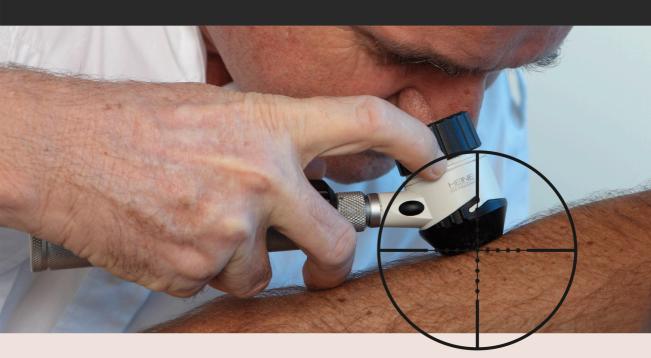
# DERMATOSCOPY AND SKIN CANCER

A HANDBOOK FOR HUNTERS
OF SKIN CANCER AND MELANOMA



**CLIFF ROSENDAHL and AKSANA MAROZAVA** 

### DERMATOSCOPY AND SKIN CANCER

## DERMATOSCOPY AND SKIN CANCER

### A HANDBOOK FOR HUNTERS OF SKIN CANCER AND MELANOMA

### CLIFF ROSENDAHL

Professor, Faculty of Medicine, The University of Queensland, Australia

### AKSANA MAROZAVA

MD, Dermatology Resident at Vitebsk State Medical University, Belarus



### © Scion Publishing Ltd, 2019

ISBN 978 1911510 338

First published 2019

All rights reserved. No part of this book may be reproduced or transmitted, in any form or by any means, without permission.

A CIP catalogue record for this book is available from the British Library.

### **Scion Publishing Limited**

The Old Hayloft, Vantage Business Park, Bloxham Road, Banbury OX16 9UX, UK www.scionpublishing.com

### **Important Note from the Publisher**

The information contained within this book was obtained by Scion Publishing Ltd from sources believed by us to be reliable. However, while every effort has been made to ensure its accuracy, no responsibility for loss or injury whatsoever occasioned to any person acting or refraining from action as a result of information contained herein can be accepted by the authors or publishers.

Readers are reminded that medicine is a constantly evolving science and while the authors and publishers have ensured that all dosages, applications and practices are based on current indications, there may be specific practices which differ between communities. You should always follow the guidelines laid down by the manufacturers of specific products and the relevant authorities in the country in which you are practising.

Although every effort has been made to ensure that all owners of copyright material have been acknowledged in this publication, we would be pleased to acknowledge in subsequent reprints or editions any omissions brought to our attention.

Registered names, trademarks, etc. used in this book, even when not marked as such, are not to be considered unprotected by law.

Cover photograph kindly provided by Brian Mallon, University of Queensland, Australia

Typeset by Evolution Design & Digital Ltd, Kent, UK Printed in the UK

Last digit is the print number: 10 9 8 7 6 5 4 3 2

### **Contents**

Prefa	face	X
Forev	eword	Xİİ
Abbr	reviations	XV
СН	IAPTER 1	
	roduction to dermatoscopy	
1.1	Why use a dermatoscope?	1
	1.1.1 The economic impact of dermatoscopy	
1.2	What is a dermatoscope?	
1.3	Colours in dermatoscopy	4
1.4	Differences between polarised and non-polarised dermatoscopy	
1.5	Uses of dermatoscopy for conditions other than tumours	11
	1.5.1 Dermatoscopy of perilesional skin	11
	1.5.2 Dermatoscopy of inflammatory conditions and dermatoses	12
	1.5.3 Dermatoscopy of hair	12
	1.5.4 Dermatoscopy of skin infestations and infections	12
	1.5.5 Dermatoscopy of nails	12
	References	13
	IAPTER 2 in – the organ	
2.1	Skin as an organ	15
2.2	Embryology of skin	
2.3	The microanatomy of skin	17
	2.3.1 Epidermis	17
	2.3.2 Melanin production	23
	2.3.3 Skin phototypes	25
	2.3.4 Melanocytic vs melanotic	25
	2.3.5 Why does melanin appear as different colours at different depths in the ski	n?26
	2.3.6 Epidermal appendages	27
	2.3.7 The basement membrane	30
	2.3.8 The dermis	31
	2.3.9 Skin blood supply	32
	2.3.10 Skin lymphatics	33
	2.3.11 Skin innervation	33
	2.3.12 Anatomy of volar skin	34

### Dermatoscopy and Skin Cancer

	2.3.13	Anatomy of the nail apparatus	34
		Anatomy of the eye in relation to melanocytic neoplasia	
		rences	
СП	APTE	D 2	
		pathology for dermatoscopists	
3.1	Fron	n the scalpel to the microscope	41
	3.1.1	Specimen processing workflow	
3.2	The	histology of normal skin	
3.3		ninology used in dermatopathology	
3.4		natoscopic histological correlation of neoplastic lesions	
		The assessment of lesions	
	3.4.2	Histology of melanoma	69
	3.4.3	Histology of basal cell carcinoma	
	3.4.4	Histology of common benign keratinocytic lesions	
	3.4.5	Histology of squamous cell carcinoma	
	Refe	rences	
	APTE		
The	lang	uage of dermatoscopy: naming and defining structures and patterns	
4.1	The	evolution of metaphoric terminology for dermatoscopic structures and patterns	91
	4.1.1	Structures in revised pattern analysis	
4.2	Revi	sed pattern analysis of lesions pigmented by melanin	92
	4.2.1	Basic structures	92
	4.2.2	Lines	92
	4.2.3	Pseudopods	101
	4.2.4	Circles	101
	4.2.5	Clods	108
	4.2.6	Dots	108
	4.2.7	Structureless	108
4.3	Patte	erns in revised pattern analysis	108
	4.3.1	Why the 2-step method of dermatoscopy is not used in revised pattern analysis	108
4.4	The	process of revised pattern analysis	110
4.5	Revi	sed pattern analysis applied to lesions with white structures	112
	4.5.1	White lines	112
	4.5.2	White clods and dots	114
	4.5.3	White structureless areas in flat lesions	114
	4.5.4	White circles in flat lesions	117
	4.5.5	White keratin clues in raised lesions	117
4.6	Revi	sed pattern analysis applied to lesions with orange, yellow and skin-coloured str	uctures
	117		
4.7	Revi	sed pattern analysis applied to vessel structures and patterns	122
	4.7.1	Vessel structures	
	4.7.2	Vessel patterns	123
	4.7.3	The anatomical correlation of vessel structures and patterns	125
4.8	The	cognition of dermatoscopy	129
	Refe	rences	131

### CHAPTER 5

### The skin examination

5.1	The skin check consultation	133
	5.1.1 Who needs a full skin examination?	133
	5.1.2 History taking	133
	5.1.3 The skin examination	134
	5.1.4 Clinical examination	134
	5.1.5 Dermatoscopic examination	135
	5.1.6 A methodical approach to complete skin examination	137
5.2	Photo-documentation	151
	5.2.1 Workflow for photo-documentation	151
5.3	Patient safety: tracking specimens and self-audit	154
5.4	The lives of lesions	155
	References	158
	APTER 6 haos and Clues': a decision algorithm for pigmented skin les	ions
6.1	'Chaos and Clues'	
6.2 6.3	Clues 164	139
0.5	6.3.1 Grey or blue structures (including grey or blue structureless)	16.4
	6.3.2 An eccentric structureless area	
	6.3.3 Thick lines reticular	
	6.3.4 Black dots and clods, peripheral	
	6.3.5 Lines radial or pseudopods, segmental	
	6.3.6 White lines	
	6.3.7 Lines parallel on the ridges (volar) or lines parallel chaotic on the na	
	6.3.8 Polymorphous vessels	
	6.3.9 Angulated lines	
6.4	Exceptions	
•	6.4.1 Any changing lesion on an adult	
	6.4.2 A nodular or small lesion which has any of the clues to malignancy	
	6.4.3 Any lesion on the head or neck with pigmented circles and/or any of	
	6.4.4 Any lesion on volar skin (palms or soles) with a parallel ridge pattern	
6.5	Excluding unequivocal seborrhoeic keratoses from biopsy	
	References	
CH.	IAPTER 7	
'Pre	ediction without Pigment': a decision algorithm for non-pig	mented skin lesions
7.1	'Prediction without Pigment'	191
	7.1.1 Prioritisation of clues in 'Prediction without Pigment'	
7.2	'Prediction without Pigment': short version	
	7.2.1 Step 1: is there ulceration?	194
	7.2.2 Step 2: are there 'white clues'?	196
	7.2.3 Step 3: is the vessel pattern consistent with malignancy?	204
7.3	Conclusion	211
	References	211

### CHAPTER 8

### Pattern analysis

3.3 Applying the aide-memoire in practice	8.1 Re	evised pattern analysis – a diagnostic algorithm	213
8.3.1 A pattern or primary pattern of lines 8.3.2 A pattern or primary pattern of pseudopods 8.3.3 A pattern or primary pattern of circles 8.3.4 A pattern or primary pattern of circles 8.3.5 A pattern or primary pattern of clods 8.3.6 A structureless pattern 8.3.6 A structureless pattern 8.3.7 A pattern or primary pattern of dots 8.3.6 A structureless pattern 8.3.8 References 8.241  CHAPTER 9  Dermatoscopic features of common and significant lesions: pigmented and non-pigmented 9.1 Melanoma: pigmented and non-pigmented 9.1 Pigmented melanoma 9.1 Pigmented melanoma 9.1 Melanomas in pigmented and non-pigmented 9.1 Melanomas on palmar and plantar skin 9.1 Melanomas on palmar and plantar skin 9.1 Melanocytic naevi: pigmented and non-pigmented 9.2 Melanocytic naevi: pigmented and non-pigmented 9.2 Pigmented acquired and congenital naevi 9.2 Pigmented acquired and congenital naevi 9.2 Pigmented acquired and congenital naevi 9.2 Pigmented acquired and scipcin pigmented naevi 9.2 Pigmented acquired and congenital naevi 9.2 Sebornhoeic keratosis-like features in congenital naevi 9.2 Pigmented naevis pigmented naevi 9.2 Pigmented naevis pigmented naevi 9.2 Pigmented naevis pigmented naevi 9.2 Pigmented naevis pigmented naevi 9.2 Pigmented naevis pigmented naevi 9.2 Pigmented naevis pigmented naevi 9.3 Basal cell carcinoma: 926 9.2 Reed and Spitz naevi 9.2 Pigmented condition pigmented 9.3 Non-pigment clues to basal cell carcinoma 9.3 Basal cell carcinoma: 927 9.3 Pigmented lues to basal cell carcinoma 9.3 Pigmented lues to basal cell carcinoma 9.3 Fibroepithelioma of Pinkus 9.4 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 9.4 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 9.4 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 9.4 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 9.4 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis	8.2 Ar	naide-memoire for revised pattern analysis of pigmented skin lesions	214
8.3.2 A pattern or primary pattern of pseudopods	8.3 A	oplying the aide-memoire in practice	215
8.33 A pattern or primary pattern of circles 8.34 A pattern or primary pattern of clods 8.35 A pattern or primary pattern of clods 8.36 A structureless pattern 8.36 A structureless pattern 8.37 References 241  CCHAPTER 9  Dermatoscopic features of common and significant lesions: pigmented and non-pigmented 9.11 Pigmented melanoma 9.11 Pigmented melanoma 9.12 Hypomelanotic and amelanotic melanoma 9.13 Metastatic melanoma 9.14 Melanomas on palmar and plantar skin 9.15 Nail apparatus melanomas 9.24 Melanocytic naevi: pigmented and non-pigmented 9.2 Melanocytic naevi: pigmented and non-pigmented 9.2 Pigmented acquired and congenital naevi 9.2 Pigmented acquired and congenital naevi 9.2 Seborrhoeic keratosis-like features in congenital naevi 9.2 Seborrhoeic keratosis-like features in congenital naevi 9.2 Red and Spitz naevu 9.2 Red and Spitz naevi 9.2 Red and Spitz naevi 9.2 Nail matrix naevi 9.2 Pigment clues to basal cell carcinoma 9.2 Pigment clues to basal cell carcinoma 9.2 Pigment clues to basal cell carcinoma 9.2 Pigment clues to basal cell carcinoma 9.3 Basal cell carcinoma: pigmented and non-pigmented 9.2 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.3 Pigment clues to basal cell carcinoma 9.4 Porokeratosis. 9.4 Seborrhoeic keratosis. pigmented and non-pigmented 9.4 Seborrhoeic keratosis. pigmented and non-pigmented 9.4 Seborrhoeic keratosis. 936 9.4 Porokeratosis. 9.4 Seborrhoeic keratosis.	8.3	.1 A pattern or primary pattern of lines	215
8.3.4 A pattern or primary pattern of clods 8.3.5 A pattern or primary pattern of dots 8.3.6 A structureless pattern 238 References 241  CHAPTER 9  Dermatoscopic features of common and significant lesions: pigmented and non-pigmented  243 9.1.1 Pigmented melanoma 9.1.2 Hypomelanotic and amelanotic melanoma 9.1.2 Hypomelanotic and amelanotic melanoma 9.1.4 Melanomas on palmar and plantar skin 9.1.5 Nail apparatus melanomas 9.1.5 Nail apparatus melanomas 9.2.1 Melanocytic naevi: pigmented and non-pigmented 9.1.2 Pigmented acquired and congenital naevi 9.1.2 Pigmented acquired and congenital naevi 9.1.3 Dysplastic naevus syndrome 9.1.4 Seborrhoeic keratosis-like features in congenital naevi 9.2.5 Non-pigmented naevi 9.2.6 Blue naevus 9.2.7 Halo naevus phenomenon 9.2.8 Growing naevi 9.2.9 Reed and Spitz naevi. 9.2.1 Nail matrix naevi. 9.2.2 Pigmented clues to basal cell carcinoma 9.2.3 Basal cell carcinoma: pigmented and non-pigmented 9.2.6 9.2.1 Naevi on volar skin 9.2.7 9.3.1 Non-pigment clues to basal cell carcinoma 9.3.3 Basal cell carcinoma: pigmented and non-pigmented 9.3.3 Pigmentelioma of Pinkus 9.3.4 Seborrhoeic keratosis: pigmented and non-pigmented 9.3.5 Sibroco keratosis: pigmented and non-pigmented 9.3.6 Benign keratinocytic lesions 9.3.1 Non-pigment clues to basal cell carcinoma 9.3.2 Pigment clues to basal cell carcinoma 9.3.3 Fibroepithelioma of Pinkus 9.3.4 Seborrhoeic keratosis: pigmented and non-pigmented 9.4.5 Succo keratosis: 9.30 9.4.4 Porokeratosis: 9.30 9.4.5 Stucco keratosis: 300 9.4.5 Stucco keratosis: 300	8.3	.2 A pattern or primary pattern of pseudopods	228
8.3.5 A pattern or primary pattern of dots 8.3.6 A structureless pattern	8.3	.3 A pattern or primary pattern of circles	229
References	8.3	.4 A pattern or primary pattern of clods	233
References	8.3	5 A pattern or primary pattern of dots	236
Dermatoscopic features of common and significant lesions: pigmented and non-pigmented  2.1 Melanoma: pigmented and non-pigmented 2.43 2.1. Pigmented melanoma 2.44 2.1. Hypomelanotic and amelanotic melanoma 2.44 2.1. Melanomas on palmar and plantar skin 2.44 2.1. Nail apparatus melanomas. 2.44 2.1. Nail apparatus melanomas. 2.44 2.2. Melanocytic naevi: pigmented and non-pigmented 2.53 2.1 Basic classification of naevi. 2.54 2.2. Pigmented acquired and congenital naevi. 2.54 2.2. Pigmented acquired and congenital naevi. 2.54 2.2. Seborrhoeic keratosis-like features in congenital naevi 2.58 2.2. Sun-pigmented naevi. 2.58 2.2. Blue naevus 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Non-pigmented naevi. 2.59 2.2. Naevi on volar skin 2.59 2.2. Naev	8.3	.6 A structureless pattern	238
Dermatoscopic features of common and significant lesions: pigmented and non-pigmented  2.1 Melanoma: pigmented and non-pigmented 243 2.1. Pigmented melanoma 244 2.1. Pigmented melanoma 244 2.1. Hypomelanotic and amelanotic melanoma 244 2.1. Melanomas on palmar and plantar skin 244 2.1. Nail apparatus melanomas 244 2.2 Melanocytic naevi: pigmented and non-pigmented 253 2.1 Basic classification of naevi 254 2.2. Pigmented acquired and congenital naevi 254 2.2. Pigmented acquired and congenital naevi 258 2.3 Dysplastic naevus syndrome 258 2.4 Seborrhoeic keratosis-like features in congenital naevi 258 2.5 Non-pigmented naevi 266 2.6 Blue naevus 266 2.7 Halo naevus phenomenon 266 2.8 Growing naevi 269 2.9 Reed and Spitz naevi 269 2.10 Naevi on volar skin 274 2.11 Nail matrix naevi 276 2.12 Nail matrix naevi 276 2.13 Basal cell carcinoma: pigmented and non-pigmented 276 2.14 Seborrhoeic keratosis -like features in congenital naevi 276 28. Pigment clues to basal cell carcinoma 276 28. Pigment clues to basal cell carcinoma 276 28. Pigment clues to basal cell carcinoma 276 28. Pigment clues to basal cell carcinoma 277 28. Pigment clues to basal cell carcinoma 276 28. Pigment clues to basal cell carcinoma 277 28. Seborrhoeic keratosis: pigmented and non-pigmented 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 29.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 280	Re	eferences	241
Melanoma: pigmented 243 9.1.1 Pigmented melanoma 244 9.1.2 Hypomelanotic and amelanotic melanoma 244 9.1.3 Metastatic melanoma 244 9.1.4 Melanomas on palmar and plantar skin 244 9.1.5 Nail apparatus melanomas 244 9.1.6 Melanocytic naevi: pigmented and non-pigmented 253 9.2.1 Basic classification of naevi. 254 9.2.2 Pigmented acquired and congenital naevi. 254 9.2.3 Dysplastic naevus syndrome 258 9.2.4 Seborrhoeic keratosis-like features in congenital naevi 258 9.2.5 Non-pigmented naevi. 266 9.2.6 Blue naevus . 266 9.2.7 Halo naevus phenomenon 266 9.2.8 Growing naevi 269 9.2.9 Reed and Spitz naevi. 269 9.2.10 Naevi on volar skin 274 9.2.11 Nail matrix naevi. 276 9.2.12 Nail matrix naevi. 276 9.2.13 In Non-pigment clues to basal cell carcinoma 276 9.3.1 Non-pigment clues to basal cell carcinoma 276 9.3.2 Pigment clues to basal cell carcinoma 277 9.3.3 Fibroepithelioma of Pinkus 288 9.4.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 9.4.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis 289 9.4.2 Seborrhoeic keratosis: pigmented and non-pigmented 288 9.4.3 Lichen planus-like keratosis 303 9.4.4 Porokeratosis 303 9.4.5 Stucco keratosis 303	CHAP	TER 9	
9.1 Melanoma: pigmented and non-pigmented 9.1.1 Pigmented melanoma			
91.1       Pigmented melanoma       244         91.2       Hypomelanotic and amelanotic melanoma       244         91.3       Metastatic melanoma       244         91.4       Melanomas on palmar and plantar skin       244         91.5       Nail apparatus melanomas       244         9.2       Melanocytic naevi: pigmented and non-pigmented       253         92.1       Basic classification of naevi       254         92.2       Pigmented acquired and congenital naevi       254         92.2       Pigmented acquired and congenital naevi       258         92.2       Seborrhoeic keratosis-like features in congenital naevi       258         92.5       Non-pigmented naevi       258         92.5       Non-pigmented naevi       269         92.6       Blue naevus       264         92.7       Halo naevus phenomenon       269         92.8       Growing naevi       269         92.9       Reed and Spitz naevi       269         92.9       Reed and Spitz naevi       269         92.1       Nail matrix naevi       276         93.1       Non-pigment clues to basal cell carcinoma       276         93.1       Non-pigmented uses to basal cell carcinoma       276	- '		
91.2       Hypomelanotic and amelanotic melanoma       244         91.3       Metastatic melanoma       244         91.4       Melanomas on palmar and plantar skin       244         91.5       Nail apparatus melanomas       244         9.2       Melanocytic naevi: pigmented and non-pigmented       253         9.2.1       Basic classification of naevi       254         9.2.2       Pigmented acquired and congenital naevi       254         9.2.3       Dysplastic naevus syndrome       258         9.2.4       Seborrhoeic keratosis-like features in congenital naevi       258         9.2.5       Non-pigmented naevi       261         9.2.6       Blue naevus       264         9.2.7       Halo naevus phenomenon       269         9.2.8       Growing naevi       269         9.2.9       Reed and Spitz naevi       269         9.2.1       Naevi on volar skin       274         9.2.1       Nail matrix naevi       276         9.3       Basal cell carcinoma: pigmented and non-pigmented       276         9.3.1       Non-pigment clues to basal cell carcinoma       277         9.3.2       Pigment clues to basal cell carcinoma       277         9.3.3       Fibroepithelioma o			
9.1.3 Metastatic melanoma		9	
9.1.4 Melanomas on palmar and plantar skin			
9.1.5 Nail apparatus melanomas			
8.2       Melanocytic naevi: pigmented and non-pigmented.       253         9.2.1       Basic classification of naevi.       254         9.2.2       Pigmented acquired and congenital naevi.       254         9.2.3       Dysplastic naevus syndrome.       258         9.2.4       Seborrhoeic keratosis-like features in congenital naevi.       258         9.2.5       Non-pigmented naevi.       261         9.2.6       Blue naevus.       264         9.2.7       Halo naevus phenomenon.       269         9.2.8       Growing naevi.       269         9.2.9       Reed and Spitz naevi.       269         9.2.10       Naevi on volar skin.       274         9.2.11       Nail matrix naevi.       276         9.3.1       Non-pigment clues to basal cell carcinoma.       276         9.3.2       Pigment clues to basal cell carcinoma.       276         9.3.2       Pigment clues to basal cell carcinoma.       277         9.3.3       Fibroepithelioma of Pinkus.       284         9.4       Benign keratinocytic lesions.       289         9.4.1       Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis.       289         9.4.2       Seborrhoeic keratosis: pigmented and non-pigmented.		·	
9.2.1 Basic classification of naevi		· ·	
9.2.2 Pigmented acquired and congenital naevi		, , , , , , , , , , , , , , , , , , , ,	
92.3 Dysplastic naevus syndrome			
9.24 Seborrhoeic keratosis-like features in congenital naevi			
9.2.5       Non-pigmented naevi			
9.2.6 Blue naevus		g .	
9.27 Halo naevus phenomenon		, 9	
9.2.8       Growing naevi	9.2		
9.2.9 Reed and Spitz naevi	9.2	·	
9.2.10 Naevi on volar skin	9.2		
9.2.11 Nail matrix naevi	9.2	.9 Reed and Spitz naevi	269
9.3Basal cell carcinoma: pigmented and non-pigmented2769.3.1Non-pigment clues to basal cell carcinoma2769.3.2Pigment clues to basal cell carcinoma2779.3.3Fibroepithelioma of Pinkus2849.4Benign keratinocytic lesions2899.4.1Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis2899.4.2Seborrhoeic keratosis: pigmented and non-pigmented2899.4.3Lichen planus-like keratosis3039.44Porokeratosis3069.4.5Stucco keratosis306			
9.3.1 Non-pigment clues to basal cell carcinoma			
9.3.2 Pigment clues to basal cell carcinoma	9.3 Ba	ısal cell carcinoma: pigmented and non-pigmented	276
9.3.3 Fibroepithelioma of Pinkus	9.3	.1 Non-pigment clues to basal cell carcinoma	276
9.4 Benign keratinocytic lesions	9.3	.2 Pigment clues to basal cell carcinoma	277
9.4.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis	9.3	.3 Fibroepithelioma of Pinkus	284
9.4.2Seborrhoeic keratosis: pigmented and non-pigmented.2899.4.3Lichen planus-like keratosis.3039.4.4Porokeratosis.3069.4.5Stucco keratosis.306	9.4 Be	nign keratinocytic lesions	289
9.4.3       Lichen planus-like keratosis       .303         9.4.4       Porokeratosis       .306         9.4.5       Stucco keratosis       .306	9.4	.1 Solar lentigo, ink spot lentigo, melanotic macule and idiopathic guttate hypomelanosis	289
9.4.4 Porokeratosis	9.4	.2 Seborrhoeic keratosis: pigmented and non-pigmented	289
9.4.5 Stucco keratosis	9.4	·	
	9.4	4 Porokeratosis	306
	9.4	.5 Stucco keratosis	306
9.4.6 Viral wart	9.4	.6 Viral wart	306
9.4.7 Clear cell acanthoma306	9.4	.7 Clear cell acanthoma	306
Actinic keratosis, squamous cell carcinoma in situ and squamous cell carcinoma	9.5 Ac	tinic keratosis, squamous cell carcinoma in situ and squamous cell carcinoma	310

### Dermatoscopy and Skin Cancer

	9.5.1	Actinic keratosis	310
	9.5.2	Squamous cell carcinoma in situ	313
	9.5.3	Distinguishing flat pigmented facial lesions: solar lentigo, pAK/SCC in situ and LM	320
	9.5.4	Squamous cell carcinoma and keratoacanthoma	331
9.6	Dern	natofibroma and dermatofibrosarcoma protuberans	338
9.7	Haer	mangioma and other vascular lesions	342
9.8	Merk	cel cell carcinoma	346
9.9	Atyp	ical fibroxanthoma	346
9.10	Adne	exal tumours	348
	9.10.1	Sebaceous gland hyperplasia	348
	9.10.2	Trichoepithelioma	348
	9.10.3	Trichilemmoma	348
	9.10.4	Cysts – trichilemmal (pilar) cyst/wen and epidermal cyst	348
	9.10.5	Pilomatrixoma	350
	9.10.6	Eccrine poroma	350
	9.10.7	Sebaceous adenoma and carcinoma	350
9.11	Neur	rofibroma	353
9.12	Moll	uscum contagiosum	353
9.13	Cuta	neous lymphoma	353
9.14	Kapo	osi sarcoma	355
	Refe	rences	356
Inde	X		359

### **Preface**

Fifty years ago when I entered my first year of medical studies in 1969, the same year that Neil Armstrong stepped onto the moon, dermatoscopy as we know it was science fiction. Much has changed. Dermatoscopy is now standard of care in the management of skin cancer and melanoma

My interest in this novel science became focused after a family member, Graham, developed metastatic melanoma. Graham did not blame the GP who dismissed a lesion of concern on his thigh a couple of years earlier, and he made the point to me that GPs were not prepared for this challenge in their training, a challenge that was thrust on them due to a rising incidence of melanoma and an inexplicable shortage of dermatologists in Australia. Graham's GP looked after him well. Right up to the moment of his death, a death which was predictably terrible, aggravated by multi-organ metastases and finally necrotising fasciitis.

My journey since then, commencing with a PhD expertly supervised by David Wilkinson and Peter Soyer and focused on improving skin cancer management in Australia, has been a very steep learning curve. I have been mentored by men of undoubted genius: Harald Kittler and David Weedon, men whose genius was only matched by their generosity. I have been assisted by exceptional colleagues: lan McColl, Iris Zalaudek, Alan Cameron, Jeff Keir, Greg Canning, Phil Tschandl, Agata Bulinska, Simon Clark and Nisa Akay. I am particularly grateful to Harald

Kittler, Stephen Hayes and Jeff Keir for their critical review of the book and to Simon Clark for reviewing and correcting the dermatopathology chapter.

This book would never have been possible without my co-author Aksana Marozava. Aksana worked with me for two years, taught me how to do a skin examination and dispelled any delusions of grandeur by repeatedly discovering significant lesions I had passed over. Her diligence and skill in collating my image collection for the book and preparing all of the graphics has hopefully made this book the masterpiece we wanted to produce.

The hunting metaphor is no accident. Hunting and gathering (Aksana insists that she is a gatherer) are as natural to *Homo sapiens* as is falling in love. The romance and thrill of the hunt elevates what we do to more than the drudgery of repetitive work, and the satisfaction of every success motivates further effort.

Finally, I am indebted to my wife Debbie for putting up with me through this journey and for effectively managing our practice and business affairs so I could focus on hunting, research, teaching and writing.

To conclude, I quote Vice Admiral Horatio Nelson, hunter extraordinaire, speaking at the battle of Copenhagen in 1801:

"It is warm work; and this day may be the last to any of us at a moment. But mark you! I would not be elsewhere for thousands"

> Cliff Rosendahl Brisbane March 2019

### **Foreword**

This new book is an important step forward in the developing art and science of dermatoscopy for skin lesion recognition. The debate as to whether the technique is any good is surely over, but more help as to how to best do it. and (vitally) to best teach it, is most welcome.

Over the last decade or so. Cliff Rosendahl. and more recently Aksana Marozava, have documented some 19.000 excised skin lesions in Cliff's clinic in Capalaba, Brisbane, and fed the data into the SCARD online database which he set up with Tobias Wilson. This book summarises the knowledge gained from the analysis of that histopathological data and the lesion images, plain and dermatoscopic. The sheer scale of the data behind this book gives it an authority that can't be ignored.

The book is built around two algorithms, 'Chaos and Clues' and 'Prediction without Pigment' which, as explained, may not always lead to a diagnosis, but to a safe decision as to whether excision is required. The selected colour images illustrate well the dermatoscopic features and terms set out in the text.

Cliff is fully committed to revised pattern analysis and the use of what he calls objective geometric terminology to describe dermatoscopic structures, building on the 'descriptive' terminology often associated with co-worker Harald Kittler of Vienna. There are no 'arborising' vessels here (if vessels are 'tree like', then what sort of tree?) but branched serpentine (admittedly, 'serpentine', i.e. snakelike, is still a metaphor, but a much more consistent one than tree-like) And it is further explained that the apparent sharp focus of such vessels in BCC is due to the superficial

cutaneous vascular plexus being clarified by the translucent BCC stroma, rather than that vessel morphology being unique to BCC.

There is more basic science here than is usual in a book aimed at beginners, but the extra effort put into appreciating the embryology, anatomy and histopathology pays rewards, particularly with regard to dermatoscopic-pathological correlation. Recognising structures like blue clods and polarising-specific white lines is good, but understanding what they mean at the microanatomical level gives insight into the modus operandi of the target of the hunt: malignant tissue.

More recently described signs such as white circles in early invasive SCC and angulated lines and polygons in melanoma in situ are detailed. I have witnessed Cliff working in his clinic and I can say that the author has a zero tolerance approach to such lesions. with approximately 80% of the melanomas diagnosed in his clinic being pre-invasive.

Dermatoscopy and Skin Cancer is a more challenging read than some earlier textbooks on this subject, but builds on hard-won, audit-backed knowledge to take us to the next level of advanced pattern analysis. It can be commended to the beginner/improver and indeed expert, who is willing to put in some work to embrace the latest evidence-based approach and terminology, which seems likely to supersede the earlier algorithms based on metaphorical language. This may mean some effort for those of us who learned dermoscopy/dermatoscopy with terms like maple leaf, arborising, comedo-like, ovoid nests, etc., but the new approach makes sense

if for no other reasons than the need for translation and utility for international research, for dermatoscopy is now highly globalised.

### **Dr Stephen Hayes**

Independent dermatoscopy educator Associate Specialist in Dermatology, University Hospital Southampton UK board member, International Dermoscopy Society

### **Abbreviations**

actinic keratosis ΑK BCCbasal cell carcinoma dermatofibroma DF

DOPA dihydroxyphenylalanine

FFG elevated, firm and continuously growing

haematoxylin and eosin H&E KΑ keratoacanthoma

LN lymph nodes

LPLK lichen planus-like keratosis

MHC major histocompatibility complex

melanocytic naevus Naevus

NMSC non-melanoma skin cancer PAM primary acquired melanosis pBCC pigmented basal cell carcinoma pIEC pigmented intraepidermal carcinoma pSCC pigmented squamous cell carcinoma

RPA revised pattern analysis RPE retinal pigmented epithelial

relative risk RR

SCARD skin cancer audit research database

SCC squamous cell carcinoma

UV ultraviolet

### CHAPTER 1

### Introduction to dermatoscopy



### 1.1 Why use a dermatoscope?

"Melanoma writes its message on the skin with its own ink and it is there for all to see. Unfortunately, some see but do not comprehend."

> Neville Davis; Modern concepts of melanoma and its management; Annals of Plastic Surgery, 1978

Since Neville Davis made this statement the advent of dermatoscopy has facilitated earlier diagnosis of melanoma, as well as enhancing diagnostic accuracy for many dermatological conditions both benign and malignant<sup>1</sup>. The handheld dermatoscope is a recent innovation, having first become available in 1987, but there has been a rapid proliferation of research commensurate with the utility and efficacy of this relatively inexpensive instrument. Studies have demonstrated that dermatoscopy improves diagnostic accuracy for pigmented skin malignancies, both melanocytic<sup>2</sup> (naevus and melanoma) and non-melanocytic<sup>3</sup> (basal cell carcinoma (BCC) and squamous cell carcinoma (SCC)), to the extent that it is now standard of care in Australasia for any clinician treating pigmented skin lesions4. It has now also been shown that dermatoscopy improves diagnostic accuracy for non-pigmented skin lesions<sup>5</sup> and it is also being increasingly applied to the diagnosis of many inflammatory skin conditions. Not only is dermatoscopy useful for the diagnosis of skin conditions, but it has also been shown to be effective for application in skin cancer surgery, where surgical margins are significantly more likely to be adequate when dermatoscopy is utilised at preoperative

marking<sup>6</sup>. The dermatoscope is an essential tool for dermatologists and, with skin conditions accounting for up to 14.8% of all consultations in general practice<sup>7</sup>, there is a compelling argument that it is as applicable for general practitioners (GPs) as the stethoscope (Figure 1.1)8.

### 1.1.1 The economic impact of dermatoscopy

An American study in 2016 found that the economic costs avoided by diagnosing melanoma 6 months earlier justified over 100 (170 when loss of earnings was considered) benign biopsies9. Anything that achieves the same rate of diagnosis and therefore prevention of delayed diagnosis, with greater specificity, will achieve these savings with a smaller investment

An Australian primary care-based study found that generalist GPs performed 17 benign biopsies for each melanoma treated, whereas GPs subspecialised in skin cancer practice, who also had a high use of dermatoscopy, discovered one melanoma for every 8.5 biopsies<sup>10</sup>. This suggests that with respect to the management of melanoma alone, derma-

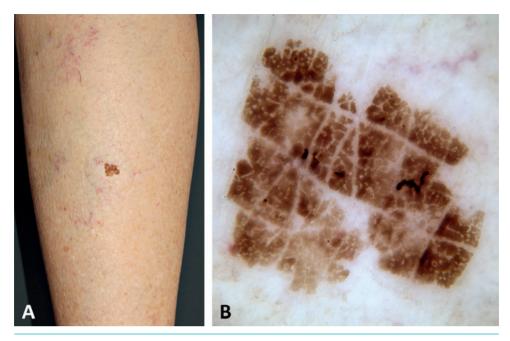


Figure 1.1: Clinically (A) this irregular pigmented lesion is suspicious for malignancy, but dermatoscopically (B) it has the unequivocal morphology of a seborrhoeic keratosis (see Figure 9.80).

toscopy could result in economic savings by earlier detection with fewer unnecessary excisions.

It is not unusual in current dermatology and primary care practice for a biopsy to precede many therapeutic surgical procedures for both pigmented and non-pigmented lesions. While suspected melanomas should have an initial excisional biopsy, we have found that with the advent of dermatoscopy and increased diagnostic confidence, a preliminary partial biopsy procedure is not necessary for the majority of suspected non-melanoma skin cancers (NMSC). Unpublished raw data from the skin cancer audit research database (SCARD) gives a snapshot of current practice in Australasia of 848 primary care doctors, either practising in dedicated skin cancer practices or in general practices with a special

interest in skin cancer. Of 429,010 specimens of NMSC treated surgically, 316,339 (73.73%) were managed in a 1-step approach, without a preceding biopsy<sup>11</sup>. This suggests that a proportion of primary care doctors are already managing NMSC through a 1-step process in the majority of cases.

There are many advantages of proceeding directly to curative surgical management followina а confident dermatoscopic diagnosis. These include the fact that margins are more likely to be more clearly definable without the inflammation that is expected after a partial biopsy. A single procedure is more convenient for the patient and, when the costs of one rather than two surgical episodes and dermatopathological assessments are considered, the economic saving is approximately 50%.

### What is a dermatoscope?

A dermatoscope is essentially a magnifying glass which eliminates surface reflection from the skin by using either a fluid interface (non-polarising contact dermatoscopes) or polarising filters (polarising contact or non-contact dermatoscopes) (Figure 1.2). This allows pigmented structures to be seen to the level of the deep dermis, up to 1mm into the skin, as well as blood vessels in the dermis when they are not obscured by pigment. A built-in light source allows the device to be used as a compact handheld instrument. Even the early dermatoscopes provided adequate visual information but initial studies were burdened by the need for expensive film photography. The advent of new dermatoscopes with even brighter optics, as well as with the option of polarised light sources, was paralleled by the availability of digital photography. This has facilitated the convenient forwarding of captured images for purposes including research and teledermatoscopy.

Although polarising dermatoscopes can be used without interface fluid, visualisation of structures can be improved with fluid and all the images displayed in this book are taken with fluid immersion, whether the dermatoscope was in polarised mode or not. Initially the interface fluid of choice was oil but that is rarely used now. Alcohol-based fluids (70% ethanol in water, isopropanol in water or alcohol hand gel) are just as effective and have the advantage of having an antiseptic effect as well as of drying very quickly. Ultrasound gel is useful with thicker lesions such as keratoacanthoma (KA) or when examining complex curved surfaces such as the nail unit. Even the use of a sterile alcohol wipe can be effective. Whatever fluid is used should be wiped from the dermatoscope footplate after use for hygienic purposes as well as to protect the footplate from a build-up of residue.

It has been claimed that polarised dermatoscopy provides a superior view of vessel structures – we have found that to be related to footplate pressure rather than to polarisation. Of course, when polarised dermatoscopes are used in non-contact mode vessels will not be compressed. The same can be achieved in contact mode with non-polarised or polarised dermatoscopy if less footplate pressure is applied to the lesion. Sometimes the use of thicker contact fluid such as hand gel or ultrasound gel may facilitate this.





Figure 1.2: Dermatoscopes are available from a variety of companies. (A) The DermLite DL4 (3Gen) and (B) the Heine delta 20T (Heine Corporation) both default to polarised mode but can easily be switched to non-polarised mode.

### Melanin

The main pigment influencing the colour of fair skin is haem. In people with darker skin