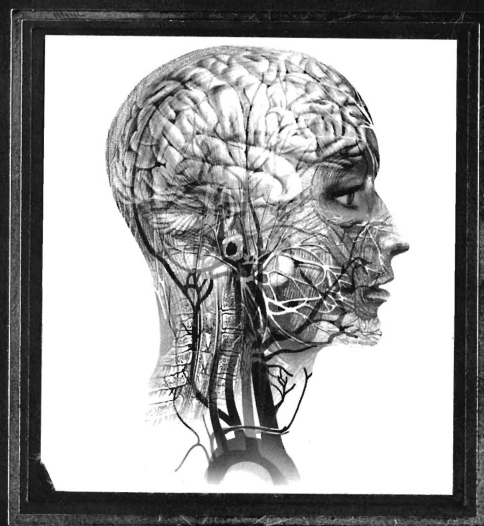


SEVENTH EDITION

Scott-Brown's
Otorhinolaryngology,
Head and Neck Surgery

VOLUME 1



Edited by

Michael Gleason

George G Browning, Martin J Burton, Roy Clarke, John Elliott,
Nicholas S Jones, Valerie J Lund, Linda M Luxon, John C Watkinson

Scott-Brown's Otorhinolaryngology, Head and Neck Surgery

Scott-Brown's Otorhinolaryngology, Head and Neck Surgery

7th edition

Lead editor: Michael Gleeson

Volume 1

- Part 1 Cell biology, edited by Nicholas S Jones
- Part 2 Wound healing, edited by Nicholas S Jones
- Part 3 Immunology, edited by Nicholas S Jones
- Part 4 Microbiology, edited by Nicholas S Jones
- Part 5 Haematology, edited by Nicholas S Jones
- Part 6 Endocrinology, edited by Nicholas S Jones
- Part 7 Pharmacotherapeutics, edited by Martin J Burton
- Part 8 Perioperative management, edited by Martin J Burton
- Part 9 Safe and effective practice, edited by Martin J Burton
- Part 10 Interpretation and management of data, edited by Martin J Burton
- Part 11 Recent advances in technology, edited by Martin J Burton
- Part 12 Paediatric otorhinolaryngology, edited by Ray Clarke

Volume 2

- Part 13 The nose and paranasal sinuses, edited by Valerie J Lund
- Part 14 The neck, edited by John Hibbert
- Part 15 The upper digestive tract, edited by John Hibbert
- Part 16 The upper airway, edited by John Hibbert
- Part 17 Head and neck tumours, edited by John Hibbert

Volume 3

- Part 18 Plastic surgery of the head and neck, edited by John C Watkinson
- Part 19 The ear, hearing and balance, edited by George G Browning and Linda M Luxon
- Part 20 Skull base, edited by Michael Gleeson
- Index
- CD-ROM

George G Browning MD FRCS

Professor of Otorhinolaryngology, MRC Institute of Hearing Research, Glasgow Royal Infirmary, Glasgow, UK

Martin J Burton MA DM FRCS

Senior Clinical Lecturer, University of Oxford; and Consultant Otolaryngologist, Oxford Radcliffe NHS Trust Oxford, UK

Ray Clarke BSc DCH FRCS FRCS (ORL)

Consultant Paediatric Otolaryngologist, Royal Liverpool University Children's Hospital, Alder Hey, Liverpool, UK

Michael Gleeson MD FRCS

Professor of Otolaryngology and Skull Base Surgery, Institute of Neurology, University College London; and Consultant, Guy's, Kings and St Thomas' and the National Hospital for Neurology and Neurosurgery, London UK; and Honorary Consultant Skull Base Surgeon, Great Ormond Street Hospital for Sick Children, London, UK

John Hibbert ChM FRCS

Formerly Consultant Otolaryngologist, Department of Otolaryngology, Guy's Hospital, London, UK

Nicholas S Jones MD FRCS FRCS (ORL)

Professor of Otorhinolaryngology, Queen's Medical Centre, University of Nottingham, Nottingham UK

Valerie J Lund MS FRCS FRCS (Ed)

Professor of Rhinology, The Ear Institute, University College London, London, UK

Linda M Luxon BSc MBBS FRCP

Professor of Audiovestibular Medicine, University of London at University College London, Academic Unit of Audiovestibular Medicine; and Consultant Physician, National Hospital for Neurology and Neurosurgery; and Honorary Consultant Physician, Great Ormond Street Hospital for Children, London, UK

John C Watkinson MSc MS FRCS (Ed, Glas, Land) DLO

Consultant Head and Neck and Thyroid Surgeon, Department of Otorhinolaryngology/Head and Neck Surgery, Queen Elizabeth Hospital, University of Birmingham NHS Trust, Birmingham, UK

Scott-Brown's Otorhinolaryngology, Head and Neck Surgery

7th edition

Volume 1

Edited by

Michael Gleeson

George G Browning, Martin J Burton,

Ray Clarke, John Hibbert, Nicholas S Jones,

Valerie J Lund, Linda M Luxon,

John C Watkinson

Hodder Arnold

www.hoddereducation.com

First published in Great Britain in 1952 by Butterworth & Co.

Second edition 1965

Third edition 1971

Fourth edition 1979

Fifth edition 1987

Sixth edition 1997

This seventh edition published in Great Britain in 2008 by Hodder Arnold

An imprint of Hodder Education, a part of Hachette Livre UK, 338 Euston Road, London NW1 3BH

<http://www.hoddereducation.com>

© 2008 Edward Arnold (Publishers) Ltd

All rights reserved. Apart from any use permitted under UK copyright law, this publication may only be reproduced, stored or transmitted, in any form, or by any means with prior permission in writing of the publishers or in the case of reprographic production in accordance with the terms of licences issued by the Copyright Licensing Agency. In the United Kingdom such licences are issued by the Copyright Licensing Agency: Saffron House, 6-10 Kirby Street, London EC1N 8TS

Whilst the advice and information in this book are believed to be true and accurate at the date of going to press, neither the author[s] nor the publisher can accept any legal responsibility or liability for any errors or omissions that may be made. In particular (but without limiting the generality of the preceding disclaimer) every effort has been made to check drug dosages; however it is still possible that errors have been missed. Furthermore, dosage schedules are constantly being revised and new side-effects recognized. For these reasons the reader is strongly urged to consult the drug companies' printed instructions before administering any of the drugs recommended in this book.

British Library Cataloguing in Publication Data

A catalogue record for this book is available from the British Library

Library of Congress Cataloging-in-Publication Data

A catalog record for this book is available from the Library of Congress

ISBN 978 0 340 808 931

1 2 3 4 5 6 7 8 9 10

Commissioning Editor: Joanna Koster

Project Editor: Zelah Pengilley

Production Controller: Lindsay Smith / Andre Sim

Text and Cover Designer: Amina Dudhia

Cover photograph © MEHAU KULYK/SCIENCE PHOTO LIBRARY

Typeset in 10 pt Minion by Macmillan India

Printed and bound in India.

What do you think about this book? Or any other Hodder Arnold title?
Please send your comments to www.hoddereducation.com

Contents

| | |
|----------------------|--------|
| Contributors | xi |
| Preface | xxix |
| How to use this book | xxxix |
| Abbreviations | xxxiii |

PART 1 CELL BIOLOGY – EDITED BY NICHOLAS S JONES **1**

| | | |
|---|--|----|
| 1 | Molecular biology <i>Michael Kuo and Richard Irving</i> | 3 |
| 2 | Genetics <i>Karen P Steel</i> | 15 |
| 3 | Gene therapy <i>Scott M Graham and John H Lee</i> | 23 |
| 4 | Mechanisms of anticancer drugs <i>Sarah Payne and David Miles</i> | 34 |
| 5 | Radiotherapy and radiosensitizers <i>Stewart G Martin and David AL Morgan</i> | 47 |
| 6 | Apoptosis and cell death <i>Michael Saunders</i> | 56 |
| 7 | Stem cells <i>A John Harris and Archana Vats</i> | 66 |

PART 2 WOUND HEALING – EDITED BY NICHOLAS S JONES **85**

| | | |
|----|--|-----|
| 8 | Soft and hard tissue repair <i>Stephen R Young and Melissa Calvin</i> | 87 |
| 9 | Skin flap physiology <i>A Graeme B Perks</i> | 110 |
| 10 | Biomaterials, tissue engineering and their applications <i>Colin A Scotchford, Matthew Evans and Archana Vats</i> | 118 |

PART 3 IMMUNOLOGY – EDITED BY NICHOLAS S JONES **131**

| | | |
|----|--|-----|
| 11 | Defence mechanisms <i>Ian Todd and Richard J Powell</i> | 133 |
| 12 | Allergy: basic mechanisms and tests <i>Stephen R Durham and Graham Banfield</i> | 144 |

| | | |
|--|--|------------|
| 13 | Evaluation of the immune system <i>Elizabeth Drewe and Richard J Powell</i> | 167 |
| 14 | Primary immunodeficiencies <i>Elizabeth Drewe and Richard J Powell</i> | 174 |
| 15 | Rheumatological diseases <i>Adrian Drake-Lee</i> | 183 |
| PART 4 MICROBIOLOGY – EDITED BY NICHOLAS S JONES | | 193 |
| 16 | Microorganisms <i>Vivienne Weston</i> | 195 |
| 17 | Viruses and antiviral agents <i>Paul Simons and Karl G Nicholson</i> | 204 |
| 18 | Fungi <i>Juliette Morgan and David W Warnock</i> | 213 |
| 19 | Antimicrobial therapy <i>Vivienne Weston</i> | 228 |
| 20 | HIV and otolaryngology <i>Thomas A Tami and Jahmal A Hairston</i> | 238 |
| PART 5 HAEMATOLOGY – EDITED BY NICHOLAS S JONES | | 251 |
| 21 | Blood groups, blood components and alternatives to transfusion <i>Fiona Regan and Ian Gabriel</i> | 253 |
| 22 | Haemato-oncology <i>Clare Wykes and Fiona Regan</i> | 265 |
| 23 | Haemostasis: normal physiology, disorders of haemostasis and thrombosis and their management <i>Fiona Regan</i> | 278 |
| PART 6 ENDOCRINOLOGY – EDITED BY NICHOLAS S JONES | | 293 |
| 24 | The pituitary gland: anatomy and physiology <i>John Hill</i> | 295 |
| 25 | The pituitary: imaging and tests of function <i>Alan P Johnson, Swarupsinh Chavda and Paul Stewart</i> | 303 |
| 26 | The thyroid gland: anatomy and physiology <i>Julian A McGlashan</i> | 314 |
| 27 | The thyroid gland: function tests and imaging <i>Susan Clarke</i> | 327 |
| 28 | The thyroid: nonmalignant disease <i>Lorraine M Albon and Jayne A Franklyn</i> | 338 |
| 29 | The parathyroid glands: anatomy and physiology <i>Mateen H Arastu and William J Owen†</i> | 367 |
| 30 | Parathyroid function tests and imaging <i>David Hosking</i> | 379 |
| 31 | Parathyroid dysfunction: medical and surgical therapy <i>E Dinakara Babu, Bill Fleming and JA Lynn</i> | 387 |
| 32 | Head and neck manifestations of endocrine disease <i>Jonathan M Morgan and Thomas McCaffrey</i> | 398 |

| | |
|---|------------|
| PART 7 PHARMACOTHERAPEUTICS – EDITED BY MARTIN J BURTON | 405 |
| 33 Drug administration and monitoring <i>Geraldine Gallagher</i> | 407 |
| 34 Corticosteroids in otolaryngology <i>Niels Mygind and Jens Thomsen</i> | 418 |
| 35 Drug therapy in otology <i>Wendy Smith and Martin Burton</i> | 429 |
| 36 Drug therapy in rhinology <i>Wendy Smith and Grant Bates</i> | 436 |
| 37 Drug therapy in laryngology and head and neck surgery <i>Wendy Smith and Rogan Corbridge</i> | 446 |
| | |
| PART 8 PERIOPERATIVE MANAGEMENT – EDITED BY MARTIN J BURTON | 455 |
| 38 Preparation of the patient for surgery <i>Adrian Pearce</i> | 457 |
| 39 Recognition and management of the difficult airway <i>Adrian Pearce</i> | 467 |
| 40 Adult anaesthesia <i>Andrew D Farmery and Jaideep J Pandit</i> | 488 |
| 41 Paediatric anaesthesia <i>Alistair Cranston</i> | 507 |
| 42 Adult critical care <i>Gavin G Lavery</i> | 526 |
| 43 Paediatric intensive care <i>Helen Allen and Rob Ross Russell</i> | 542 |
| | |
| PART 9 SAFE AND EFFECTIVE PRACTICE – EDITED BY MARTIN J BURTON | 549 |
| 44 Training, accreditation and the maintenance of skills <i>Paul O’Flynn</i> | 551 |
| 45 Communication and the medical consultation <i>Damian Gardner-Thorpe and Richard Canter</i> | 559 |
| 46 Clinical governance: Improving the quality of patient care <i>Debbie Wall, Patrick J Bradley and Aidan Halligan</i> | 568 |
| 47 Medical ethics <i>Katherine Wasson</i> | 581 |
| 48 Medical jurisprudence and otorhinolaryngology <i>Maurice Hawthorne and Desmond Watson</i> | 594 |
| | |
| PART 10 INTERPRETATION AND MANAGEMENT OF DATA – EDITED BY MARTIN J BURTON | 613 |
| 49 Epidemiology <i>Jan HP van der Meulen and David A Lowe</i> | 615 |
| 50 Outcomes research <i>Iain RC Swan</i> | 633 |
| 51 Evidence-based medicine <i>Martin J Burton</i> | 645 |
| 52 Critical appraisal skills <i>Martin Dawes</i> | 649 |

| | | |
|--|--|------------|
| PART 11 RECENT ADVANCES IN TECHNOLOGY – EDITED BY MARTIN J BURTON | | 673 |
| 53 | Functional magnetic resonance imaging: Principles and illustrative applications for otolaryngology <i>Paul M Matthews</i> | 675 |
| 54 | Positron emission tomography and integrated PET/computed tomography <i>Wai Lup Wong</i> | 684 |
| 55 | Image-guided surgery, 3D planning and reconstruction <i>Ghassan Alusi and Michael Gleeson</i> | 701 |
| 56 | Ultrasound in ear, nose and throat practice <i>Keshthra Satchithananda and Paul S Sidhu</i> | 711 |
| 57 | Interventional techniques <i>James V Byrne</i> | 731 |
| 58 | Laser principles in otolaryngology, head and neck surgery <i>Brian JG Bingham</i> | 742 |
| 59 | Electrophysiology and monitoring <i>Patrick R Axon and David M Baguley</i> | 748 |
| 60 | Optical coherence tomography <i>Mariah Hahn and Brett E Bouma</i> | 755 |
| 61 | Contact endoscopy <i>Mario Andrea and Oscar Dias</i> | 762 |
| PART 12 PAEDIATRIC OTORHINOLARYNGOLOGY – EDITED BY RAY CLARKE | | 769 |
| 62 | Introduction <i>Ray Clarke</i> | 771 |
| 63 | The paediatric consultation <i>Ray Clarke and Ken Pearman</i> | 776 |
| 64 | ENT input for children with special needs <i>Francis Lannigan</i> | 783 |
| 65 | Head and neck embryology <i>T Clive Lee</i> | 792 |
| 66 | Molecular otology, development of the auditory system and recent advances in genetic manipulation <i>Henry Pau</i> | 811 |
| 67 | Hearing loss in preschool children: screening and surveillance <i>Kai Uus and John Bamford</i> | 821 |
| 68 | Hearing tests in children <i>Glynnis Parker</i> | 834 |
| 69 | Investigation and management of the deaf child <i>Sujata De, Sue Archbold and Ray Clarke</i> | 844 |
| 70 | Paediatric cochlear implantation <i>Joseph G Toner</i> | 860 |
| 71 | Congenital middle ear abnormalities in children <i>Jonathan P Harcourt</i> | 869 |
| 72 | Otitis media with effusion <i>George Browning</i> | 877 |
| 73 | Acute otitis media in children <i>Peter Rea and John Graham</i> | 912 |

| | | |
|----|---|------|
| 74 | Chronic otitis media in childhood <i>John Hamilton</i> | 928 |
| 75 | Management of congenital deformities of the external and middle ear <i>David Gault and Mike Rothera</i> | 965 |
| 76 | Disorders of speech and language in paediatric otolaryngology <i>Ray Clarke and Siobhan McMahon</i> | 990 |
| 77 | Cleft lip and palate <i>Chris Penfold</i> | 996 |
| 78 | Craniofacial anomalies: genetics and management <i>Dean Kissun, David Richardson, Elizabeth Sweeney and Paul May</i> | 1019 |
| 79 | Vertigo in children <i>Gavin AJ Morrison</i> | 1040 |
| 80 | Facial paralysis in childhood <i>SS Musheer Hussain</i> | 1052 |
| 81 | Epistaxis in children <i>Ray Clarke</i> | 1063 |
| 82 | Nasal obstruction in children <i>Michelle Wyatt</i> | 1070 |
| 83 | Paediatric rhinosinusitis <i>Glenis Scadding and Helen Caulfield</i> | 1079 |
| 84 | The adenoid and adenoidectomy <i>Peter J Robb</i> | 1094 |
| 85 | Obstructive sleep apnoea in childhood <i>Helen M Caulfield</i> | 1102 |
| 86 | Stridor <i>David Albert</i> | 1114 |
| 87 | Acute laryngeal infections <i>Susanna Leighton†</i> | 1127 |
| 88 | Congenital disorders of the larynx, trachea and bronchi <i>Martin Bailey</i> | 1135 |
| 89 | Laryngeal stenosis <i>Michael J Rutter and Robin T Cotton</i> | 1150 |
| 90 | Paediatric voice disorders <i>Ben Hartley</i> | 1167 |
| 91 | Juvenile-onset recurrent respiratory papillomatosis <i>Michael Kuo and William J Primrose</i> | 1174 |
| 92 | Foreign bodies in the ear and the aerodigestive tract in children <i>A Simon Carney, Nimesh Patel and Ray Clarke</i> | 1184 |
| 93 | Tracheostomy and home care <i>Michael Saunders</i> | 1194 |
| 94 | Cervicofacial infections in children <i>Ben Hartley</i> | 1210 |
| 95 | Diseases of the tonsil <i>William S McKerrow</i> | 1219 |
| 96 | Tonsillectomy <i>William S McKerrow and Ray Clarke</i> | 1229 |
| 97 | Salivary gland disorders in childhood <i>Peter D Bull</i> | 1242 |
| 98 | Tumours of the head and neck in childhood <i>Fiona B MacGregor</i> | 1251 |

| | | |
|-----|--|------|
| 99 | Branchial arch fistulae, thyroglossal duct anomalies and lymphangioma <i>Peter D Bull</i> | 1264 |
| 100 | Gastro-oesophageal reflux and aspiration <i>Haytham Kubba</i> | 1272 |
| 101 | Diseases of the oesophagus, swallowing disorders and caustic ingestion <i>Lewis Spitz</i> | 1282 |
| 102 | Imaging in paediatric ENT <i>Neville Wright</i> | 1295 |
| 103 | Medical negligence in paediatric otolaryngology <i>Maurice Hawthorne</i> | 1305 |

Please note: The table of contents for all three volumes can be found on the *Scott-Brown* website at: www.scottbrownENT.com.

The index for all three volumes is included in Volume 3.

Contributors

Victor J Abdullah MBBS FRCS (Eng) FRCS (Edin)

Consultant, Department of ENT
United Christian Hospital; and
Chief of Service in ENT
Kowloon East Cluster, Hospital Authority; and
Honorary Clinical Associate Professor
Department of Otorhinolaryngology, Head and Neck Surgery
The Chinese University of Hong Kong
Shatin, Hong Kong

Jose M Acuin MD MSc

Professor in Otolaryngology–Head and Neck Surgery
De La Salle Health Sciences Campus
Dasmarinas, Cavite, Philippines

Richard Adamson MB BS FRCS

Consultant Otolaryngologist/Head and Neck Surgeon
Fife Hospitals NHS Trust
Scotland, UK

David Albert FRCS

Lead clinician
Department of Otolaryngology
Hospital for Sick Children
Great Ormond Street
London, UK

Lorraine M Albion MSc MRCP

Consultant Endocrinologist and Acute Physician
Queen Alexandra Hospital
Portsmouth, UK

Helen Allen MBChB MRCP MRCPCH

Specialist Registrar in Paediatrics
Addenbrookes Hospital
Cambridge, UK

Ghassan Alusi PhD FRCS (ORL–HNS)

Consultant Otolaryngologist
ENT Department
Barts and the London NHS Trust
London, UK

Mario Andrea MD PhD

Professor and Chairman
Department of Otolaryngology, Voice and Communication
Disorders
Faculty of Medicine of Lisbon
Lisbon, Portugal

Jawaher Ansari MBBS MRCP FRCR

Specialist Registrar in Clinical Oncology
Queen Elizabeth Hospital
Birmingham, UK

Mateen H Arastu MBBS BSc MRCS (Eng)

Specialist Registrar in Trauma and Orthopaedic Surgery
South West Thames Region, UK

Sue Archbold MPhil

Education Co-ordinator
The Ear Foundation
Nottingham, UK

Stig Arlinger PhD

Professor of Technical Audiology
Department of Clinical and Experimental Medicine
Division of Technical Audiology
Linköping University
Linköping, Sweden

Marcus Atlas

Professor of Otolaryngology
University of Western Australia; and
Director, Ear Science Institute Australia
Sir Charles Gairdner Hospital
Nedlands, Western Australia

Patrick R Axon MD FRCS (ORL–HNS)

Consultant Otolaryngologist and Skull Base Surgeon
Department of Otolaryngology
Cambridge University Hospitals
Cambridge, UK

E Dinakara Babu MS FRCS (Eng) FRCS (Ire) FRCS (Glas) FRCS (Inter collegiate) Diploma
in Laparoscopy (France)

Consultant and Clinical Head
Breast and Endocrine Surgery
Hillingdon Hospital
Uxbridge, UK

Claus Bachert MD PhD

Department of Oto-Rhino-Laryngology
Ghent University Hospital
Ghent, Belgium

Lydia Badia FRCS (ORL-HNS)

Consultant ENT Surgeon
Rhinology and Facial Plastics
Royal National Throat, Nose and Ear Hospital
London, UK

Jose V Sebastian Bagan MD DDS PhD

Hospital General Universitario de Valencia
Valencia, Spain

Dan Bagger-Sjöbäck MD PhD

Professor in Ear, Nose and Throat Diseases
Karolinska Institute
Stockholm, Sweden

David M Baguley BSc MSc MBA PhD

Consultant Audiological Scientist
Cambridge University Hospitals NHS Foundation Trust
Cambridge, UK

S Bahadur MS FAMS PhD

Professor of Otolaryngology and Head/Neck Surgery
All India Institute of Medical Sciences
New Delhi, India

Martin Bailey BSc FRCS

Consultant Paediatric Otolaryngologist
Great Ormond Street Hospital for Children
London, UK

John Bamford BA PhD

Ellis Llywd Jones Professor of Audiology and Deaf Education
School of Psychological Sciences
Faculty of Medical and Human Sciences
University of Manchester
Manchester, UK

Doris-Eva Bamiou MD MSc PhD

Consultant in Audiological Medicine
Department of Neuro-otology
National Hospital for Neurology and Neurosurgery; and
Honorary Senior Lecturer
Academic Unit of Audiological Medicine
University College London Institute of Child Health
Great Ormond Street Hospital
London, UK

Dev Banerjee BSc MBChB MD MRCP

Consultant Respiratory and Sleep Physician
Sleep and Ventilation Unit
Department of Respiratory Medicine
Birmingham Heartlands Hospital
Birmingham, UK

Graham Banfield MD DLO FRCS Ed (ORL-HNS)

Consultant ENT Surgeon
The Great Western Hospital
Swindon, UK

Jane A Baran PhD

Professor and Chair
Department of Communication Disorders
University of Massachusetts Amherst
Amherst, MA, USA

Michael E Baser†

Formerly of Department of Environmental Health Sciences
Johns Hopkins School of Hygiene and Public Health
Baltimore, MD, USA

Grant Bates BSc (Hons) BM Bch FRCS

Consultant ENT Surgeon
John Radcliffe Hospital
Oxford, UK

Nigel Beasley FRCS (ORL-HNS)

Consultant in Otolaryngology
Queen's Medical Centre
Nottingham, UK

Michael S Benninger MD

Chairman Department of Otolaryngology-Head and Neck Surgery
Henry Ford Hospital
Detroit, USA

Barry KB Berkovitz MSc PhD BDS FDSRCS (Eng)

Department of Anatomy and Human Sciences
School of Biomedical and Health Sciences
King's College
London, UK

Thanos Bibas MSc (Lond) DrMed FRCSI (Otol)

Consultant and Lecturer in Otolaryngology
Hippokrateion Hospital University of Athens
Athens, Greece

Carsten Bindslev-Jensen MD PhD DSc

Head, Allergy Center
Department of Dermatology
Odense University Hospital
Odense, Denmark

Brian JG Bingham MBChB FRCS Ed Glas

Consultant ENT Surgeon
Department of Otolaryngology
Victoria Infirmary; and
Honorary Senior Lecturer in Otorhinolaryngology
University of Glasgow
Glasgow, UK

† Deceased

Martin A Birchall MD (Cantab) FRCS FRCS (Oto) FRCS (ORL)
 Professor of Laryngology
 University of Bristol
 Bristol, UK; and
 Consultant in Otolaryngology Head and Neck Surgery
 Royal United Hospital Bath NHS Trust
 Bath, UK

Ian D Bottrill BM FRCS (ORL)
 Consultant Otolaryngologist; and
 Honorary Senior Lecturer
 ENT Department
 University of Oxford
 John Radcliffe Hospital
 Oxford, UK

An Boudewyns MD PhD
 Professor of Otorhinolaryngology
 Department of Otorhinolaryngology, Head and Neck Surgery
 University of Antwerp Hospital – University of Antwerp
 Antwerp, Belgium

Brett E Bouma PhD
 Associate Professor, Department of Dermatology
 Member of the Faculty of the Harvard-MIT Division of Health
 Science and Technology
 Harvard Medical School
 Boston, MA, USA

Jean Bousquet MD
 Service des Maladies Respiratoires
 Hôpital Arnaud de Villeneuve
 Montpellier, France

Patrick J Bradley MBA FRCS FRACS (Hon) FRCSLT (Hon)
 Professor of Head and Neck Oncologic Surgery
 Department of ORL-HNS
 Nottingham University Hospitals
 Queen's Medical Centre Campus
 Nottingham, UK

Stefan Brew MB ChB MHB (Hons) MSc FRANZCR
 Consultant Neuroradiologist
 The National Hospital for Neurology and Neurosurgery
 London, UK

Steven M Bromley MD
 Clinical Assistant Professor of Neurology (Medicine)
 and Attending Neurologist
 University of Medicine and Dentistry of New Jersey
 Robert Wood Johnson Medical School
 Cooper University Hospital, Camden, NJ; and
 Bromley Neurology PC, Audubon, NJ; and
 Smell and Taste Center
 University of Pennsylvania Medical Center
 Philadelphia, PA
 USA

Adolfo M Bronstein MD PhD FRCP
 Professor of Clinical Neuro-otology; and
 Head, Neuro-otology Unit
 Division of Neuroscience and Mental Health
 Charing Cross Hospital, Imperial College London; and
 Honorary Consultant Neurologist
 Charing Cross Hospital; and
 National Hospital for Neurology and Neurosurgery
 Queen Square, London, UK

Gerald Brookes FRCS
 Consultant Otologist and Neuro-Otologist
 The National Hospital for Neurology and Neurosurgery; and
 The Royal National Throat, Nose and Ear Hospital
 London, UK

George G Browning MD FRCS
 Professor of Otorhinolaryngology
 MRC Institute of Hearing Research
 Glasgow Royal Infirmary
 Glasgow, UK

Peter D Bull MB FRCS
 Consultant Otolaryngologist
 Royal Hallamshire Hospital and Sheffield Children's Hospital; and
 Honorary Senior Clinical Lecturer
 University of Sheffield
 Sheffield, UK

Martin J Burton MA DM FRCS
 Senior Clinical Lecturer
 University of Oxford; and
 Consultant Otolaryngologist
 Oxford Radcliffe NHS Trust
 Oxford, UK

James V Byrne MD FRCS FRCS
 Professor of Neuroradiology
 University of Oxford; and
 Consultant Neuroradiologist
 John Radcliffe Hospital
 Oxford, UK

Melissa Calvin MRCOG PhD BSc (Hons)
 Department of Obstetrics and Gynaecology
 Lister Hospital
 Stevenage, UK

Richard Canter PhD FRCS FRCS (Otol) Hon FRCS (Edin)
 Consultant Otolaryngologist
 Royal United Hospital; and
 Honorary Senior Lecturer
 University of Bath
 Bath, UK

Paul Carding FRCSLT
 Professor of Voice Pathology
 The Medical School, Newcastle University; and
 Head of Speech, Voice and Swallowing Department
 Otolaryngology Directorate, Freeman Hospital
 Newcastle on Tyne, UK

A Simon Carney BSc (Hons) MBChB FRCS FRACS MD

Associate Professor and Head of ENT Unit
Flinders University and Flinders Medical Centre
Adelaide, South Australia

Anna Cassoni BSc FRCP FRCR

Consultant in Clinical Oncology
University College Hospitals NHS Foundation Trust
London, UK

Helen Caulfield MBBS FRCS (ORL-HNS)

Consultant ENT Surgeon
Department of Otolaryngology
The Royal Free Hospital
London, UK

Roderick Cawson† MD FRCPATH

Formerly Emeritus Professor of Oral Medicine
Guy's Hospital
London, UK

Borka Ceranic MD ENTspec PhD

Consultant in Audiological Medicine
Department of Audiology
St George's Hospital
London, UK

Swarupsinh V Chavda MBChB DMRD FRCR

Consultant Neuroradiologist and Honorary Senior Lecturer
Queen Elizabeth Hospital
University Hospital Birmingham Foundation Trust
Birmingham, UK

Elfy B Chevretton BSc MBBS FRCS MS

Consultant Otolaryngologist
Department of ENT Surgery
Guy's and St Thomas' NHS Trust
London, UK

Peter Clarke BSc FRCS

Consultant ENT Surgeon
Charing Cross Hospital
London, UK

Ray Clarke BSc DCh FRCS FRCS (ORL)

Consultant Paediatric Otolaryngologist
Royal Liverpool University Children's Hospital
Alder Hey, Liverpool, UK

Susan Clarke MSc FRCP FRCR

Senior Lecturer and Consultant Physician
Department of Nuclear Medicine
Guy's and St Thomas' Hospital
London, UK

Rogan Corbridge MBBS BSc FRCS FRCS (ORL)

Consultant ENT Surgeon
Oxford Centre for Head and Neck Oncology
John Radcliffe Hospital
Oxford, UK

Robin T Cotton MD

Director, Pediatric Otolaryngology–Head and Neck Surgery
Children's Hospital Medical Center; and
Professor of Pediatric Otolaryngology
Department of Otolaryngology, Head and Neck Surgery
University of Cincinnati College of Medicine
Cincinnati, OH, USA

Graham J Cox BDS FRCS

Consultant ENT Surgeon; and
Macmillan Head and Neck Surgical Oncologist
John Radcliffe Hospital
Oxford, UK

Alistair Cranston MBBS FRCA

Consultant Anaesthetist
Birmingham Children's Hospital
Birmingham, UK

Cor WRJ Cremers

Department of Otorhinolaryngology
University Medical Center
St Radboud
Nijmegen, The Netherlands

Ian S Curthoys PhD

Emeritus Professor
School of Psychology
University of Sydney
Sydney, Australia

Rosalyn A Davies FRCP PhD

Consultant in Audiovestibular Medicine
Department of Neuro-Otology
The National Hospital for Neurology and Neurosurgery; and
Honorary Senior Lecturer
Institute of Neurology
Queen Square, London, UK

Martin Dawes MB BS MD (Lond) FRCGP

Chair, Family Medicine
McGill University
Quebec, Canada

Ranit De MPhil FRCS (ORL-HNS)

Consultant ENT Surgeon
University Hospital North Staffordshire NHS Trust; and
Stoke-on-Trent and Mid-Staffordshire NHS Trust
Stafford, UK

Sujata De MBBS FRCS (ORL-HNS)
Consultant Paediatric Otorhinolaryngologist
Alder Hey Children's Hospital
Liverpool, UK

Charles Diamond FRCS (Glas) Dip Pall Med
Honorary Consultant Otolaryngologist
Freeman Hospital and St. Oswald's Hospice
Newcastle upon Tyne, UK

Oscar Dias MD PhD
Associate Professor
Department of Otolaryngology, Voice and Communication
Disorders
Faculty of Medicine of Lisbon
Lisbon, Portugal

Harvey Dillon B Eng (Hons I) PhD
Director of Research
National Acoustic Laboratories
Chatswood, Australia

Richard L Doty PhD
Smell and Taste Center
University of Pennsylvania Medical Center
Philadelphia, PA, USA

M Stephen Dover FDSRCS FRCS
Consultant Oral and Maxillofacial Surgeon, and
Honorary Senior Lecturer
Department of Maxillofacial Surgery
University Hospital Birmingham NHS Foundation Trust
Birmingham, UK

Wolfgang Draf Prof Dr Med Dr HC FRCS
Director, Department of Ear, Nose and
Throat Diseases, Head and Neck Surgery
International Neuroscience Institute
Hanover, Germany

Adrian Drake-Lee MMed PhD FRCS
Consultant ENT Surgeon
Queen Elizabeth Hospital
University Hospital, NHS Trust
Birmingham, UK

Elizabeth Drewe MBBS PhD MRCP MRCPATH
Consultant Clinical Immunologist
Nottingham University Hospitals NHS Trust
Nottingham, UK

Stephen R Durham MA MD FRCP
Professor of Allergy and Respiratory Medicine
Imperial College School of Medicine
National Heart and Lung Institute
London, UK

Sunil Narayan Dutt MS DNB PhD FRCS Ed (ORL-HNS) DLO (Eng) DORL
Senior Consultant and Clinical Coordinator
Department of Otolaryngology and Head and Neck Surgery
Apollo Group of Hospitals
Bangalore, India

Charles East FRCS
Consultant Otolaryngologist, Head and Neck Surgeon
The Royal Free Hampstead NHS Trust
London, UK

Ronald Eccles BSc PhD DSc
Director, Common Cold Centre and
Healthcare Clinical Trials
Cardiff School of Biosciences
Cardiff University
Cardiff, Wales, UK

D Gareth R Evans MD FRCP
Professor, Department of Medical Genetics
St Mary's Hospital
Manchester, UK

Matthew Evans PhD
Editorial Manager
Caudex Medical
Oxford, UK

Johannes J Fagan MBChB, FCS (SA) MMed (Otol)
Professor and Chairman
Division of Otolaryngology
University of Cape Town and Groote Schuur Hospital
Cape Town, South Africa

Andrew D Farmery BSc MA MD FRCA
Senior Lecturer in Anaesthetics
Nuffield Department of Anaesthetics
University of Oxford
Oxford, UK

Neil Fergie FRCS MD
Consultant in Otorhinolaryngology Head and Neck Surgery
Kings Mill Hospital, Mansfield; and
Queens Medical Centre
Nottingham, UK

John Fleetham MB BS FRCP(C)
Professor of Medicine
Respiratory Division
University of British Columbia and Vancouver Hospital
Vancouver, BC, Canada

Bill Fleming FRACS FRCS
Consultant Endocrine Surgeon
Hammersmith Hospital
Imperial Healthcare NHS Trust
London, UK

Liam M Flood MB BS FRCS

ENT Consultant
James Cook University Hospital
Middlesbrough, UK

Adrian Fourcin PhD FLoA

Emeritus Professor, Department of Phonetics and Linguistics
University College London
London, UK

Jayne A Franklyn MD PhD FRCP FMedSci

Professor of Medicine
Division of Medical Sciences
University of Birmingham
Queen Elizabeth Hospital
Birmingham, UK

Nicole JM Freling MD PhD

Department of Radiology
Academic Medical Centre
Amsterdam, The Netherlands

David N Furness BSc PhD

School of Life Sciences
Keele University
Staffordshire, UK

Ian Gabriel MBBS BSc (Hons) MRCP (UK) DipRCPath

Department of Haematology
St Mary's Hospital Campus
Imperial College School of Medicine
London, UK

Geraldine Gallagher FRCSI

Antrim Area Hospital
Belfast, Northern Ireland

Damian Gardner-Thorpe MRCP (UK) MRCS (Eng) MRCGP (UK)

General Practitioner
The Pulteney Practice
Bath, UK

David Gault MB ChB FRCS

Consultant Plastic Surgeon
London Centre for Ear Reconstruction
The Portland Hospital
London, UK

Garrick A Georgeu MB ChB FRCS (Ed) FRCS PLAS MSc

Plastic Surgery Department
Selly Oak Hospital
University Hospital Birmingham
Birmingham, UK

Kevin P Gibbin

Consultant Otolaryngologist
University Hospital
Nottingham, UK

Ralph W Gilbert MD FRCS (C)

Professor of Otolaryngology/Head and Neck Surgery
University of Toronto
University Health Network
Princess Margaret Hospital
Toronto, Ontario, Canada

John Glaholm BSc FRCP FRCR (Clin Oncol)

Consultant Clinical Oncologist
Cancer Centre, Queen Elizabeth Hospital
Birmingham, UK

Michael Gleeson MD FRCS

Professor of Otolaryngology and Skull Base Surgery
Institute of Neurology
University College London; and
Consultant
Guy's, Kings and St Thomas' and the National Hospital for
Neurology and Neurosurgery
London UK; and
Honorary Consultant Skull Base Surgeon
Great Ormond Street Hospital for Sick Children
London, UK

Kees Graamans MD PhD

Professor and Chairman
Department of Otorhinolaryngology
University Medical Centre Nijmegen
Nijmegen, The Netherlands

John Graham MA BM BCh FRCS

Consultant Otolaryngologist
The Royal National Throat, Nose, and Ear Hospital
Gray's Inn Road
London, UK

Scott M Graham MD

Professor
Department of Otolaryngology – Head and Neck Surgery
The University of Iowa; and
Director of Rhinology, University of Iowa Hospital and Clinics
Iowa City, IA, USA

Luisa F Grymer MD

Grymer Private Hospital
Aarhus, Denmark

Carole M Hackney BSc PhD

Department of Physiology, Development and Neuroscience
University of Cambridge
Cambridge, UK

Mariah Hahn PhD

Assistant Professor
Department of Chemical Engineering
Texas A&M University
Texas, USA

Jahmal A Hairston MD
Department of Otolaryngology
University of Cincinnati College of Medicine
Cincinnati, OH, USA

Aidan Halligan FRCP FRCOG MA MD MRCPI FFPHM
Chief Executive, Elision Health Ltd; and
Deputy Chief Medical Officer, England (2003–2005); and
Director of Clinical Governance for the NHS (1999–2006)
Leicester, UK

G Michael Halmagyi MD FRACP
Clinical Professor
Department of Neurology
Royal Prince Alfred Hospital
Sydney, Australia

John Hamilton FRCS
Department of Otolaryngology
Gloucestershire Royal Hospital
Gloucester, UK

Ravinder PS Harar FRCS (ORL-HNS)
Specialist Registrar Otolaryngology, Head and Neck Surgery
The National Hospital for Neurology and Neurosurgery; and
The Royal National Throat, Nose and Ear Hospital
London, UK

Jonathan P Harcourt MA FRCS
Consultant ENT Surgeon
Charing Cross Hospital
London, UK

Meredydd Harries FRCS MSc (Voice)
Consultant ENT Surgeon
The Royal Sussex County Hospital
Brighton, UK

A John Harris PhD
Developmental Biology Laboratory
Department of Physiology
University of Otago
Dunedin, New Zealand

Douglas Harrison FRCS
Consultant Plastic Surgeon
The Wellington Hospital
London, UK

Ben Hartley MBBS BSc FRCS
Consultant Paediatric Otolaryngologist
Great Ormond Street Hospital for Children
London, UK

Richard J Harvey BScMed MB BS FRACS
Nuffield Fellow, University of Oxford, UK; and
Rhinologist and Endoscopic Skull Base Surgeon
St Vincent's Hospital
Sydney, Australia

Peter Haughton BSc PhD
Formerly Clinical Scientist and Head of Audiology
Department of Medical Physics
Royal Hull Hospitals
Kingston upon Hull, UK

Maurice Hawthorne FRCS
Consultant Otolaryngologist, Head and Neck Surgeon
James Cook University Hospital
Middlesbrough, UK

John Hibbert ChM FRCS
Formerly Consultant Otolaryngologist
Department of Otolaryngology
Guy's Hospital
London, UK

John M Hilinski MD
Facial Plastic and Reconstructive Surgery
San Diego Face and Neck Specialties
University of California, San Diego Medical Center
San Diego, CA, USA

John Hill FRCS FRCSed
Department of ORL-HNS
The Freeman Hospital
Newcastle upon Tyne, UK

Malcolm P Hilton MA BM BCh FRCS (ENG) FRCS (ORL-HNS)
Consultant Otolaryngologist
Royal Devon and Exeter Hospital; and
Honorary Clinical Lecturer
Peninsula Medical School
University of Exeter
Exeter, UK

Lisa J Hirst BSc PhD Cert MCRSLT
Head of Service
Speech and Language Therapist
Salisbury District Hospital
Wiltshire, UK

Simon Holmes BDS MBBS (Hons) FDS RCS Eng FRCS (OMFS)
Consultant Oral and Maxillofacial Surgeon
Barts and The London NHS Trust
London, UK

David Hosking MD FRCP
Consultant Physician
Division of Mineral Metabolism
City Hospital
Nottingham, UK

David J Howard BSc FRCS FRCS (Ed)
Emeritus Senior Lecturer
University College London; and
Consultant Head and Neck Surgeon
Royal Throat, Nose and Ear Hospital; and
Charing Cross Hospital
London, UK

SS Musheer Hussain MB MSc (Manc) FRCS (ORL)

Consultant Otolaryngologist and Head
ENT and Audiology Services
Ninewells Hospital and Medical School; and
Honorary Senior Lecturer and Director
Temporal Bone Laboratory; and
Licenced Teacher of Anatomy
Department of Otolaryngology
University of Dundee
Dundee, UK

Richard M Irving MD FRCS (ORL-HNS)

Consultant in Neurotology
University Hospital Birmingham NHS Trust and
Diana Princess of Wales (Birmingham Childrens) Hospital; and
Honorary Senior Lecturer
University of Birmingham
Birmingham, UK

Mark E Izzard MB BS FRACS

Senior Lecturer in Otolaryngology
University of Auckland; and
Consultant Head and Neck Surgeon
Auckland District Health Board
Auckland, New Zealand

Jean-Pierre Jeannon MB ChB FRCS (OTO) FRCS (ORL)

Consultant Ear Nose and Throat/Head and Neck Surgeon
Guy's and St Thomas' Hospital
London, UK

Chris R Jennings

Department of Otolaryngology
The Queen Elizabeth Hospital
Birmingham, UK

Dan Jiang PhD FRCSI (Otol) FRCS (ORL-HNS)

Consultant Otolaryngologist
Department of Otolaryngology, Head and Neck Surgery
Guy's, St Thomas' and Evelina Children's Hospitals
London, UK

Alan P Johnson FRCS

Department of Otolaryngology
Queen Elizabeth Hospital
Birmingham, UK

Andrew S Jones MB BCh MD FRCSE FRCS

Professor, School of Cancer Studies
Division of Surgery and Oncology
Royal Liverpool University Hospital
Liverpool, UK

Nicholas S Jones MD FRCS FRCS (ORL)

Professor of Otorhinolaryngology
Queens Medical Centre
University of Nottingham
Nottingham, UK

Petros D Karkos MPhil FRCSI

Specialist Registrar in Otolaryngology
Mersey Deanery
Chester, UK

Gerard Kelly MD FRCS (ORL-HNS)

Consultant Ear, Nose and Throat and Skull Base Surgeon; and
Clinical Director of Otolaryngology
The Leeds Teaching Hospitals NHS Trust
Leeds, UK

Andras Armand Kemeny MD FRCS

The National Centre for Stereotactic Radiosurgery
Royal Hallamshire Hospital
Sheffield, UK

David W Kennedy MD FACS FRCSI

Department of Otorhinolaryngology-Head and Neck Surgery
University of Pennsylvania
Philadelphia, PA, USA

Richard SC Kerr BSc MS MBBS FRCS

Consultant Neurosurgeon
Oxford Skull Base Unit
Oxford Radcliffe NHS Trust; and
Honorary Senior Lecturer, University of Oxford
Oxford, UK

Dean Kissun FRCS (OMFS)

Consultant Maxillofacial Surgeon
NHS Lothian
Edinburgh, UK

Jean Michel Klossek MD

ENT Professor, University of Poitiers; and
ENT and Head and Neck Surgery Department
University Hospital Jean Bernard
Poitiers, France

Gary Kroukamp MBChB FCORL (SA)

Faculty of Health Sciences
University of Stellenbosch, Tygerberg Hospital
Tygerberg, South Africa

Haytham Kubba MPhil MD FRCS (ORL-HNS)

Consultant Paediatric Otolaryngologist, Head and Neck Surgeon
The Royal Hospital for Sick Children
Glasgow, UK

Michael Kuo PhD FRCS (Eng) FRCS (ORL-HNS) DCH

Consultant Otolaryngologist – Head and Neck Surgeon
Birmingham Children's Hospital
Birmingham, UK

Francis Lannigan MB ChB MD FRCS (Eng) Ed (ORL) FRACS

Department of Otolaryngology – Head and Neck Surgery
Princess Margaret Hospital for Children; and
Clinical Professor, The University of Western Australia
Perth, Western Australia

Gavin G Lavery MB BCH BAO FCARCSI MD
 Director of Critical Care Services
 Royal Hospitals, Belfast, UK; and
 Visiting Professor, Faculty of Life and Health Sciences
 University of Ulster
 Northern Ireland

Brian Leatherbarrow BSc MBChB DO FRCS FRCOphth
 Consultant Ophthalmic, Oculoplastic and Orbital Surgeon
 Manchester Royal Eye Hospital
 Manchester, UK

John H Lee MD
 Assistant Professor
 Department of Otolaryngology – Head and Neck Surgery
 University of Iowa
 Iowa City, IA, USA

T Clive Lee MA MSc MD PhD FRCSI FRCSEd CEng FIEI
 Professor of Anatomy
 Royal College of Surgeons in Ireland
 Dublin, Ireland

Susanna Leighton† BSc FRCS (ORL-HNS)
 Formerly Consultant Paediatric Otolaryngologist
 Great Ormond Street Hospital for Children
 London, UK

Paula Leslie PhD Cert MRCSLT
 Associate Professor
 Communication Science and Disorders
 University of Pittsburgh
 Pittsburgh, PA, USA

Tristram HJ Lesser AKC FRCSEd MS
 Otolaryngology/Head and Neck Surgery
 University Hospital
 Liverpool, UK

James W Loock MB ChB (UCT) FCS (SA) ORL
 Professor and Head
 Department of Otorhinolaryngology
 University of Stellenbosch
 Tygerberg Hospital
 Cape Town, South Africa

David A Lowe BSc FRCSEd FRCS
 Research Fellow
 Clinical Effectiveness Unit
 The Royal College of Surgeons of England
 London, UK

Valerie J Lund MS FRCS FRCS (Ed)
 Professor of Rhinology
 The Ear Institute
 University College London
 London, UK

Linda M Luxon BSc MBBS FRCP
 Professor of Audiovestibular Medicine
 University of London at University College London
 Academic Unit of Audiovestibular Medicine; and
 Consultant Physician, National Hospital for
 Neurology and Neurosurgery; and
 Honorary Consultant Physician
 Great Ormond Street Hospital for Children
 London, UK

JA Lynn MS FRCS
 Consultant Surgeon
 Cromwell Hospital
 London, UK

Fiona B MacGregor MBChB FRCS (ORL HNS)
 Consultant Otolaryngologist
 Royal Hospital for Sick Children
 Glasgow, UK

Ian S Mackay FRCS
 Consultant ENT Surgeon
 Royal Brompton Hospital and Charing Cross Hospital
 London, UK

Kenneth MacKenzie MB ChB FRCS (Ed)
 Consultant Otorhinolaryngologist and Honorary Senior Lecturer
 Glasgow Royal Infirmary
 University of Glasgow
 Glasgow, UK

Marcelle Macnamara MA MBBS FRCS MPhil FRCS (ORL-HNS)
 Retired Consultant Otolaryngologist, Head and Neck Surgeon
 Heart of England Foundation Trust
 Birmingham, UK

Arnold GD Maran MD DSc FRCS (Ed, Eng, Glasg) FRCP FDS
 Emeritus Professor of Otolaryngology
 University of Edinburgh
 Edinburgh, UK

Andrew H Marshall BSc MBBS FRCS
 Consultant Otolaryngologist
 Department of Otorhinolaryngology and Head and Neck Surgery
 University Hospital
 Nottingham, UK

Stewart G Martin BSc (Hons) MSc PhD
 Associate Professor of Oncology
 MSc Course Director and Head of Translational Radiation
 Biology Research Group
 University of Nottingham
 Nottingham, UK

Robert C Mason BSc ChM MD FRCS
 Consultant Upper GI Surgeon
 Guy's and St Thomas' Hospitals
 London, UK

† Deceased

Lesley Mathieson FRCSLT

Visiting Lecturer in Voice Pathology
The Ear Institute
University College London; and
Honorary Research Adviser
Speech and Language Therapy Department
Royal National Throat Nose and Ear Hospital
London, UK

Paul M Matthews MA (Oxon) MD DPhil FRCP

Vice-President for Imaging Genetics and for Neurology; and
Head, GSK Clinical Imaging Center
Clinical Pharmacology and Discovery Medicine GlaxoSmithKline;
and Professor of Clinical Neurosciences
Department of Clinical Neurosciences
Imperial College, London; and
(Hon.) MRC Clinical Research Professor
Department of Clinical Neurology
University of Oxford
Oxford, UK

Paul May MBBS FRCS FRCPCH

Consultant Paediatric Neurosurgeon
Craniofacial Unit, Alder Hey Children's Hospital
Liverpool, UK

Thomas McCaffrey MD PhD

Professor and Chair
Department of Otolaryngology Head and Neck Surgery
University of South Florida
Tampa, FL, USA

Leo McClymont MBChB MD FRCS(Ed) FRCS(Glas)

Raigmore Hospital
Highland Acute Hospitals NHS Trust Inverness
Inverness, UK

Andrew McCombe MD FRCS (ORL)

Consultant ENT Surgeon
Frimley Park Hospital
Camberley, UK

Gerald W McGarry MD FRCS (RCPSGlasg) FRCS(Ed) FRCS (ORL-HNS)

Consultant Otorhinolaryngologist
Glasgow Royal Infirmary; and
Honorary Senior Lecturer
University of Glasgow
Glasgow, UK

Julian A McGlashan MBBS FRCS (ORL)

Special Lecturer and Consultant
Department of Otorhinolaryngology
Queen's Medical Centre Campus
Nottingham University Hospitals
Nottingham, UK

Mark McGurk MD BDS FRCS FDSRCS DLO

Consultant in Oral and Maxillofacial Surgery
Guy's Hospital
London, UK

Stephen McHanwell BSc PhD MBIol CBiol

Professor of Anatomical Sciences; and
National Teaching Fellow 2007; and
Director of Stage 1 & 2 BDS
School of Dental Sciences
Dental School
Newcastle upon Tyne, UK

Michael J McKenna MD

Professor, Department of Otolaryngology
Harvard Medical School; and
Surgeon, Department of Otolaryngology
Massachusetts Eye and Ear Infirmary
Boston, MA, USA

William S McKerrow MB ChB MRCPG (exam) FRCS(Ed & Glasg)

Consultant Otolaryngologist
Department of ENT/Head and Neck Surgery
Raigmore Hospital
Inverness, UK

Siobhan McMahon BSc MRCSLT

Speech and Language Therapy
Department Alder Hey Hospital
Liverpool, UK

Brent A McMonagle MBBS FRACS

Department of Otolaryngology
Guy's Hospital
London, UK

Hisham Mehanna BMedSc (Hons) MBChB (Hons) FRCS (ORL-HNS)

Consultant ENT – Head and Neck and Thyroid Surgeon; and
Honorary Senior Lecturer
University Hospitals Coventry and Warwickshire
Walsgrave Hospital
Coventry, UK

Saamil N Merchant MD

Gudrun Larsen Eliassen and Nels Kristian Eliassen
Professor of Otolaryngology
Harvard Medical School; and
Surgeon in Otolaryngology and
Director of Otopathology Laboratory
Department of Otolaryngology
Massachusetts Eye and Ear Infirmary
Boston; and
Affiliate Faculty Member
Harvard University–Massachusetts Institute of Technology
Division of Health Sciences and Technology
Cambridge, MA, USA

David Miles FRCP MD

Consultant in Medical Oncology
Mount Vernon Cancer Centre
Middlesex, UK

Christopher A Milford FRCS

Consultant Otolaryngologist
Oxford Skull Base Unit
John Radcliffe Hospital
Oxford, UK

Robert Mills MS MPhil FRCS (Eng) FRCS (Ed)

Otolaryngology Unit
University of Edinburgh
Royal Infirmary of Edinburgh
Edinburgh, UK

Steven Ross Mobley MD

Director of Facial Plastic and Reconstructive Surgery
Division of Otolaryngology-HNS
University of Utah School of Medicine
Salt Lake City, UT, USA

David Moffat BSc MA FRCS

Consultant Neuro-Otologist
Department of Otoneurological and Skull Base Surgery
Addenbrookes
Cambridge University Teaching Hospital NHS Foundation Trust;
and Associate Lecturer, Cambridge University
Cambridge, UK

Brian CJ Moore MA PhD FMedSci FRS

Professor of Auditory Perception
Department of Experimental Psychology
University of Cambridge
Cambridge, UK

David AL Morgan FRCR

Consultant Clinical Oncologist
Department of Clinical Oncology
Nottingham University Hospitals
Nottingham, UK

Jonathan M Morgan MD

Instructor
Department of Otolaryngology Head and Neck Surgery
University of South Florida
Tampa, FL, USA

Juliette Morgan MD

Division of Foodborne Bacterial and Mycotic Diseases
National Center for Zoonotic, Vector-Borne and Enteric Diseases
Centers for Disease Control and Prevention
Atlanta, GA, USA

Gavin AJ Morrison MA MBBS FRCS

Consultant ENT Surgeon
Guy's, St Thomas' and Evelina Hospitals
London, UK

Randall P Morton MB MSc FRACS

Professor of Otolaryngology
University of Auckland; and
Consultant Otolaryngologist-Head and Neck Surgeon
Counties Manukau and Auckland District Health Boards
Auckland, New Zealand

Frank E Musiek PhD

Professor and Director of Auditory Research
Department of Communication Sciences; and
Professor of Otolaryngology
School of Medicine University of Connecticut
Storrs, CT, USA

Niels Mygind MD

Formerly Consultant in Lung Medicine
Department of Respiratory Medicine
University Hospital of Aarhus
Aarhus, Denmark

Karl G Nicholson MBBS MRCS FRCP

Professor of Infectious Diseases
Department of Infectious Diseases and Tropical Medicine
Leicester Royal Infirmary
Leicester, UK

Andrew J Nicol MBChB, FCS (SA)

Associate Professor
General Surgery; and
Head of Trauma Unit
Groote Schuur Hospital
Cape Town, South Africa

Gilbert J Nolst Trenité MD PhD

Professor of Otorhinolaryngology
Academic Medical Center
University of Amsterdam
The Netherlands

Desmond A Nunez FRCS (ORL) MD

Director, Department of Otolaryngology
North Bristol NHS Trust; and
Honorary Senior Lecturer
University of Bristol
Bristol, UK

Michael O'Connell BSc, MPhil, FRCS

Consultant Otorhinolaryngologist,
Facial Plastic Surgeon and Honorary Senior Lecturer
Brighton and Sussex University Hospitals NHS Trust
Brighton, UK

Alec Fitzgerald O'Connor FRCS

Consultant Otolaryngologist
St Thomas Hospital
London, UK

Paul O'Flynn FRCS

Consultant ENT/Head and Neck Surgeon
University College Hospitals; and
Honorary Consultant
The Royal National Throat, Nose and Ear Hospital
London, UK

Stephen O'Leary MB BS BMedSc PhD FRACS

The Department of Otolaryngology
Royal Victorian Eye and Ear Hospital
East Melbourne, Australia

Morten Osterballe MD

Allergy Center, Odense University Hospital
Odense, Denmark

Peter O'Sullivan BSc MPhil FRCSI (ORL-HNS)

Clinical Fellow, Neurotology
Department of Otolaryngology
Sir Charles Gairdner Hospital
Nedlands, Western Australia

William J Owen† MS FRCS

Formerly Oesophageal Investigation Unit
Department of Surgery, St Thomas' Hospital
London, UK

Jaideep J Pandit MA BM DPhil FRCA

Consultant Anaesthetist
Nuffield Department of Anaesthetics
University of Oxford
Oxford, UK

Andrew J Parker MBChB (hons) DLO ChM FRCS

Consultant ENT Surgeon
Department of Otolaryngology
Royal Hallamshire Hospital
Sheffield, UK

Glynnis Parker MB ChB FRCP DCH MSc

Audiovestibular Physician
Sheffield Children's Hospital
Sheffield, UK

Nimesh Patel MBChB FRCS FRCS (ORL-HNS)

Consultant Otolaryngologist
Southampton General Hospital
Liverpool, UK

John P Patten BSc MB FRCP

Consultant Neurologist (retired)
South West Thames Regional Health Authority
London, UK

Henry Pau MD MBChB FRCSed FRCS Ed (ORL-HNS) FRCS

Consultant Otorhinolaryngologist; and
Honorary Senior Lecturer
University Hospitals of Leicester
Leicester, UK

Santdeep H Paun FRCS (ORL-HNS)

Consultant Nasal and Facial Plastic Surgeon
St Bartholomew's Hospital
London, UK

Sarah Payne BSc (Hons) MRCP

SpR in Medical Oncology
Centre for Tumour Biology
Institute of Cancer and the CR-UK Clinical Centre
Barts and the London
Queen Mary's School of Medicine and Dentistry
London, UK

Adrian Pearce FRCA

Consultant Anaesthetist
Department of Anaesthesia
Guy's and St Thomas' Hospital
London, UK

Ken Pearman FRCS

Consultant Paediatric Otolaryngologist
Children's Hospital
Birmingham, UK

Chris Penfold FDSRCS FRCS

Consultant Oral and Maxillofacial Surgeon
Alder Hey Children's Hospital
Liverpool, UK

A Graeme B Perks FRCS FRCS (Plast) FRACS

Consultant Plastic Surgeon
The City Hospital
Nottingham, UK

Alison Perry PhD FRCSLT

Chair, School of Human Communication Sciences
Faculty of Health Sciences
La Trobe University
Melbourne, Australia

James O Pickles MA MSc PhD DSc

Head of Hearing Unit
Vision, Touch and Hearing Research Centre
Department of Physiology and Pharmacology
University of Queensland
Brisbane, Australia

Lisa Pitkin BSc MSc FRCS ORL-HNS

Specialist Registrar in Otolaryngology
South (West) Thames Otolaryngology Training Region
Royal Marsden NHS Foundation Trust
London, UK

Laysan Pope BSc MB BS MRCS

Specialist Registrar in Otolaryngology, Head and Neck Surgery
John Radcliffe Hospital
Oxford, UK

Stephen R Porter BSc MD PhD FDS RCS FDS RCSE

Professor of Oral Medicine
UCL Eastman Dental Institute
London, UK

Richard J Powell MBBS DM FRCP FRCPath

Consultant and Professor in Clinical Immunology
University of Nottingham
Nottingham, UK

Paul Pracy BSc MBBS FRCS (Glas) FRCS (ORL-HNS)

Consultant Head and Neck Surgeon
Department of Otorhinolaryngology/Head and Neck Surgery
Queen Elizabeth Hospital
University Hospital Birmingham NHS Trust
Birmingham, UK

Hillel Pratt PhD

Evoked Potentials Laboratory
Technion – Israel Institute of Technology
Haifa, Israel

Tim Price BSc MBChB MRCS DLO FRCS (ORL-HNS)

Consultant Otolaryngologist, Head and Neck Surgeon
Dorset County Hospital
Dorchester, UK

William J Primrose MB FRCS

Consultant Otolaryngologist/Head and Neck Surgeon
Royal Victoria Hospital, Belfast
Northern Ireland, UK

Matthias Radatz MD FRCS

The National Centre for Stereotactic Radiosurgery
Royal Hallamshire Hospital
Sheffield, UK

Ullas Raghavan FRCS (ORL-HNS)

Consultant Ear Nose and Throat and Facial Plastic Surgeon
Doncaster Royal Infirmary
Doncaster, UK

Gunesh P Rajan MD FMH (Ch) FRACS

Senior Lecturer of Otolaryngology, Head and Neck Surgery
Department of Otolaryngology, Head and Neck Surgery
University of Western Australia
Fremantle, Australia

James Ramsden PhD FRCS

Specialist Registrar in Otolaryngology/Head and Neck Surgery
John Radcliffe Hospital
Oxford, UK

Richard Ramsden FRCS

Manchester Royal Infirmary
Manchester, UK

Sheila C Rankin FRCR

Consultant Radiologist
Guy's and St Thomas' Hospital NHS Trust
London, UK

Helge Rask-Andersen MD PhD

Professor in Experimental Otology
Department of Otolaryngology
Uppsala University Hospital
Uppsala, Sweden

Peter Rea MA FRCS (Eng) FRCS (ORL-HNS)

Consultant Otolaryngologist
Leicester Royal Infirmary
Leicester, UK

Fiona Regan MBBS FRCP FRCPath

Consultant Haematologist
Department of Haematology
Imperial College School of Medicine; and
Honorary Senior Lecturer and Consultant Haematologist
National Blood Service
London, UK

Claud Regnard FRCP (Lon)

Consultant in Palliative Care Medicine
St. Oswald's Hospice, Newcastle-upon-Tyne; and
Freeman Hospital (Newcastle Hospitals NHS Trust)
Newcastle-upon-Tyne and
Northumberland Tyne and Wear NHS Trust
Northumberland, UK

Evan Reid BSc MB ChB PhD FRCP

Wellcome Trust Senior Research Fellow in Clinical Science; and
Honorary Consultant in Medical Genetics
Department of Medical Genetics and
Cambridge Institute for Medical Research
Addenbrooke's Campus, University of Cambridge
Cambridge, UK

Gerhard Rettinger Prof Dr Med

Head ENT-University-Department
Ulm, Germany

David Richardson FRCS FDSRCS

Consultant Maxillofacial Surgeon
Supra Regional Paediatric Craniofacial Unit
Royal Liverpool Childrens Hospital; and
Maxillofacial Unit
University Hospital Aintree
Liverpool, UK

Peter J Robb BSc (Hons) MB BS FRCS FRCSEd

Epsom and St Helier University Hospitals NHS Trust
Surrey, UK

David Roberts FRCS

St Thomas and Guy's Hospital NHS Trust
London, UK

Philip J Robinson MB ChB FRCS FRCS (Otolaryngology)

Consultant Otolaryngologist
ENT Department, Southmead Hospital
Bristol, UK

Nicholas J Roland MBChB MD FRCS

Consultant ENT/Head and Neck Surgeon
University Hospital Aintree
Liverpool, UK

Geoffrey E Rose DSc MS MRCP FRCS FRCOphth

Consultant Orbital Surgeon
Moorfields Eye Hospital
London, UK

Rob Ross Russell MD FRCPCH

Consultant in Paediatric Intensive Care and Respiratory Medicine
Addenbrooke's Hospital
Cambridge, UK

Mike Rothera MBBS FRCS

Consultant Paediatric ENT Surgeon
Royal Manchester Childrens' Hospital
Manchester, UK

Jeremy Rowe MA DM FRCS (SN)

The National Centre for Stereotactic Radiosurgery
Royal Hallamshire Hospital
Sheffield, UK

Julian Rowe-Jones MB BS FRCS (ORL)

Consultant Rhinologist and Nasal Plastic Surgeon
Department of Otorhinolaryngology – Head and Neck/
Facial Plastic Surgery
Royal Surrey County Hospital
Guildford, UK

Claudia Rudack PD Dr Med

ENT-University-Department
Münster, Germany

Michael J Rutter FRACS

Division of Pediatric Otolaryngology/Head and Neck Surgery,
Cincinnati Children's Hospital Medical Center; and
Associate Professor of Pediatric Otolaryngology
Department of Otolaryngology, Head and Neck Surgery
University of Cincinnati College of Medicine
Cincinnati, OH, USA

Shakeel R Saeed MBBS (Lon) FRCS (Ed) FRCS (Eng) FRCS (Orl) MD (Man)

Consultant ENT and Skull Base Surgeon
University Department of Otolaryngology–
Head and Neck Surgery
Manchester Royal Infirmary and Hope Hospital
Manchester, UK

Hesham Saleh MBBCh FRCS FRCS (ORL-HNS)

Consultant Rhinologist/Facial Plastic Surgeon
Charing Cross Hospital and the Royal Brompton Hospital; and
Honorary Senior Lecturer
Imperial College of Medicine
London, UK

Robert J Sanderson† MB ChB FRCS (Ed) FRCS (Eng) FRCS (ORL-HNS)

Formerly Consultant Otolaryngologist/Head and Neck Surgeon
Western General Hospital
Edinburg, UK

Keshthra Satchithananda BDS FDSRCS MB BS FRCS FRCR

Consultant Radiologist
Department of Radiology
Charing Cross Hospital
London, UK

Michael Saunders MD FRCS

Consultant Otolaryngologist
Department of Otorhinolaryngology, Head and Neck Surgery
St Michael's Hospital
Bristol, UK

Glenis Scadding MA MD FRCP

Consultant Immunologist, Rhinologist and Allergy Specialist
Royal National Throat Nose and Ear Hospital
London, UK

Jochen Schacht PhD

Professor and Director
Kresge Hearing Research Institute
Department of Otolaryngology
University of Michigan
Ann Arbor, MI, USA

Rodney J Schlosser MD

Department of Otolaryngology
Medical University of South Carolina
Charleston, SC, USA

Stephan Schmid MD

Professor of Otolaryngology
Department of Otorhinolaryngology, Head and Neck Surgery
Universitätsspital Zurich
Zurich, Switzerland

Colin A Scotchford PhD

Associate Professor
School of Mechanical, Materials and Manufacturing Engineering
University of Nottingham
Nottingham, UK

Andrew Scott FRCS (ORL-HNS) MPhil

The Royal Shrewsbury Hospital
Shrewsbury, UK

Crispian Scully CBE MD PhD MDS MRCS FDSRCS FDSRCPs FFDRCsI FDSRCSCE FRCPath

FMedSci DSc
Professor of Oral Medicine, Pathology and Microbiology
University of London; and
Professor of Special Care Dentistry
UCL-Eastman Dental Institute
London, UK

Su-Hua Sha MD

Research Investigator
Kresge Hearing Research Institute
Department of Otolaryngology
University of Michigan
Ann Arbor, MI, USA

Naomi Sibtain MBBS MRCP FRCR

Consultant Neuroradiologist
King's College Hospital
London, UK

Paul S Sidhu BSc MB BS MRCP FRCR DTM&H

Senior Lecturer and Consultant Radiologist
Department of Radiology
King's College Hospital
London, UK

Richard Sim MD FRCS (Oto)

Department of Ear, Nose and Throat
Royal United Hospital
Bath, UK

† Deceased

Paul Simons MBBS BSc MRCP MRCGP DCH DRCOG DFFP

Marcham Road Health Centre
Abingdon, UK

Robert Slack BSc MB ChB FRCS (Ed) FRCS (Eng)

Department of Ear, Nose and Throat
Royal United Hospital
Bath, UK

Wendy Smith BPharm MBBS DLO FRCS (ORL-HNS)

Locum Consultant Otorhinolaryngology
The Leeds Teaching Hospitals NHS Trust
Leeds, UK

Lewis Spitz PhD FRCS

Institute of Child Health (University College London) and
Great Ormond Street Hospital for Children
London, UK

Jacob Bertram Springborg MD PhD

University Clinic of Neurosurgery
The Neuroscience Centre
Copenhagen University Hospital
Copenhagen, Denmark

Nicholas D Stafford MB FRCS

Director, Postgraduate Medical Institute
University of Hull
Hull, UK

H Stammberger MD Hon FRCS (Ed) Hon FRCS (Engl)

Professor and Head
Department of General ORL, H & NS
Medical University
Graz, Austria

Michael Stearns BDS MB BS FRCS

The Royal Free Hospital
London, UK

Karen P Steel PhD FMedSci

The Wellcome Trust Sanger Institute
Hinxton, UK

Paul Stewart FRCP

Department of Medicine
Queen Elizabeth Hospital
Birmingham, UK

Iain RC Swan MB ChB MD FRCS (Ed)

Department of Otolaryngology
North Glasgow University NHS Trust
Glasgow, UK

Elizabeth Sweeney FRCP DRGOC MD

Consultant Clinical Geneticist
Craniofacial Unit
Alder Hey Children's Hospital
Liverpool, UK

Andrew C Swift ChM FRCS FRCS (Ed)

Consultant in Otorhinolaryngology
University Hospital Aintree
Liverpool, UK

Andra E Talaska BS

Kresge Hearing Research Institute
Department of Otolaryngology
University of Michigan
Ann Arbor, MI, USA

Thomas A Tami MD

Professor of Otolaryngology
Department of Otolaryngology
University of Cincinnati College of Medicine
Cincinnati, OH, USA

Rinze A Tange MD PhD UHD

Associate Professor of Otolaryngology
Department of ORL, Head and Neck Surgery
Academic Medical Centre
University of Amsterdam
Amsterdam, The Netherlands

A Thakar MS FRCS

Associate Professor of Otolaryngology and Head/Neck Surgery
All India Institute of Medical Sciences
New Delhi, India

J Regan Thomas MD

Francis L. Lederer Professor and Head
University of Illinois at Chicago
Department of Otolaryngology – Head and Neck Surgery
Chicago, IL, USA

Jens Thomsen MD DMSc FRCS

Professor of Otolaryngology
Department of Otorhinolaryngology, Head and Neck Surgery
Gentofte Hospital, University of Copenhagen
Hellerup, Denmark

Matthew J Thurtell MSc (Med) MBBS FRACP

Neuro-Ophthalmology Fellow
Department of Neurology
University Hospitals of Cleveland
Cleveland, OH, USA

Bo Tideholm MD PhD

ENT Specialist
Department of Otorhinolaryngology
University Hospital
Malmö, Sweden

Paul Tierney BA BM BCh (Oxon) FRCS (Eng.) FRCS (ORL-HNS)

Consultant Otolaryngologist – Head and Neck Surgeon
North Bristol NHS Trust; and
Honorary Senior Lecturer
Bristol University
Bristol, UK

Ian Todd PhD

Associate Professor and Reader in Cellular Immunopathology
University of Nottingham
Nottingham, UK

Joseph G Toner MB MA FRCS

Consultant/Honorary Senior Lecturer, Otolaryngology
Belfast HSC Trust
Queens University
Belfast, UK

Michael Chi Fai Tong MBChB (CUHK) MD (CUHK) FRCS (Ed) FHKAM (ORL)

Professor and Head of Academic Divisions
Department of Otorhinolaryngology, Head and Neck Surgery
The Chinese University of Hong Kong
Hong Kong

Dean M Toriumi MD

Division of Facial Plastic and Reconstructive Surgery
Department of Otolaryngology – Head and Neck Surgery
University of Illinois at Chicago
Chicago, IL, USA

Mirko Tos Prof MD DMSc Dr Hc

Emeritus Professor, Ear, Nose and Throat Department
Gentofte Hospital
University of Copenhagen
Hellerup, Denmark; and
Professor of Otolaryngology
University of Maribor
Maribor, Slovenia

Stephen C Toynton MB FRCS (ORL)

Consultant Otorhinolaryngologist
Derriford Hospital, Plymouth Hospitals NHS Trust; and
Otology Advisor to Diving Diseases Research Centre and
Hyperbaric Medical Unit
Plymouth, UK

Kai Uus MD PhD

Lecturer in Audiology
School of Psychological Sciences
Faculty of Medical and Human Sciences
University of Manchester
Manchester, UK

Peter Valentine BSc FRCS (ORL-HNS)

Consultant Otolologist and ENT Surgeon
Royal Surrey County Hospital NHS Trust
Guildford; and
Ashford and St Peter's Hospitals NHS Trust
Chertsey, UK

Jan HP van der Meulen PhD FFPH

Reader in Clinical Epidemiology
Health Services Research Unit
London School of Hygiene and Tropical Medicine
London, UK

C Andrew van Hasselt MBChB FRCS FRCS (Edin) FCS (SA)

Chairman, Department of Surgery; and
Professor of Surgery (Otorhinolaryngology)
Department of Otorhinolaryngology, Head and Neck Surgery
The Chinese University of Hong Kong
Shatin, Hong Kong

Adriaan F van Olphen MD PhD

ENT Surgeon
University Medical Centre Utrecht
Utrecht, The Netherlands

Archana Vats MA (Cantab) FRCS (Eng) FRCS (Oto) PhD

Imperial College and St. Mary's NHS Trust
London, UK

Antonio M Vignola†

Formerly of Istituto di Fisiopatologia Respiratoria
Università Palermo
Palermo, Italy

Alexander C Viantis MBBCh FCS (SA) FCSHK

Associate Professor
Department of Otorhinolaryngology, Head and Neck Surgery
The Chinese University of Hong Kong
Shatin, Hong Kong

Sherryl Wagstaff FRACS

Consultant Otolologist
Royal Victorian Eye and Ear Hospital
Melbourne University Teaching Hospital
East Melbourne, Australia

Debbie Wall BEd (Hons) MA

Senior Researcher
NHS Clinical Governance Support Team
Leicester, UK

David Ward MBBS FRCS FRCS (Ed)

Consultant Plastic Surgeon
Leicester Royal Infirmary
Leicester, UK

David W Warnock PhD FRCPath

Division of Foodborne Bacterial and Mycotic Diseases
National Center for Zoonotic, Vector-Borne and Enteric Diseases
Centers for Disease Control and Prevention
Atlanta, GA, USA

Katherine Wasson BA PhD MPH

Chief, Clinical Ethics Service; and
Assistant Professor, Critical Care
The University of Texas M.D. Anderson Cancer Center
Houston, Texas, USA

John C Watkinson MSc MS FRCS (Ed, Glas, Lond) DLO

Consultant Head and Neck and Thyroid Surgeon
Department of Otorhinolaryngology/Head and Neck Surgery
Queen Elizabeth Hospital
University of Birmingham NHS Trust
Birmingham, UK

† Deceased

Desmond Watson BM BCh MA FRCS

Former Consultant Ear Nose and Throat Surgeon and Advisor
Medical Protection Society
Leeds, UK

Keith Webster MMedSci FRCS FRCS (OMFS) FDSRCS

Consultant Oral and Maxillofacial Surgeon
University Hospital Birmingham NHS Foundation Trust; and
Honorary Senior Lecturer
Faculty of Dentistry and Medicine
University of Birmingham
Birmingham, UK

Vivienne Weston MBBS FRCP MSc FRCPath

Consultant Medical Microbiologist
Nottingham University Hospitals NHS Trust
Nottingham, UK

Richard Wight MB BS FRCS Eng (Otol) FRCS Ed (Otol)

Consultant Head and Neck Surgeon
James Cook University Hospital
Middlesbrough, UK

Janet A Wilson BSc MD FRCSEd FRCS Eng

Professor of Otolaryngology, Head and Neck Surgery
Newcastle University Freeman Hospital
Newcastle Upon Tyne, UK

Wai Lup Wong BA (Hons) MRCP FRCR

Paul Strickland Scanner Centre
Mount Vernon Hospital
Northwood, UK

John Kong Sang Woo MBBS FCSHK FRCSEd FHKAM (Otorhinolaryngology)

Consultant, Department of ENT, Prince of Wales Hospital; and
Chief of Service in ENT
New Territories East Cluster, Hospital Authority; and
Honorary Clinical Associate Professor
Department of Otorhinolaryngology, Head and Neck Surgery
The Chinese University of Hong Kong
Hong Kong

Tim J Woolford MD FRCS (ORL)

Consultant in Otorhinolaryngology
Manchester Royal Infirmary
University of Manchester
Manchester, UK

Peter-John Wormald MD FRACS FRCS (Ed) FCS (SA) MBChB

Department of Otolaryngology, Head and Neck Surgery
Adelaide and Flinders Universities
Adelaide, Australia

Steve Worrollo FIMPT

Consultant Maxillofacial Prosthetist
Department of Maxillofacial Surgery
University Hospital Birmingham NHS Trust
Birmingham, UK

Neville Wright DMRD FRCR

Consultant Paediatric Radiologist
Central Manchester and
Manchester Children's Hospitals NHS Trust
Department of Radiology
Royal Manchester Children's Hospital
Manchester, UK

Tony Wright LLM DM FRCS Tech RMS

Professor of Otolaryngology
UCL Ear Institute
London, UK

Floris L Wuyts PhD

Professor of Medical Physics
University of Antwerp; and
Head of AUREA (Antwerp University Research Centre for
Equilibrium and Aerospace)
Department of ENT
University Hospital of Antwerp
Antwerp, Belgium

Michelle Wyatt MA FRCS (ORL-HNS)

Consultant Paediatric Otolaryngologist
Great Ormond Street Hospital
London, UK

Clare Wykes BSc MRCP DipRCPPath

Haematology SpR
Hammersmith Hospitals NHS Trust
London, UK

Stephen R Young BSc (Hons) PhD

Faculty of Science
The American International University in London
Surrey, UK

Preface

Fifty-five years have passed since the first edition of *Scott-Brown's Otorhinolaryngology: Head and Neck Surgery* was published. Many otorhinolaryngologists have read at least one edition, committed it to memory and passed their specialist examinations because of it. All will have kept referring to it throughout their careers and remember it with affection. Looking back it is apparent that a radical change in structure and format has taken place every 15 to 20 years. It is 20 years since Alan Kerr made the last radical change with the publication of the 5th edition, 20 years that have seen an information technology explosion. The internet, on-line libraries, e-delivery of journals and increasingly books, computerised search engines, CD-ROMs, DVDs, digital photography; the list goes on. These technological advances have transformed medical education, influenced significantly the way the current generation learns and the methods by which their competencies and knowledge are assessed. Certainly sufficient time has elapsed for *Scott-Brown* to evolve dramatically once more. This edition has been completely re-written. It bears little resemblance to its predecessors other than by title, and in its philosophy to provide a complete resume of the knowledge base that underpins modern ORL practice and which will guide clinicians in their everyday patient care for years to come. The number of chapters has almost doubled, with large topics dissected into more digestible parts. This reflects the expansion of our specialty such that it is now a group of subspecialties linked by the common thread, each concerned with, and committed to, the care of patients with disorders of the head and neck.

Our authors are the leading experts in their respective fields of interest and have been selected from all over the world. Almost all the text is illustrated in colour and it comes with its own CD-ROM, containing all the text and illustrations in an accessible and searchable form, with references linked to PubMed.

So what else could the trainee or practising otorhinolaryngologist want from the definitive reference to the field at the beginning of the new millennium? Quite simply, the level of evidence for the advice we offer and the practice we undertake. Nowadays specialties need to define best clinical practice, if only to guide and remind health care providers of their duty to their patients to practice in accordance with accepted evidence and to

strive for excellence in clinical standards at all times. Surgeons also need to know how their actions might be viewed by the courts and the areas of practice that are currently exercising the legal profession. This edition has tried to provide that information.

It has not been an easy task for our contributors, some of whom were not writing in their mother tongue. That they were able to write to a structured format was much to their credit. I was fortunate to recruit, and am extremely grateful to, my team of section editors all of whom worked tirelessly with a common purpose. George G Browning, Martin J Burton, Ray Clarke, John Hibbert, Nicholas S Jones, Valerie J Lund, Linda M Luxon and John C Watkinson represent some of the very best and most respected clinicians in the United Kingdom, each one an international authority, each one with a heavy professional commitment. Alan Kerr's advice and encouragement throughout was always welcome and extremely useful. Marcelle McNamara came to my aid and assistance numerous times during the project. She gave tirelessly of her time and energy during a very serious illness, writing chapters and putting others into format and a more readable form. She was an example and inspiration throughout.

The creation of this edition has also been an interesting experience for the publishing staff. During a lengthy period of gestation, this text has changed ownership several times as the publishing houses traded and realigned their lists. Without the drive and perseverance of Zelah Pengilly and Jo Koster from Hodder Education it would surely have fallen by the wayside. Words cannot express my gratitude to them adequately. Understanding when clinical work overwhelmed me, they hid their frustrations over slow progress or irritatingly incomplete manuscripts. They buoyed us all up when the end seemed so far away.

Sadly, some of our contributors will never see their chapters in print as they have died during the preparation of this text. Some had long, unpleasant illnesses but wrote despite them. Others were cut down unexpectedly in their prime but have now left a legacy, and a few were my close friends and colleagues. I am proud to have my name linked permanently through this publication with Michael Baser, Roderick Cawson, Susanna Leighton,

William Owen, Robert Sanderson and Antonio Vignola. We hope that their families will draw some comfort also by seeing their loved ones live on in this book.

Finally, there are four very special people whose constant love and affection drives me on through life. They are of course my wife, Ann, and our children,

Andrew, Clare and Mark. They too will breathe a deep sigh of relief with the publication of this text and I thank them with all my heart.

Michael Gleeson
September 2007

How to use this book

This new edition of *Scott-Brown's Otorhinolaryngology, Head and Neck Surgery* incorporates some special features to aid the readers' understanding and navigation of the text. These are described below.

SEARCH STRATEGY

The majority of the chapters feature a search strategy indicating the key words used by the author when conducting their literature review in order to prepare the chapter, so that the reader can repeat and develop the search.

EVIDENCE SCORING

For the major sections in each chapter, the authors have used a hierarchical system to indicate the level of evidence supporting their statements. This is shown in the text in the form [***], with the number of stars indicating the level of evidence. The key to this system is shown in the table below.

| Level | Category of evidence |
|-------|--|
| **** | Systematic reviews, meta-analyses of randomized controlled trials and randomized controlled trials |
| *** | Non-randomised studies |
| ** | Observational or non-experimental studies |
| * | Expert opinion |

Where no level is shown, the quality of supporting evidence, if any exists, is of low grade only (for example, case reports, clinical experience etc.). For more information on evidence scoring, please refer to Chapter 304, Evidence-based medicine; and 305 Critical appraisal skills.

CLINICAL RECOMMENDATIONS

The authors have indicated the basis on which they have made clinical recommendations by grading them according to the level of the supporting evidence. This is shown in the text in the form [Grade A], with the grade indicating the level of evidence supporting the recommendation. The key to this system is shown in the table below.

| Grade | Nature of supporting evidence |
|-------|--|
| A | Recommendation based on evidence from meta-analyses of randomized controlled trials |
| B | Recommendation based on evidence from high quality case-controlled or cohort studies |
| C | Recommendation based on evidence from low quality case-controlled or cohort studies |
| D | Recommendation based on evidence from clinical series or expert opinion |

Recommendations are graded where the author is satisfied that the literature supports such a grading; otherwise a grading may not be given.

REFERENCE ANNOTATION

The reference lists are annotated with an asterisk, where appropriate, to guide readers to key primary papers and major review articles. We hope that this feature will render the lists of references more useful to the reader and will encourage self-directed learning among both trainees and practicing physicians.

Abbreviations

| | | | |
|---------|--|--------------------|---|
| 2,3DPG | 2,3-diphosphoglycerate | Ad-VEGF | adenovirus-encoding vascular endothelial growth factor |
| 2D | two-dimensional | AECRS | acute exacerbation of chronic rhinosinusitis |
| 3,4-DAP | 3,4-diaminopyridine | AED | aerodynamic equivalent diameter |
| 3D | three-dimensional | AEDS | atopic eczema dermatitis syndrome |
| 5-FdUMP | 5-fluoro-2 deoxyuridine monophosphate | AEF | auditory-evoked cortical magnetic field |
| 5-FU | 5-fluorouracil | AF | atrial fibrillation; or anterior fontanelle |
| 5-FUMP | 5-fluorouridine monophosphate | AFB | acid-fast bacilli |
| 5-HT | 5-hydroxytryptamine | AFRS | allergic fungal rhinosinusitis |
| 6MP | 6-Mercaptopurine | AgNOR | silver staining nucleolar organizer region |
| 18-FDG | 2-18-fluoro-2-deoxy-D-glucose | AHA | American Heart Association |
| A | adenine; or anterior | AHCPR | Agency for Health Care Policy and Research (USA) |
| AABR | automated auditory brainstem response | AHI | apnoea/hypopnoea index |
| AAHL | age-associated hearing loss | AI | apoptotic index |
| AAOHS | American Academy of Otolaryngologists/Head and Neck Surgeons | AICA | anterior inferior cerebellar artery |
| AAV | adeno-associated virus | AIDS | acquired immunodeficiency syndrome |
| ABC | aspiration biopsy cytology | AIRE | autoimmune regulator gene |
| ABEP | auditory nerve and brainstem evoked potential | AJCC | American Joint Committee on Cancer |
| ABG | air-bone gap | ALD | assistive listening device |
| ABI | auditory brainstem implant | ALEP | auditory long-latency (or late) evoked potential |
| ABLB | alternate binaural loudness balance | ALL | acute lymphoblastic leukaemia |
| ABPA | allergic bronchopulmonary aspergillosis | $\alpha 2\beta 2$ | two α and two β globin chains |
| ABR | auditory brainstem response; or acoustic brainstem evoked response | $\alpha 2\delta 2$ | HbA2 |
| ABRS | acute bacterial rhinosinusitis | $\alpha 2\gamma 2$ | foetal haemoglobin |
| AC | air conduction; or alternating coupled | ALPS | autoimmune lymphoproliferative syndrome |
| ACC | adenoid cystic carcinoma; or American College of Cardiology | ALS | amyotrophic lateral sclerosis |
| ACE | angiotensin-converting enzyme | ALT | alternative lengthening of telomere; or alternating chemoradiotherapy |
| ACF | anterior cranial fossa | ALTB | acute laryngotracheobronchitis |
| ACh | Acetylcholine | ALTE | apparent life-threatening event |
| AChR | acetyl choline receptor | AML | acute myeloid leukaemia |
| ACT | Aid for Children with Tracheostomies | AN | acoustic neuroma; or auditory neuropathy; or audiovestibular nerve |
| ACTH | adrenocorticotrophic hormone | ANA | anti-nuclear antibody |
| A/D | analogue-to-digital | AN/AD | auditory neuropathy/auditory dyssynchrony |
| AD | Alzheimer's disease | ANCA | antineutrophil cytoplasmic antibody |
| ADA | adenosine deaminase | AND | allow a natural death |
| ADAM-33 | A disintegrin and metalloprotease 33k | ANUG | acute necrotizing ulcerative gingivitis |
| ADCC | antibody-dependent cellular cytotoxicity | AOAE | automated otoacoustic emission |
| ADH | antidiuretic hormone | AoCD | anaemia of chronic disease |
| ADHD | attention deficit hyperactivity disorder | AOM | acute otitis media |
| ADR | adverse drug reaction | | |

| | | | |
|--------|---|----------|--|
| AON | anterior olfactory nucleus | BAHNO | British Association of Head and Neck Oncologists |
| AP | anterior–posterior; or action potential | BAO-HNS | British Association of Otorhinolaryngologists – Head and Neck Surgeons |
| APB | ALT-associated promyelocytic leukaemia body | BAPO | British Association for Paediatric Otolaryngology |
| APC | antigen presenting cell; or activated protein C; or argon plasma coagulation; or adenomatous polyposis coli | BCC | basal cell carcinoma |
| APD | auditory processing disorder | BCG | Bacillus Calmette–Guérin |
| APECED | autoimmune polyendocrinopathy–candidiasis–ectodermal dystrophy | BCHA | bone conductor hearing aid |
| APHAB | Abbreviated Profile of Hearing Aid Benefit | BCSH | British Committee for Standards in Haematology |
| APL | anti-phospholipid | BDP | beclomethasone dipropionate |
| APMET | aggressive papillary middle ear tumour | BE | bulla ethmoidalis |
| APQ | amplitude perturbation quotient | BF | biofeedback |
| APTT | activated partial thromboplastin time | BFU-E | burst-forming unit erythroid |
| APUD | amine precursor uptake and decarboxylation | BiPAP | bilevel positive airway pressure |
| ARF | acute renal failure | BIPP | bismuth and iodoform paraffin paste |
| ARIA | allergic rhinitis and its impact on asthma | BL | Burkitt's lymphoma |
| ARR | absolute risk reduction | BMA | British Medical Association |
| ARS | acute rhinosinusitis | BMI | body mass index |
| ART | advanced rotating tomograph; or antiretroviral therapy | BMP | bone morphogenetic protein; or bone morphogenic protein |
| ARTA | age-related typical audiogram | BMS | burning mouth syndrome |
| ASA | aspirin-induced asthma; or aspirin-sensitive asthma; or American Society of Anesthesiologists | BMT/SCT | bone marrow stem cell transplantation |
| a-SCC | anterior semicircular canal | BOA | behavioural observation audiometry |
| ASIC | application specific integrated circuit | BOLD | blood oxygenation level-dependent |
| ASL | American sign language; or arterial spin labelling | BOR | brachio-oto-renal |
| ASPO | American Society of Pediatric Otolaryngologists | BP | blood pressure |
| ASSR | auditory steady state response | BPD | bronchopulmonary dysplasia |
| AST | arterial spin tagging | BPPV | benign positional paroxysmal vertigo |
| AT | ataxia telangiectasia; or auditory therapy or training | BPV | benign paroxysmal vertigo; or benign positional vertigo |
| ATD | ascending tract of Deiters | BS | Behçet's syndrome |
| ATIII | antithrombin III | BSE | bedside swallowing examination; or bovine spongiform encephalopathy |
| ATN | auriculotemporal nerve | BSL | British sign language |
| ATP | adenosine triphosphate | BTE | behind the ear |
| ATRA | all-trans retinoic acid | BVF | bilateral vestibular failure |
| AUC | area under the curve | C | cytosine |
| AV | apical vesicles; or arteriovenous | CAD | caspase-activated DNase |
| AVCN | anteroventral cochlear nuclei | CADCAM | computer-aided design, computer-aided manufacture |
| AVM | arteriovenous malformation | CAGE | cerebral air gas embolism |
| aVOR | angular VOR | cAMP | 3',5'-monophosphate |
| AZT | 3'azido3'-deoxythymidone zidovudine; or azothiaprine | CANS | central auditory nervous system |
| | | CAP | compound action potential; or category of auditory performance |
| | | CAPD | central auditory processing disorder |
| BAC | bacterial artificial chromosome | CaR | calcium sensing receptor |
| BACDA | British Association of Community Doctors in Audiology | CAS | computer-assisted surgery |
| BADS | British Association of Day Surgery | CATCH-22 | cardiac defects, abnormal facies, thymic hypoplasia, cleft palate and hypocalcaemia-22 |
| BAES | British Association of Endocrine Surgeons | CB | concha bullosa; or critical bandwidth |
| BAHA | bone-anchored hearing aid | CBF | ciliary beat frequency |

| | | | |
|--------|--|-----------------|---|
| CBT | cognitive-behavioural therapy | CNO | chronic nasal obstruction |
| CCA | common carotid artery | CNS | central nervous system |
| CCDU | colour-coded duplex ultrasonography | CO ₂ | carbon dioxide |
| CCR | chemokine receptor | COAD | chronic obstructive airway disease |
| CCW | counter-clockwise | COM | chronic otitis media |
| CD | cluster of differentiation; or colloid droplets; or compact disk | COPD | chronic obstructive pulmonary disease |
| CDA | cold dry air | COR | conditioned orientation reflex |
| CDC | Centers for Disease Control and Prevention | COSI | Client Oriented Scale of Improvement |
| CDK | cyclin-dependent kinase | COX-2 | cyclo-oxygenase 2 |
| CDP | computerized dynamic posturography | CP | cleft palate |
| CE-CT | contrast-enhanced computed tomography | CPA | cerebellopontine angle |
| CEA | carcinoembryonic antigen | CPAP | continuous positive airway pressure |
| CEPOD | Confidential Enquiry into Perioperative Deaths | CPD | citrate phosphate dextrose; or continuing professional development |
| CER | control event rate | CPG | central pattern generator |
| CERA | cortical evoked response audiometry | CPO | cleft palate only |
| CEVMP | click-evoked vestibular myogenic potential | CPPIH | Commission for Patient and Public Involvement in Health (UK) |
| CF | cystic fibrosis; or characteristic frequency | CPR | cardiopulmonary resuscitation |
| CFD | colour-flow duplex Doppler | CQI | continuous quality improvement |
| CFR | craniofacial resection | CREST | calcinosis, Raynaud's, oesophageal involvement, sclerodactyly, telangiectasis |
| CFTR | cystic fibrosis transmembrane conductance regulator | CRF | corticotrophin-releasing factor |
| CFU | colony-forming unit | CRH | corticotropin-releasing hormone |
| CFU-GM | colony-forming unit, granulocyte-macrophage | CROS | contralateral routing of signal or sound |
| CFU-Mk | colony-forming unit, megakaryocyte | CRP | C-reactive protein; or canalith repositioning procedure |
| CG | clinical governance | CRRT | continuous renal replacement therapy |
| CGD | chronic granulomatous disease | CRS | chronic rhinosinusitis; or congenital rubella syndrome |
| CGH | comparative genomic hybridization | CRSS | chronic rhinosinusitis |
| CGRP | calcitonin gene-related peptide | CS | corticosteroid |
| CGST | Clinical Governance Support Team | CSCI | Commission for Social Care Inspection (UK) |
| CHARGE | coloboma, heart defects, atresia choanae, retardation of growth, genital anomalies and ear abnormalities | CSF | cerebrospinal fluid |
| CHART | continuous, hyperfractionated, accelerated radiotherapy | CSM | Committee on Safety of Medicines |
| CHI | Commission for Healthcare Improvement (UK) | CSOM | chronic suppurative otitis media |
| CI | cochlear implant; or cardiac index; or confidence interval; or concha inferior | CT | computed tomography |
| CID | Central Institute for the Deaf | CTA | composite tissue allograft |
| CJD | Creutzfeldt–Jakob disease | CTL | cytotoxic T-lymphocyte |
| CL | cleft lip | CTLA | cytotoxic T-lymphocyte-associated antigen |
| CL/P | cleft lip with or without cleft palate | CTLL | cytotoxic T-lymphocyte leukaemic |
| CLL | chronic lymphatic leukaemia; or chronic lymphocytic leukaemia | CTM | cricothyroid membrane |
| CM | concha media; or cochlear microphonic; or cricothyroid muscle | cTNM | clinical tumour, nodes, metastases |
| CMAP | compound muscle action potential | CTR | cricotracheal resection |
| CME | continuing medical education | CTZ | chemoreceptor trigger zone |
| CMI | cell-mediated immunity | Cu-ATSM | Cu(II)-diacetyl-bis-N4-methylthiosemicarbozone |
| CML | chronic myeloid leukaemia | CUP | carcinoma of unknown primary origin |
| CMT | Charcot–Marie–Tooth | CUSA | cavitation ultrasonic surgical aspirator |
| CMV | <i>Cytomegalovirus</i> | CVA | cerebrovascular accident |
| CN | cranial nerve; or cochlear nuclei; or cochlear nerve | CVD | central vestibular disorder |
| | | CVI | common variable immunodeficiency |
| | | CVP | central venous pressure |
| | | CW | clockwise |
| | | CXR | chest x-ray |
| | | CYP | cytochrome P450 |

| | | | |
|---------|---|--------|--|
| DACH | diaminocyclohexane | EAC | external auditory canal; or external acoustic canal |
| DAHANCA | Danish Head and Neck Cancer Study | EAL | ethmoidal artery ligation |
| DAHNO | Data for Head and Neck Oncology (UK) | EAM | external auditory meatus |
| dB | decibel | EB | epidermolysis bullosa |
| dB SPL | decibel sound pressure level | EBM | evidence-based medicine |
| DBPCFC | double-blind placebo-controlled food challenge | EBNA | Epstein-Barr virus-associated nuclear antigen |
| DCIA | deep circumflex iliac artery | EBP | evidence-based practice |
| DCN | dorsal cochlear nucleus | EBV | Epstein-Barr virus |
| DCR | dacryocystorhinostomy | EC | embryonic carcinoma |
| DD | death domain | ECA | external carotid artery |
| DDHS | Direct Drive Hearing System | ECAL | external carotid artery ligation |
| DFF | DNA fragmentation factor | ECAP | electrically evoked compound action potential |
| DFN3 | deafness type 3 | ECC | extracorporeal circuit |
| DFO-H | deferroxamine-hespan | ECG | electrocardiogram |
| DHA-S | dehydroepiandrosterone sulphate | ECM | extracellular matrix |
| DHE | dihaematoporphyrinether | ECMO | extracorporeal membrane oxygenation |
| DHI | dizziness handicap inventory | EcochG | electrocochleography |
| DHTR | delayed haemolytic transfusion reaction | ECog | electrocochleogram |
| DIC | disseminated intravascular coagulation | ECOG | Eastern Cooperative Oncology Group (USA) |
| DIEP | deep inferior epigastric perforator | ECP | eosinophil cationic protein |
| DILS | diffuse infiltrated lymphocytosis syndrome | ECR | extracapsular rupture |
| DIT | diiodotyrosine | EDGT | early goal-directed therapy |
| DLE | discoid lupus erythematosus | EDN | eosinophil-derived neurotoxin |
| DM | diabetes mellitus | EDS | excessive daytime sleepiness |
| DMD | Duchenne muscular dystrophy | EDTA | ethylenediaminetetraacetic acid |
| DMSA | dimercapto succinic acid | EDV | end diastolic velocity |
| DMSO | dimethylsulfoxide | EE | external frontoethmoidectomy |
| DNA | deoxyribonucleic acid | EEG | electroencephalography; or electroencephalogram |
| DNAR | do not attempt resuscitation | EER | experimental event rate |
| DNL | nasolacrimal duct | EFS | event-free survival |
| DNR | do not resuscitate | EG | embryonic germ |
| dNTP | deoxynucleoside triphosphate | EGF | epidermal growth factor |
| DP | directional preponderance | EGFR | epidermal growth factor receptor |
| DPA | Data Protection Act (UK) | EIA | enzyme immunoassay |
| DPOAE | distortion product otoacoustic emission | ELDCR | endonasal laser dacryocystorhinostomy |
| DR | death receptor; or drug resistance | ELG | electrolaryngography |
| DRS | Dysphagia Research Society | ELISA | enzyme-linked immunosorbent assay |
| DSA | digital subtraction angiography | ELST | endolymphatic sac tumour |
| DSI | Dysphonia Symptom Index | EM | erythema multiforme |
| DSL | desired sensation level | EMEA | European Agency for the Evaluation of Medicinal Products |
| DTD | DT-diaphorase | EMG | electromyography |
| DTIC | dimethyl triazeno imidazole carboxamide | EMI | elective mucosal irradiation |
| dTMP | deoxythymidine monophosphate | EN | enteral nutrition |
| DTPA | diethylene triamine pentacetic acid | ENA | extra nuclear antigen |
| dUMP | deoxyuridine monophosphate | ENG | electronystagmography |
| DVB | degree of voice break | ENoG | electroneurography |
| DVLA | Driver and Vehicle Licensing Authority (UK) | ENT | ear, nose and throat |
| DVN | descending vestibular nuclei | EOG | electroolfactogram; or electrooculography |
| DVT | deep vein thrombosis | EORTC | European Organisation for Research and Treatment of Cancer |
| DWI | diffusion weighted image | EP | endolymphatic potential |
| EA | episodic ataxia; or early antigen | | |
| EAACI | European Academy of Allergology and Clinical Immunology | | |

| | | | |
|----------------|---|-----------|--|
| EPO | erythropoietin | FN | facial nerve |
| EQ-5D | EuroQol | FNA | fine-needle aspiration |
| ER | enhancement ratio; or endoplasmic reticulum | FNAB | fine-needle aspiration biopsy |
| ERB | equivalent rectangular bandwidth | FNAC | fine-needle aspiration cytology |
| ERM | ezrin, radixin, moesin | FOAR | fronto-orbital advancement and remodelling |
| ERP | event-related potential | FOI | fibreoptic orotracheal intubation |
| ERT | external radiotherapy | FPANS | fluticasone propionate aqueous nasal spray |
| Er:YAG | erbium:yttrium-aluminium-garnet | FS | folliculostellate |
| ES | embryonic stem; or endolymphatic sac | FSH | follicle-stimulating hormone |
| ESPAL | endonasal ligation of the sphenopalatine artery | FT | fibrous tissue |
| ESR | erythrocyte sedimentation rate | FTA | fluorescent treponemal antibody |
| ESS | endoscopic sinus surgery; or Epworth Sleepiness Scale | FTA-ABS | fluorescent treponemal antibody test |
| ET | essential thrombocytosis; or endotracheal tube | FTC | frequency threshold curve |
| ET-1 | endothelin-1 | FTP | Fitness to Practise |
| ETT | endotracheal tube | G | guanine |
| EU | European Union | G6PD | glucose-6-phosphate deficiency |
| EUA | examination under anaesthesia | Ga-67 | gallium |
| EVAS | enlarged vestibular aqueduct syndrome | GABA | gamma-aminobutyric acid |
| EXIT | extrauterine intrapartum treatment | GABHS | group A beta-haemolytic streptococcus |
| F ₀ | fundamental frequency | GAG | glycosaminoglycan |
| FAAF | four alternative auditory feature | GALT | gut-associated lymphoid tissue |
| Fab | fragment antigen binding | GAS | Goal Attainment Scaling |
| FACS | fluorescence-activated cell sorter | G&S | group and screen |
| FACT | functional assessment of cancer therapy | GBI | Glasgow Benefit Inventory |
| FAMM | facial artery myomucosal flap | GBLC | geometric broken line closure |
| Fas-L | Fas ligand | GCS | Glasgow Coma Score |
| FBC | full blood count | G-CSF | granulocyte-colony stimulating factor |
| Fc | fragment crystallizable | GD | Graves' disease |
| FD | fibrous dysplasia | GERD | gastrooesophageal reflux disease |
| FDA | Food and Drug Administration (USA) | GH | growth hormone |
| FDG | fluorodeoxyglucose; or 2-[¹⁸ F] fluoro-2-deoxy-D-glucose; or F18-fluoro-2-deoxy-D-glucose | GHABP | Glasgow Hearing Aid Benefit Profile |
| FDG-PET | 2-[¹⁸ F] fluoro-2-deoxy-D-glucose-positron emission tomography; or fluorine-18-labelled deoxyglucose positron emission tomography | GHRH | growth hormone-releasing hormone |
| FEES | fibreoptic endoscopic evaluation of swallowing | GI | gastrointestinal |
| FEESST | fibreoptic endoscopic evaluation of swallowing with sensory testing | GIA | gravitoinertial acceleration |
| FESS | functional endoscopic sinus surgery | GIC | glass ionomer cement |
| FETNIM | fluorine-18 fluoroerythronitroimidazole | GIST | gastrointestinal stromal tumour |
| FFP | fresh frozen plasma | GLM | ground lamella of middle turbinate, middle (frontal) portion |
| FFT | fast Fourier transform | GMC | ganglion mother cell; or General Medical Council (UK) |
| FGF | fibroblast growth factor | GM-CSF | granulocyte-macrophage colony-stimulating factor |
| FHH | familial hypocalcaemic hypercalcaemia | GN | glossopharyngeal nerve |
| FISH | fluorescence <i>in situ</i> hybridization | GNE | glottal-to-noise excitation |
| FIV | feline immunodeficiency virus | GnRH | gonadotropin-releasing hormone |
| FLAIR | fluid attenuated inversion recovery | GOR | gastro-oesophageal reflux |
| FMISO | fluorine-18 fluoromisonidazole | GORD | gastro-oesophageal reflux disease |
| fMRI | functional magnetic resonance imaging | GOSH | Great Ormond Street Hospital (UK) |
| | | gp | glycoprotein |
| | | GP | general practitioner |
| | | GPN | glossopharyngeal neuralgia |
| | | GPP | gingivo-periosteoplasty |
| | | G protein | guanine nucleotide-binding regulatory protein |
| | | GPB2 | growth factor receptor binding protein 2 |

| | | | |
|---------|--|---------------|---|
| GSH | glutathione | HMWC | high molecular weight compound |
| GSPN | greater superficial petrosal nerve | HNC | head and neck cancer |
| GST | glutathione S-transferase | HNR | harmonics-to-noise ratio |
| GSW | gun shot wound | HNRQ | Head and Neck Radiotherapy Questionnaire |
| GTN | nitroglycerin | HNSCC | head and neck squamous cell carcinoma |
| GTR | guided tissue regeneration | HPA | hypothalamic–pituitary–adrenal |
| GVHD | graft-versus-host disease | HPC | haemangiopericytoma |
| H&E | haematoxylin and eosin | HPD | haematoporphyrin derivative |
| H&N | head and neck | HPL | horizontal partial laryngectomy |
| H2 | histamine receptor type 2 | HPT | hyperparathyroidism |
| HA | hydroxyapatite | HPV | human papillomavirus; or human herpes virus 8 |
| HAART | highly active antiretroviral therapy | HRA | Human Rights Act |
| HAE | hereditary angioedema | HRCT | high-resolution computed tomography |
| HAEM | HSV-associated erythema multiforme | HRM | high-resolution manometry |
| HAPI | Hearing Aid Performance Inventory | HRQOL | health-related quality of life |
| HB | House–Brackmann | HRT | hormone replacement therapy |
| Hb | haemoglobin | HS | hiatus semilunaris |
| HbA | adult haemoglobin | h-SCC | horizontal semicircular canal |
| HBO | hyperbaric oxygen | HSCT | haemopoietic stem cell transplant |
| HBOT | hyperbaric oxygen therapy | HSMN | hereditary sensory-motor neuropathy |
| HBsAg | hepatitis B surface antigen | HSPG | heparin sulphate proteoglycan |
| HC | Healthcare Commission (UK) | HSV | herpes simplex virus |
| HCA | hydroxycarbonate apatite | HSV-1 | herpes simplex virus type 1 |
| HCG | human chorionic gonadotrophin | HSV-2 | herpes simplex virus type 2 |
| HCSU | Health Care Standards Unit (UK) | HSV-TK | herpes simplex thymidine kinase |
| Hct | haematocrit | HT | hydroxytryptamine |
| HCV | hepatitis C virus; or human T-lymphocytic virus 1 | hTERT | human telomerase reverse transcriptase |
| HD | haemodialysis | hTR | human telomerase RNA |
| HDL | high-density lipoprotein | HU | Hounsfield unit |
| HDM | house dust mite | HUI | Health Utilities Index |
| HDPE | high-density polyethylene | HUS | haemolytic uraemic syndrome |
| HDU | high dependency unit | Hz | hertz |
| He-Ne | helium-neon | HZV | herpes zoster virus |
| HEp-2 | human epithelial type 2 | IAC | internal auditory canal |
| HFT | hereditary familial telangiectasia | IAM | internal auditory meatus |
| HGF | hepatocyte growth factor | IBP | invasive monitoring of blood pressure |
| HHI | Hearing Handicap Inventory | IC | inferior colliculus; or immunochemistry |
| HHIE | Hearing Handicap Inventory for the Elderly | ICA | internal carotid artery |
| HHT | hereditary haemorrhagic telangiectasia | ICAM | intercellular adhesion molecule |
| HHV-6 | human herpesvirus 6 | ICAM-1 | intercellular adhesion molecule 1 |
| HHV-8 | human herpesvirus 8 | ICD | International Classification of Disease |
| HI | hearing impaired | ICM | intensive care medicine |
| HiB | <i>Haemophilus influenzae</i> B | ICP | intracranial pressure |
| HIT | heparin-induced thrombocytopenia | ICRA | International Collegium of Rehabilitative Audiology |
| HITT | heparin-induced thrombocytopenia with thrombosis | ICU | intensive care unit |
| HIV | human immunodeficiency virus | ID | inferior dental |
| HIV-SGD | HIV-associated salivary gland disease | IDA | iron deficiency anaemia |
| HJB | high jugular bulb | IDT | infant distraction test |
| HL | hearing loss; or hearing level; or hairy leukoplakia | IDU | intravenous drug user |
| HLA | human leukocyte antigen | IF | intrinsic factor |
| HM | history of migraine; or hemifacial microsomia | IFN | interferon |
| HMW | high molecular weight | IFN- α | interferon-alpha |
| | | IFN- β | interferon-beta |
| | | IFN- γ | interferon gamma |

| | | | |
|-----------------|--|--------|---|
| IFNP | idiopathic facial nerve paralysis | K | Kirschner |
| Ig | immunoglobulin | KAR | killer activating receptor |
| IgE | immunoglobulin E | keV | kilo electron volt |
| IGF | insulin-like growth factor | KIR | killer inhibitory receptor |
| IGFI | insulin-like growth factor I | KS | Kaposi's sarcoma |
| IGFII | insulin-like growth factor II | KSS | Kearns-Sayre syndrome |
| IgG | immunoglobulin G | KTP | potassium titanyl phosphate |
| IGS | image-guided surgery | | |
| IHAFF | International Hearing Aid Fitting Forum | LA | lymphangioma |
| IHC | immunohistochemistry; or inner hair cell | LAD | leukocyte adhesion defect |
| | | LAP | left anteroposterior |
| IHS | International Headache Society | LARP | left anterior-right posterior |
| IL | interleukin | LAUP | laser-assisted uvulopalatoplasty |
| IL-1 | interleukin-1 | LB | lateral bundle |
| IL-2 | interleukin-2 | LCH | Langerhans' cell histiocytosis |
| IL-3 | interleukin-3 | LCM | laser capture microdissection |
| IL-6 | interleukin-6 | LD | lymphocytic depleted |
| ILMA | intubating laryngeal mask airway | LDH | lactate dehydrogenase |
| IMA | internal maxillary artery | LDL | low-density lipoprotein; or loudness discomfort level |
| IMAL | internal maxillary artery ligation | | |
| IMF | intermaxillary fixation | LDUH | low-dose unfractionated heparin |
| IMRT | intensity-modulated radiation therapy | LED | light-emitting diode |
| IMSPAC | imitative test of speech pattern contrast perception | LFA | lymphocyte-function associated antigen |
| | | LGOB | loudness growth in octave bands |
| INC | immunonuclear chemistry | LH | luteinizing hormone |
| INE | intranasal ethmoidectomy | LHRH | leuteinizing hormone-releasing hormone |
| INO | internuclear ophthalmoplegia | LIF | leukaemia-inhibitory factor |
| iNOS | inducible nitric oxide synthase | LINKs | Local Involvement Networks (UK) |
| INR | international normalized ratio; or interventional neuroradiology | LL | lateral lemniscus |
| | | LM | laryngeal mask |
| IOC | Interim Orders Committee (UK) | LMA | laryngeal mask airway |
| IOPI | Iowa Oral Performance Instrument | LMN | lower motor neuron |
| IP ₃ | 1,4,5-inositol triphosphate | LMW | low molecular weight |
| IPSS | inferior petrosal sinus sampling | LMWC | low molecular weight compound |
| IRMA | immunoradiometric assay | LMWH | low molecular weight heparin |
| IRS | Intergroup Rhabdomyosarcoma Study | LOD | logarithm to the base 10 of the odds that the markers are linked at a recombination distance of <i>N</i> centimorgans |
| ISAAC | International Study of Asthma and Allergies in Childhood | | |
| ISEL | <i>in situ</i> end labeling | LOH | loss of heterozygosity |
| ISJ | incudostapedial joint | LOS | length of stay; or lower oesophageal sphincter |
| ISO | International Standards Organization | | |
| ISS | immunostimulatory DNA sequence | LP | lamina papyracea; or lichen planus; or lymphocyte predominant |
| ISSNHL | idiopathic sudden sensorineural hearing loss | LPC | linear predictive coding |
| | | LPR | laryngopharyngeal reflux |
| IT | inferior turbinate | LR | likelihood ratio |
| ITA | inferior thyroid artery | LREC | local research ethics committee |
| ITE | in the ear | LSCC | lateral semicircular canal |
| ITP | idiopathic thrombocytopenic purpura | IT | leukotriene |
| ITU | intensive therapy unit | LTAS | long-term average spectrum |
| IUCC | International Union against Cancer | LTASS | long-term average speech spectrum |
| i.v. | intravenous | LTB | laryngotracheobronchitis; or laryngotracheobronchoscopy |
| IVIg | intravenous immunoglobulin | | |
| | | LTC4-S | leukotriene C4 synthase |
| JFC | just-follow-conversation | LTR | laryngotracheal reconstruction |
| JNA | juvenile nasopharyngeal angiofibroma | LTRA | leukotriene receptor antagonists |
| JORPP | juvenile-onset recurrent respiratory papillomatosis | LVA | large vestibular aqueduct |

| | | | |
|--------|---|----------------|--|
| LVAS | large vestibular aqueduct syndrome | MIDD | maternally inherited diabetes and deafness |
| LVN | lateral vestibular nuclei | MIP | minimally invasive open parathyroidectomy; or maximum intensity projection; or macrophage inflammatory protein |
| LVOR | linear vestibulo-ocular reflex | | |
| M | metastases | | |
| MAb | monoclonal antibodies | MIP-1 α | macrophage inflammatory protein-1 α |
| MABP | mean arterial blood pressure | MISS | minimally invasive sinus surgery |
| MAC | membrane attack complex; or <i>Mycobacterium avium</i> complex | MIT | monoiodotyrosine |
| MACS | magnetic-activated cell sorter; or minimal access cranial suspension | MIVAP | minimally invasive video-assisted parathyroidectomy |
| MAF | minimum audible field | ML | mixed cellularity |
| MALT | mucosa-associated lymphoid tissue | MLD | masking level difference |
| MAOI | monoamine oxidase inhibitor | MLF | medial longitudinal fascicle or fasciculus |
| MAP | minimum audible pressure | MLR | middle latency response |
| MAPK | mitogen-activated protein kinase | MLTB | microlaryngotracheobronchoscopy |
| MAS | mandibular advancement splint | MM | malignant melanoma |
| MB | medial bundle | MMC | mitomycin C |
| MBL | mannose-binding lectin | MMN | mismatch negativity |
| MBP | major basic protein | MMP | mucous membrane pemphigoid; or matrix-metalloprotease |
| MBS | modified barium swallow | MMR | measles, mumps and rubella |
| MCP | monocyte chemotactic protein | MMS | Moh's micrographic surgery |
| MCP-1 | monocyte chemotactic protein-1 | MND | motor neurone disease |
| MCS | mental component summary | MNG | multinodular goitre |
| M-CSF | macrophage-colony stimulating factor | MOC | medial olivocochlear |
| MCV | mean corpuscular volume | MODS | multiple organ dysfunction syndrome |
| MDC | macrophage derived chemokine | MOT | malignant odontogenic tumour |
| MDR | multiple drug resistance | MPA | microscopic polyangiitis |
| MDRTB | multidrug resistant tuberculosis | MPL | monophosphoryl lipid A |
| MDS | myelodysplastic syndrome | MPO | myeloperoxidase |
| MDT | multidisciplinary team | MPT | maximum phonation time |
| MDVP | Multidimensional Voice Program | MPTP | 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine |
| ME | middle ear | | |
| MEE | medial edge epithelium | MR | magnetic resonance |
| MEG | magnetoencephalography | MRA | magnetic resonance angiography |
| MEK | MAPK/extracellular signal related kinase | MRC | Medical Research Council (UK) |
| MELAS | mitochondrial encephalopathy, lactic acidosis and stroke-like episode | MREC | multicentre regional ethics committee |
| MEMS | microelectromechanical system | MRI | magnetic resonance imaging |
| MEN | multiple endocrine neoplasia | MRL | minimal response level |
| MERRF | myoclonic epilepsy and ragged red fibre | mRNA | messenger ribonucleic acid |
| MeSH | medical subject heading | MRND | modified radical neck dissection |
| MESS | microscopic endonasal sinus surgery | MRS | Melkersson-Rosenthal syndrome; or magnetic resonance sialography |
| MET | middle ear transducer | MRSA | methicillin-resistant <i>Staphylococcus aureus</i> |
| MF | middle fossa | MRV | migraine-related vestibulopathy |
| M-FISH | multifluor FISH | MS | multiple sclerosis |
| MFR | mean airflow rate | MSA | multiple systems atrophy |
| MGB | medial geniculate body | MSBOS | maximum surgical blood ordering schedule |
| MGSA | melanoma growth stimulating activity | MSG | monosodium glutamate |
| MGUS | monoclonal gammopathy of uncertain significance | MST | maximal stimulation test |
| MHC | major histocompatibility complex | MT | maxilloturbinal; or middle turbinate |
| MI | myocardial infarction | MTC | medullary thyroid carcinoma |
| MIBG | metaiodobenzylguanidine; or iodine-123-metaiodobenzylguanidine | MTD | muscle tension dysphonia |
| MIBI | sestamibi; or technetium-99m | mtDNA | mitochondrial DNA |
| MIC | minimum inhibitory concentration | MTHFR | methylenetetrahydrofolate reductase |
| | | mTHPC | meso-tetra (hydroxyphenyl) chlorin |
| | | MUS | medically unexplained symptom |

| | | | |
|------------------|--|-----------------|---|
| MV | mechanical ventilation | NMCC | nasal mucociliary clearance |
| MVN | medial vestibular nuclei | NMDA | N-methyl-d-aspartate; or National Minimum Data Set (UK) |
| N | nodal | NNE | normalized noise energy |
| NA | noradrenaline | NNT | number needed to treat |
| NADP | nicotinamide adenine dinucleotide phosphate | NO | nitric oxide |
| NADPH | reduced form of nicotinamide adenine dinucleotide phosphate | NO ₂ | nitric dioxide |
| NAL | National Acoustic Laboratories (Australia) | NOE | naso-orbito-ethmoid |
| NAMCS | National Ambulatory Medical Care Survey (USA) | non-REM | nonrapid eye movement sleep |
| NANIPER | nonallergic noninfectious perennial rhinitis | NOS | not otherwise specified |
| NARES | nonallergic rhinitis with eosinophilia syndrome | NP | nasopharynx; or nasopharyngeal |
| NATA | National Anonymous Tonsil Archive | NPC | nasopharyngeal cancer; or nasopharyngeal carcinoma |
| NBCA | n-butyl-2-cyanoacrylate; or N-butyl-cyanoacrylate | NPSA | National Patient Safety Agency (UK) |
| NBT | nitro blue tetrazolium | NPTA | National Prospective Tonsillectomy Audit (UK) |
| NCAA | National Clinical Assessment Authority (UK) | NPV | negative predictive value |
| NCAS | National Clinical Assessment Service (UK) | NPY | neuropeptide Y |
| NCASP | National Clinical Audit Support Programme (UK) | NRA | nucleus retroambigualis |
| NCCG | non-consultant career-grade | NRLS | National Reporting and Learning System (UK) |
| NCCN | National Comprehensive Cancer Network | NRT | neural response telemetry |
| NCDB | National Cancer Data Base (USA) | NS | nodular sclerosing |
| NCEPOD | National Confidential Enquiry into Patient Outcome Death (UK) | NSAID | nonsteroidal antiinflammatory drug |
| NCIC | National Cancer Institute of Canada | NSCAG | National Specialist Commissioning Advisory Group (UK) |
| NEET | nose, ear, eye and temple | NSF | national service framework |
| NESSTAC | North of England and Scotland Study on Tonsillectomy and Adenoidectomy in Children | NSHPT | neonatal severe hyperparathyroidism |
| NET | nerve excitability test | NSRAN | nonsyndromic recessive auditory neuropathy |
| NFκB | nuclear factor kappa B | NT | nasoturbinal |
| NF1 | neurofibromatosis type 1 | NTD | neural tube defect |
| NF2 | neurofibromatosis type 2 | NTM | non-tuberculous mycobacteria |
| NFA | nonfunctioning pituitary adenomas; or nasofrontal approach | NTS | nucleus tractus solitarius |
| NG | nasogastric | NYHA | New York Heart Association |
| NH | normal hearing | O ₃ | ozone |
| NHL | non-Hodgkin's lymphoma | OAE | otoacoustic emission |
| NHS | National Health Service (UK) | OAN | olfactory neuroblastoma |
| NHSP | Newborn Hearing Screening Programme | OAS | oral allergy syndrome |
| NIBP | automatic noninvasive blood pressure | OB | olfactory bulb |
| NICE | National Institute for Health and Clinical Excellence (UK) | OCB | olivocochlear bundle |
| NICU | nonimmunological contact urticaria; or neonatal intensive care unit | OCFC | open controlled food challenge |
| NIDDM | noninsulin dependent diabetes mellitus | OCT | optical coherence tomography |
| NIH | National Institutes of Health (USA) | ODI | oxygen desaturation index |
| NIHL | noise-induced hearing loss | ODT | olfactory detection threshold |
| NIPF | nasal inspiratory peak flow | OEC | olfactory ensheathing cell |
| NIS | Na ⁺ /I ⁻ symporter | OFG | orofacial granulomatosis |
| NK | natural killer | OGTT | oral glucose tolerance test |
| N/m ² | Newtons/square metre | OHC | outer hair cell |
| | | OHL | oral hairy leukoplakia |
| | | OHS | obesity hypoventilation syndrome |
| | | OKN | optokinetic nystagmus |
| | | OM | occipitomenal |
| | | OMC | ostiomeatal complex |
| | | OME | otitis media with effusion |
| | | OMENS | orbit, mandible, ears, nerves and soft-tissue |

| | | | |
|------------|---|------------------|--|
| OMIM | Online Mendelian Inheritance in Man | PET | polyethylene terephthalate; or positron emission tomography |
| OPCS | Office for Population Censuses and Surveys (UK) | PET-CT | positron emission tomography/computed tomography |
| OPG | orthopantomogram | PF | posterior fontanelle; or cisplatinum/5-fluorouracil |
| OR | occupational rhinitis | PF4 | platelet factor 4 |
| OREP | olfactory event-related potential | PFAPA | periodic fever, aphthous stomatitis, pharyngitis and cervical adenitis |
| ORL | otorhinolaryngology | PFC | perfluorocarbon |
| OS | osteosarcoma | PFG | percutaneous fluoroscopic gastrostomy |
| OSA | obstructive sleep apnoea | PGA | polyglycolic acid |
| OSAH | obstructive sleep apnoea/hypopnoea | PGE ₁ | prostaglandin-E ₁ |
| OSAHS | obstructive sleep apnoea/hypopnoea syndrome | PGE ₂ | prostacycline; or prostaglandin I ₂ |
| OSAS | obstructive sleep apnoea syndrome | PGL | persistent generalized lymphadenopathy |
| OSC | overview and scrutiny committee | pHPT | primary hyperparathyroidism |
| OSPH | ostium of sphenoid sinus | PI | pulsatility index |
| OSPL | output sound pressure level | PI3-K | phosphatidylinositol 3 |
| OTOF | otoferlin | PICA | posterior inferior cerebellar artery |
| OVAR | off-vertical axis rotation | PICU | paediatric intensive care unit |
| P | phosphate; or posterior | PIF | prolactin release inhibiting factor |
| PA | pernicious anaemia | PIFR | peak inspiratory flow |
| PAC | P1 artificial chromosome; or pulmonary artery catheter | PIHA | partially implantable hearing aid |
| PAD | preoperative autologous deposit | PIII | parathyroid III |
| PAF | platelet-activating factor | PIV | parainfluenza virus; or parathyroid IV |
| PAG | periaqueductal grey matter | PIVC | parietoinsular vestibular cortex |
| PAI-1 | plasminogen activator inhibitor type 1 | PLA | polylactic acid |
| PALS | Patient Advice and Liaison Service (UK) | PLD | potentially lethal damage |
| PA-RT | partly accelerated radiotherapy | PLF | congenital perilymphatic fistula |
| PAS | periodic acid-Schiff | PLG | polylactide-coglycolide |
| PBP | progressive bulbar palsy | PLMD | periodic limb movement disorder |
| PCA | patient-controlled analgesia | PLS | primary lateral sclerosis |
| PCC | prothrombin complex concentrate; or Professional Conduct Committee (UK) | PM | particulate matter |
| PCD | primary ciliary dyskinesia | PMS | pharyngeal mucosal space |
| PCHI | permanent childhood hearing impairment | PNP | purine nucleoside phosphorylase; or paraneoplastic pemphigus |
| PCNA | proliferating cell nuclear antigen | PNS | peripheral nervous system; or postnasal space |
| PCR | polymerase chain reaction | POGO | prescription of gain and output |
| Pcrit | critical pressure | PONV | postoperative nausea/vomiting |
| PCS | physical component summary | PORP | partial ossicular replacement prosthesis |
| PCT | primary care trust | PP | pyrophosphate |
| PCTR | partial cricotracheal resection | PPC | Preliminary Proceedings Committee (UK) |
| PD | Parkinson's disease | PPD | purified protein derivative |
| PD-ECGF | platelet-derived endothelial cell growth factor | PPI | proton pump inhibitor; or patient and public involvement |
| PDGF | platelet-derived growth factor | PPRF | parapontine reticular formation; or paramedian pontine reticular formation |
| PDGFR | platelet-derived growth factor receptor | PPS | parapharyngeal space |
| PDL | pulsed dye laser | PPV | positive predictive value |
| PDR | <i>Physicians' Desk Reference</i> | PR3 | proteinase 3 |
| PDS | polydimethylsiloxane | PRCT | prospective randomized controlled trial |
| PDT | photodynamic therapy | PRL | prolactin |
| PE | polyethylene; or pulmonary embolism; or pharyngo-oesophageal | PRP | platelet-rich plasma |
| PEEP | positive-end expiratory pressure | PRPP | 5-phospho-alpha-D-ribose 1-diphosphate |
| PEG | percutaneous endoscopic gastrostomy | PRS | persistent rhinosinusitis |
| PEMA/THFMA | poly (ethylmethacrylate)/tetrahydrofurfuryl methacrylate | PRV | polycythaemia rubra vera |

| | | | |
|--------------|--|----------|--|
| PSA | prostate-specific antigen; or pleomorphic salivary adenoma; or persistent stapedial artery | REM | rapid eye movement |
| p-SCC | posterior semicircular canal | rEPO | recombinant erythropoietin |
| PSG | polysomnography | RET | rearranged during transfection |
| PS-OCT | polarization-sensitive OCT | RFS | rhinofrontal sinuseptomy |
| PSP | progressive supranuclear palsy | RFTVR | radiofrequency tissue volume reduction |
| PSV | peak systolic velocity | RFVR | radiofrequency volumetric reduction |
| PT | prothrombin time | RHD | Reported Hearing Disability |
| PTA | pure tone average; or peritonsillar abscess | RI | resistance index |
| PTC | psychophysical tuning curve | RIA | radioimmuno assay |
| PTFE | polytetrafluoroethylene | riMLF | rostral interstitial nucleus of the medial longitudinal fasciculus |
| PTH | parathyroid hormone | RLN | recurrent laryngeal nerve |
| PTHrP | parathyroid hormone-related protein; or parathyroid hormone-related peptide | RLS | restless leg syndrome |
| pTNM | pathological tumour, nodes, metastases | RMS | root mean square; or rhabdomyosarcoma |
| PTP | post-transfusion purpura | RNA | ribonucleic acid |
| PTS | permanent threshold shift | RND | radical neck dissection |
| PTU | propylthiouracil | RNID | Royal National Institute for Deaf and Hard of Hearing People (UK) |
| PU | uncinate process | RNP | ribonucleoprotein |
| PV | pemphigus vulgaris | ROC | receiver operating characteristic |
| PVA | polyvinyl alcohol | ROI | region of interest; or reactive oxygen intermediate |
| PVC | polyvinyl chloride | ROM | range of motion |
| PVCN | posteroventral cochlear nuclei | ROOF | retro-orbicularis orbital fat |
| PVP | pause vestibular position; or position vestibular pause | ROS | reactive oxygen species |
| PVS | persistent vegetative state | RP | rapid prototyping |
| PZT | lead zirconate titanate | RPA | retropharyngeal abscess |
| QALY | quality adjusted life year | RPT | rapid pull through |
| QOL | quality of life | RR | relative risk |
| QTL | quantitative trait loci | RRP | recurrent respiratory papillomatosis |
| RA | retinoic acid | RRR | relative risk reduction |
| RAE | Ring, Adair, Elwyn | RS | retrosgmoid |
| RAI | radioactive iodine | RSDI | Rhinosinusitis Disability Index |
| RALP | right anterior–left posterior | RSOM | rhinosinusitis outcome measure |
| RAM | Rahmonic amplitude | RSTL | relaxed skin tension line |
| RANTES | regulated on activation, normal T-cell expressed and secreted | RSV | respiratory syncytial virus |
| RAP | right anteroposterior | RT | radiotherapy |
| RAR α | retinoic acid receptor α gene | rT3 | reverse triiodothyronine |
| RARS | recurrent acute rhinosinusitis | RTK | receptor tyrosine kinase |
| RAS | recurrent aphthous stomatitis | RTL | right thyroid artery |
| RAST | radioallergosorbent test | rTMS | repetitive low-frequency transcranial magnetic stimulation |
| RAT | rapid antigen testing | RT-PCR | reverse transcriptase-polymerase chain reaction |
| RB | retinoblastoma | RUDS | reactive upper airways dysfunction syndrome |
| RBC | red blood cell | SACE | serum angiotensin converting enzyme |
| rCBF | regional cerebral blood flow | SAD | supraglottic airway device |
| RCPCH | Royal College of Paediatrics and Child Health | SAGM | saline-adenine-glucose-mannitol |
| RCT | randomized controlled trial | SALT | speech and language therapist |
| RDI | respiratory disturbance index | SANS | subacute necrotizing sialadenitis |
| REAG | real-ear aided gain | SAP | signalling lymphocyte activation molecule associated protein |
| REAL | Revised European American Lymphoma | SAPALDIA | Swiss Study on Air Pollution and Lung Diseases in Adults |
| RECD | real ear to coupler difference | SPE | side building syndrome |
| REIC | real ear insertion gain | | |

| | | | |
|-------------------|--|-----------------|---|
| s.c. | subcutaneous | SNOMED CT | Systematized Nomenclature of Medicine – Clinical Terms |
| SCBU | special care baby unit | SNOT | sino-nasal outcome test |
| SCC | squamous cell carcinoma or cancer; or semicircular canal | SNR | signal-to-noise ratio |
| SCCA | squamous cell carcinoma antigen | SNUC | sinonasal undifferentiated carcinoma |
| SCCHN | squamous cell carcinoma of the head and neck | SO ₂ | sulphur dioxide |
| SCD | sickle cell disease | SOAE | spontaneous otoacoustic emission |
| SCF | stem cell factor | SOC | superior olivary complex |
| SCID | severe combined immunodeficiency | SOM | secretory otitis media |
| SCN | severe congenital neutropenia | SOOF | suborbicularis oculi fat |
| SCUBA | self-contained underwater breathing apparatus | SOS | guanine nucleotide exchange factor (son of sevenless) |
| ScvO ₂ | central venous oxygen saturation | SP | substance P; or summing potential |
| SEAC | Spongiform Encephalopathy Advisory Committee | SPECT | single photon emission computed tomography |
| SEM | scanning electron microscopy | SPET | single photon emission tomography |
| sEMG | surface electromyography | SPF | sphenopalatine foramen |
| SF-36 | Medical Outcome Study Short-Form 36-Item Health Survey | SPI | soft phonation index |
| SfBH | Standards for Better Health (UK) | SPIO | superparamagnetic iron oxide |
| SFF | speaking fundamental frequency | SPL | sound pressure level |
| SFOAE | stimulus frequency otoacoustic emission | SPT | skin prick test; or station pull through |
| SGC | spiral ganglion cell | SRS | subacute rhinosinusitis |
| Shh | sonic hedgehog | SRS-A | slow reacting substance of anaphylaxis |
| SHO | senior house officer | SRT | speech recognition threshold; or speech reception threshold |
| SHOT | serious hazards of transfusion | SSC | superior semicircular canal |
| SIADH | syndrome of inappropriate antidiuretic hormone | SSEP | steady-state potential |
| SIDS | sudden infant death syndrome | SSG | split skin graft |
| sIg | surface immunoglobulin | SSLP | simple sequence length polymorphism |
| SIGN | Scottish Intercollegiate Guidelines Network | SSNHL | sudden sensorineural hearing loss |
| SIMEHD | semi-implantable middle ear electromagnetic hearing device | SSPE | subacute sclerosing panencephalitis |
| SIP | sickness impact profile | SSPL | saturation sound pressure level |
| SIR | speech intelligibility rating; or standardized incidence ratio | SSR | steady-state response |
| SIRS | systemic inflammatory response syndrome | SSRI | selective serotonin reuptake inhibitor |
| SL | sensation level | ST | superior turbinate |
| SLD | sublethal damage | STAT | signal transducer and activator of transcription |
| SLE | systemic lupus erythematosus | STD | standard deviation |
| SLIT | sublingual immunotherapy | STIR | short time inversion recovery |
| SLN | superior laryngeal nerve | STRP | short tandem repeat polymorphism |
| SLNB | sentinel lymph node biopsy | SUV | standardized uptake value |
| SLP | superficial lamina propria | SVCO | superior vena caval obstruction |
| SLT | speech and language therapist | SVL | strobovideolaryngoscopy |
| SMAS | superficial or subcutaneous musculoaponeurotic system | SVN | superior vestibular nuclei; or superior vestibular nerve |
| SMOFIT | submucous resection of the turbinate | SVV | subjective visual vertical |
| SMR | submucosal resection | SVZ | subventricular zone |
| SMS | short message service; or indium-111 pentetreotide | SWS | slow wave sleep |
| S/N | speech-to-noise | T | thymine; or tumour |
| SNC | sinonasal cancer | T1WI | T1-weighted images |
| SNHL | sensorineural hearing loss | T2WI | T2-weighted images |
| SNOMED | Systematized nomenclature of medicine | T3 | triiodothyronine |
| | | T4 | thyroxine |
| | | T/A | tonsillectomy and/or adenoidectomy |
| | | TAGVHD | transfusion-associated graft-versus-host disease |

| | | | |
|-----------------|---|------------------|--|
| TARC | thymus and activation-regulated chemokine | TRALI | transfusion-related acute lung injury |
| TARGET | Trial of Alternative Regimens in Glue Ear Treatment | TRAM | transverse rectus abdominis myocutaneous |
| TB | tuberculosis; or <i>Mycobacterium tuberculosis</i> | TRH | thyrotropin-releasing hormone |
| TBG | thyroxine-binding globulin | tRNA | transfer ribonucleic acid |
| Tc | T cytotoxic | TRP | transient receptor potential |
| Tc-99m | technetium | TRT | tinnitus retraining therapy |
| Tc-99m (v) DMSA | pentavalent dimercaptosuccinic acid | TSG | tumour suppressor gene |
| TC | thyroid cartilage | TSH | thyroid-stimulating hormone; or thyrotropin |
| TCF | tracheocutaneous fistula | TSHoma | TSH-secreting adenoma |
| TCI | target-controlled infusion | TSS | transitional space surgery |
| TCP | tricalcium phosphate | TT | thrombin time |
| TCR | T cell receptor | TTN | thalamic taste nucleus |
| TdT | terminal deoxynucleotidyl transferase | TTP | thrombotic thrombocytopenic purpura |
| TEC | Tissue Engineering and Regenerative Medicine Centre | TTR | transthyretin |
| TENS | transcutaneous electrical nerve stimulation | TTS | temporary threshold shift |
| TEOAE | transient evoked otoacoustic emission | TUNEL | TdT-mediated nick end labelling |
| TEP | tracheo-oesophageal puncture | TXA ₂ | thromboxane A ₂ |
| TFG | temporalis fascia graft | U | uracil |
| TFT | thyroid function test | UADT | upper aerodigestive tract |
| TG | thyroglobulin | UARS | upper airway resistance syndrome |
| TGF | transforming growth factor | UCL | uncomfortable loudness level |
| TGF- α | transforming growth factor alpha | UICC | International Union Against Cancer |
| TGF- β | transforming growth factor beta | UK-CCSG | United Kingdom Children with Cancer Study Group |
| TGF- β 1 | transforming growth factor beta 1 | UKCISG | UK Cochlear Implant Study Group |
| Th | T helper | UMN | upper motor neuron |
| TIA | transient ischaemic attack | UMP | uridine monophosphate |
| TIBC | total iron binding capacity | UNICEF | United Nations Children's Fund |
| TICA | totally implantable cochlear amplifier | UOS | upper oesophageal sphincter |
| TKI | tyrosine kinase inhibitor | UP | uncinate process |
| TM | tympic membrane | UPP | uvulopalatopharyngoplasty |
| TMC1 | transmembrane channel-like gene 1 | UPPP | uvulopalatopharyngoplasty |
| TMD | temporomandibular disorder | UPSIT | University of Pennsylvania Smell Identification Test |
| TMJ | temporomandibular joint | URT | upper respiratory tract |
| TMTF | temporal modulation transfer function | URTI | upper respiratory tract infection |
| TN | trigeminal neuralgia; or trigeminal nerve | US | ultrasound; or ultrasonography |
| TNF | tumour necrosis factor | USH | Usher syndrome |
| TNF- α | tumour necrosis factor alpha | USH1B | Usher syndrome type 1B |
| TNM | tumour, node, metastasis | USPIO | ultra-small super paramagnetic iron oxide |
| TOAE | transient evoked otoacoustic emission | UV | ultraviolet |
| TOE | transoesophageal echocardiography; or <i>Trichophyton</i> , <i>Oidiomycetes</i> and <i>Epidermophyton</i> | uVD | unilateral vestibular deafferentiation |
| TOF | tracheo-oesophageal fistula | UVPP | uvulopalatopharyngoplasty |
| TOF-o-gram | tracheo-oesophageal fistulogram | UWQOL | University of Washington Quality of Life Questionnaire |
| TORP | total ossicular replacement prosthesis | VA | Veterans' Affairs; or vestibular aqueduct |
| TPA | tissue polypeptide antigen | VAAP | voice activity and participation |
| TPF | docetaxel/cisplatinum/5-fluorouracil; or temporoparietal fascia | VAC | vacuum-assisted closure |
| TPHA | <i>T. pallidum</i> haemagglutination test; or treponemal haemagglutination | VAM | variation of amplitude |
| TPI | <i>T. pallidum</i> immobilization | VAS | visual analogue scale; or visual analogue score |
| TPN | total parenteral nutrition | VATER | vertebral, anal, tracheoesophageal and radial |
| TPO | thyroid peroxidase; or thyroperoxidase | VCA | viral capsid antigen |
| Tpot | potential doubling times | | |
| TOM | total quality management | | |

| | | | |
|--------|---|--------|--|
| VCAM-1 | vascular cell adhesion molecule-1 | VPQ | patient questionnaire of vocal performance |
| vCJD | variant Creutzfeldt-Jakob disease | VRA | visual reinforcement audiometry |
| VCR | vestibulocollic reflex | VRE | vancomycin-resistant enterococci |
| VDRL | Venereal Disease Research Laboratory | V-RQOL | voice-related quality of life |
| VEES | video endoscopic evaluation of swallowing | VS | vestibular schwannoma |
| VEGF | vascular endothelial growth factor | VSM | velocity storage mechanism |
| VEMP | vestibular-evoked myogenic potential | VSR | vestibulospinal reflex |
| VEP | vestibular evoked potential | VTE | venous thromboembolism |
| VFSS | videofluoroscopic swallowing study | VVI | vocal velocity index |
| VHI | Voice Handicap Index | vWD | von Willebrand disease |
| VHI-10 | Voice Handicap Index-10 | vWF | von Willebrand factor |
| VHL | Von Hippel-Lindau | VZV | varicella zoster virus |
| VHQ | Vertigo Handicap Questionnaire | | |
| VHT | vestibular habituation training | WAS | Wiskott Aldrich syndrome |
| VILI | ventilator induced lung injury | WBC | white blood cell |
| VIP | vasoactive intestinal polypeptide | WHO | World Health Organization |
| VLA | very late activation antigen | WMD | weighted mean difference |
| VLA4 | very late activation antigen 4 | WOB | work of breathing |
| LDL | very low-density lipoprotein | WP | Woodruff's plexus |
| VMA | vanillylmandelic acid | WPC | WARN, PAUSE, CHECK |
| VN | vestibular nucleii; or vagus nerve | | |
| VOC | volatile organic compound | XHIM | X-linked hyper immunoglobulin M |
| VOG | video-oculography | XLA | X-linked agammaglobulinaemia |
| VoiSS | voice symptom scale | XLP | X-linked lymphoproliferative syndrome |
| VOR | vestibulo-ocular reflex | | |
| VORP | vibrating ossicular prosthesis | YAC | yeast artificial chromosome |
| VORS | vestibulo-ocular reflex suppression | YAG | yttrium aluminium garnate |
| VPI | velopharyngeal insufficiency | | |

PART 1

CELL BIOLOGY

EDITED BY NICHOLAS S JONES

| | |
|---|-----------|
| 1 Molecular biology | 3 |
| <i>Michael Kuo and Richard Irving</i> | |
| 2 Genetics | 15 |
| <i>Karen P Steel</i> | |
| 3 Gene therapy | 23 |
| <i>Scott M Graham and John H Lee</i> | |
| 4 Mechanisms of anticancer drugs | 34 |
| <i>Sarah Payne and David Miles</i> | |
| 5 Radiotherapy and radiosensitizers | 47 |
| <i>Stewart G Martin and David AL Morgan</i> | |
| 6 Apoptosis and cell death | 56 |
| <i>Michael Saunders</i> | |
| 7 Stem cells | 66 |
| <i>A John Harris and Archana Vats</i> | |

Molecular biology

MICHAEL KUO AND RICHARD IRVING

| | | | |
|--|----|--|----|
| Introduction | 3 | Mapping and identification of genes associated with | |
| Molecular genetics: DNA structure and function | 3 | disease | 11 |
| Key points | 5 | Key point | 11 |
| Methods in molecular biology | 5 | Deficiencies in current knowledge and areas for future | |
| Key points | 8 | research | 11 |
| Molecular aberrations of cellular biology | 8 | References | 13 |
| Key points | 10 | Further reading | 14 |

SEARCH STRATEGY

The data in this chapter are supported by a Medline search using the key words molecular biology, genetics, and cell biology.

INTRODUCTION

Molecular biology describes the study of the biochemical processes that govern the behaviour of cells. These processes form the fundamental mechanisms by which cell function, cell–cell interactions and cell turnover are regulated. Disruption of this regulation may lead to disease, whilst an understanding of these mechanisms allows the physician to attempt to predict disease behaviour and to explore methods of restoring this regulation at a molecular level. This chapter reviews the principles of molecular genetics and outlines aspects of the molecular biology of the cell in the context of otolaryngological disease processes and describes some of the techniques that form the backbone of current molecular biology. It should give the reader sufficient background knowledge of molecular biology to understand subsequent chapters discussing the molecular biology of specific otolaryngological conditions.

MOLECULAR GENETICS: DNA STRUCTURE AND FUNCTION

Hereditary information in eukaryotes is stored in the form of double-stranded deoxyribonucleic acid (DNA)

and is referred to as the genome. DNA forms a double-helix structure as a result of hydrogen bonds between complementary pairs of nucleotides, adenine (A) with thymine (T) and cytosine (C) with guanine (G). The nucleotides on each strand are organized linearly in triplets, known as codons. Each specific sequence determines a single specific amino acid, for example ACU specifies threonine. However, as there are more triplet combinations (64) than commonly encountered amino acids (20), some proteins may be represented by different codons (e.g. lysine by AAA as well as AAG) and some codons (UAA, UGA and UAG) are ‘stop’ codons, constituting a signal for arrest of translation. The overwhelming majority of this DNA (99.9 percent) exists in the cell nucleus as the nuclear genome, which, in the human, is estimated to be 3000 megabase pairs in physical size and encodes 30,000–35,000 genes. The remaining DNA (16.6 kilobase pairs) forms the mitochondrial genome, encoding 37 genes. The mitochondrial genome and its potential role in cancer diagnostics will be discussed later.

Each DNA molecule is packaged into a chromosome by complex folding of the DNA around proteins. Diploid human cells contain 22 pairs of autosomes (1 to 22) and a

pair of sex chromosomes (XX or XY) which determines the sex of the organism. One of each pair of chromosomes is maternally inherited and the other is paternally inherited. Each chromosome has a distinctive shape, size and banding pattern, but have the common appearance of two arms apparently separated by a constriction. The centromere is microscopically recognizable as the central constriction separating the chromosome into a long arm (q for queue) and a short arm (p for petit), but its biological role lies in anchoring the chromosome to the mitotic spindle for segregation during cell division. The ends of the chromosomes are capped by telomeres, which are specialized structures containing unique simple repetitive sequences. They maintain the structural integrity of the chromosome and provide a solution for complete replication of the extreme ends of the chromosome. The conventional nomenclature for chromosomal locus assignment is given by the chromosome number, followed by the arm and finally the position on the arm, for example, 3p21 indicates position 21(two-one) on the short arm of chromosome three.

During normal cell division, DNA replication is achieved by the separation of the two strands by DNA helicase. Each separated single strand then acts as a template for polymerization, catalyzed by DNA polymerase, of nucleotides forming a new complementary strand and thus double-stranded DNA identical to the original dsDNA. As each daughter DNA consists of one original and one newly synthesized DNA strand, the process is known as semi-conservative replication. The specificity of the complementary relationship between the nucleotides on each strand forms the basis for many techniques of modern molecular biology and molecular cytogenetics.¹ The accuracy with which DNA replication takes place is remarkable with an estimated error rate of less than one in 10^9 nucleotide additions. Such accuracy is of vital importance to the individual as a permanent change in DNA, or mutation may cause inactivation of a gene essential to cell survival or cell cycle control. The high fidelity of DNA sequence replication is achieved by unidirectional 5'-to-3' direction of DNA replication, a rigorous DNA proofreading mechanism which detects mismatched DNA and efficient DNA repair pathways which excise and repair DNA damage. Failure of these mechanisms, such as is encountered in xeroderma pigmentosum, Fanconi's anaemia and ataxia telangiectasia, leads to accumulation of DNA replication errors and a high incidence of malignancies.

Although the human nuclear genome is 3×10^9 base pairs in size, about 90 percent of it is noncoding, with all the genes being coded by the remaining 10 percent of the DNA. Within the noncoding DNA are dispersed short arrays of repeat units of pairs or triplets of nucleotides (di-/trinucleotides). The exact function of these microsatellite repeats is not entirely clear, but their existence and frequency of dispersion throughout the genome have greatly facilitated study of the genetics of

tumours and many inherited disorders, which will be discussed later.

A gene is a region of the chromosomal DNA that produces a functional ribonucleic acid molecule (RNA). It comprises regulatory DNA sequences which determine when and in which cell types that gene is expressed, exons which are coding sequences and interspersed introns which are noncoding DNA sequences. These regulatory sequences often consist of CpG islands, short stretches of DNA rich in dinucleotides of cytosine and guanine. The methylation status of these CpG islands determines whether that gene is expressed in a particular cell or tissue, being unmethylated in tissues where the genes are expressed. As will be discussed later, aberration of this control is one of the mechanisms of tumour suppressor gene inactivation. Transcription is the intranuclear process driven by RNA polymerase whereby one of the two DNA strands acts as a template for the synthesis of a single RNA strand which is complementary to the DNA, except that uracil replaces thymine in RNA. This primary RNA transcript then undergoes post-transcriptional processing, or splicing.² Traditional dogma held that one gene produces one protein and therefore splicing was considered to occur simply in order to remove the noncoding intronic sequences, producing messenger RNA (mRNA). It is now known that by 'alternative splicing', one gene can result in the production of several different but often related proteins in different tissues.³

The mature mRNA then migrates into the cytoplasm where it acts as a template for the synthesis of a polypeptide during translation, a process regulated and catalyzed by cytoplasmic ribosomes. Successive amino acids are added to the polypeptide chain according to the triplet code on the mRNA, which is recognized by the transfer RNA (tRNA), to which each corresponding amino acid is covalently bound. Translation is commenced upon recognition of an initiation codon (usually but not exclusively AUG/methionine) and terminated upon recognition of a stop codon. The polypeptide subsequently undergoes a variable degree of post-translational modification and/or cleavage to produce the mature protein product, which may have an intracellular role or may be exported to the endoplasmic reticulum and hence to the extracellular space to execute its function.

The mitochondrial genome is considerably smaller than the nuclear genome, but it deserves mention here because of the increasing recognition of the role of mitochondrial DNA (mtDNA) mutations in human disease. The mitochondrial genome is only 16.6 kb in size, comprising 37 genes, which encode polypeptides which are principally involved in the respiratory chain. mtDNA is double-stranded but does not form a double-helix nor does it form chromosomes, but instead it takes the form of a circular double-stranded DNA structure with a heavy and a light strand. Unlike the nuclear

genome, which is inherited from mother and father, the mitochondrial genome of an individual is entirely maternally inherited.

KEY POINTS

- The double-stranded alpha helical structure of DNA, mainly located in the nucleus, consists of nucleotide triplets called codons which code for specific amino acids and stop signals, and forms the substrate for hereditary information in eukaryotes.
- The 22 pairs of autosomes and one pair of sex chromosomes, each with their distinctive shape, size and banding pattern, represent a complex folding of DNA around proteins to give the characteristic shape of a central constriction (centromere) separating the chromosome into a long arm (q) and a short arm (p) with a telomere cap at each end to maintain structural integrity.
- Chromosome locus nomenclature: chromosome number – 3p21 – position on chromosome arm.
- Semiconservative replication of DNA during normal cell division results in the separation of two strands of DNA by DNA helicase, each strand then acting as a template for polymerization by DNA polymerase. High fidelity is vital to prevent permanent change or mutations.
- A gene is a region of chromosomal DNA which produces functional RNA consisting of:
 - regulatory DNA sequences;
 - exons, which are coding sequences;
 - introns, which are noncoding sequences.
- Transcription is the intranuclear process driven by RNA polymerase whereby one of the two DNA strands acts as a template for single-stranded RNA synthesis complementary to the DNA, except that in RNA U is replaced by T. Splicing refers to post-transcriptional processing of RNA.
- Translation is the cytoplasmic process in which mRNA acts as a template for the synthesis of polypeptide by adding successive amino acids to the polypeptide chain, according to the triplet codon of the mRNA which is recognized by the tRNA to which the corresponding amino acid is covalently bonded. This process is regulated and catalyzed by cytoplasmic ribosomes. Post-translational modification produces mature proteins.

METHODS IN MOLECULAR BIOLOGY

Basic techniques of DNA fragmentation and identification

Unlike RNA, DNA is extremely stable, which is understandable from the function that each has in the cell. For purposes of studying the DNA and in order to clone specific DNA, the DNA molecule needs to be divided into manageable fragments. Although the ability to cut (and also to join up) DNA molecules now appears to be a very straightforward process, it was only 1970 when the first restriction endonuclease was identified in a strain of *Haemophilus influenzae*, hence its name *HindII* (pronounced Hin-dee-two). It is believed that this restriction endonuclease acts *in vivo* in bacteria as an immune or host-defence system, recognizing non-self DNA in bacteriophages and cleaving them. By surveying many different bacteria, a wide range of restriction endonucleases is now available, each of which recognize specific target sites based on sequences of four to eight nucleotides. As a specific, seven nucleotide sequence (heptanucleotide) will occur less frequently than a four nucleotide sequence (tetranucleotide), statistically, endonucleases recognizing heptanucleotide targets will cut less frequently thereby yielding larger fragments than those recognizing tetranucleotides. As the DNA is double-stranded, the resultant fragments may have blunt ends or cohesive ('sticky') ends (Figure 1.1). The nature of the ends of DNA fragments thus generated impact upon the way in which they can be ligated (joined) into recombinant molecules. Ligation of DNA fragments with cohesive ends is more efficient than joining of blunt-ended fragments.

ELECTROPHORESIS

Negatively charged phosphate groups on the DNA backbone confer a net negative charge on linear DNA. This allows fragments of different sizes to be resolved within a suitable gel matrix by the application of an electric current across the matrix. The DNA will migrate toward the positive electrode with the smaller fragments travelling faster than the larger fragments.⁴ The size of the fragment can be estimated by the use of a graduated DNA

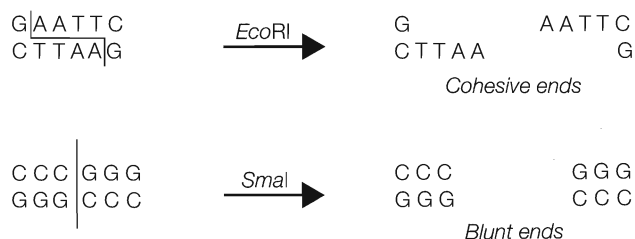


Figure 1.1 DNA cleavage by restriction endonucleases. Derived from Ref. 11, with permission.

ladder containing fragments of known molecular weight. The choice of the particular matrix depends on the fragment sizes that one is trying to resolve. Polyacrylamide gels can resolve differences of just one base pair between fragments of several hundred base pairs in size by virtue of a small pore size in the gel matrix. These gels can be used for DNA sequencing and resolution of alleles varying in only one dinucleotide repeat. Agarose gels can resolve fragment sizes from around 100 bp to 20 kb. Beyond that size, electrophoretic mobility is no longer proportional to fragment size. Resolution of fragments sizes in excess of 50 kb, such as larger bacterial artificial chromosomes (BAC) or yeast artificial chromosomes (YAC) require the use of pulsed field electrophoresis.

HYBRIDIZATION

Hybridization is the specific annealing of single DNA (or RNA) strands, the probe, to a DNA sample, the target. It serves to detect the presence of a specific sequence of DNA either in the cell or on a hybridization membrane and recognition that hybridization has occurred is achieved either by radioactively labelling the probe and localizing the radioactivity by autoradiography or by labelling the probe with fluorochromes which fluoresce when excited by light of specific wavelengths (**Figure 1.2**). Hybridization on a membrane requires the initial transfer of DNA on to a nitrocellulose membrane from an agarose gel. This elegantly simple process is eponymously known as Southern blotting after the scientist who described the process in 1975. Two other commonly used transfer techniques have their names derived from Southern blotting as jargon terms. Northern blotting is essentially the same process used for transfer of RNA to a membrane. Western blotting is one of the mainstays of protein analysis and involves the transfer of electrophoresed protein bands from a polyacrylamide gel on to a nitrocellulose or nylon membrane to which they bind strongly. Detection of the protein is usually achieved by the use of antibodies to specific antigens presented by the protein with the antibody being labelled radioactively, enzymatically or fluorescently.

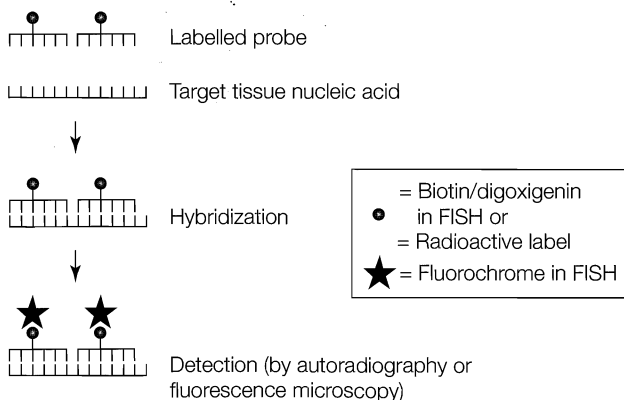


Figure 1.2 *In situ* hybridization.

CYTOGENETICS AND MOLECULAR CYTOGENETICS

Although microscopy had already reached high levels of resolution in the early 1930s, the correct number of human chromosomes was not determined until 1958. The era of classical cytogenetics was thus begun. Cytogenetics is the study of chromosomal abnormalities and rearrangements. It currently has a major role to play in prenatal diagnosis of Down's syndrome and other congenital syndromes characterized by numerical chromosomal abnormalities. In the early part of this century, Theodore Boveri proposed that cancer arose from chromosomal alterations. This hypothesis was not proven until the consistent chromosomal translocation, t(9;22), was demonstrated in chronic myeloid leukaemia. Since that time, cytogenetic analysis has been the mainstay of genetic analysis in reticuloendothelial malignancies, being responsible for the identification of consistent translocations in different leukaemias. Its use in solid tumours has been hampered by the difficulties of establishing short-term primary cultures from head and neck cancers for chromosomal analysis and the erratically acquired chromosomal changes in long-term cell lines, which may have occurred *in vitro*, influenced by culture conditions. Nevertheless, some studies have identified chromosomal areas consistently showing frequent breakpoints suggesting the location of putative tumour suppressor genes (including 3p21, 5p14, 8p11, 17p21, 18q21) and gain or amplification implying the presence of putative proto-oncogenes at other sites (including 3q, 5p, 8q, 11q13). Although the refinement of karyotyping has been radically enhanced by the introduction of 24-colour combinatorial multicolor FISH (M-FISH), the resolution and therefore utility of solid tumour karyotyping remains limited.⁵

Hybridization to target DNA in cells, using fluorescence detection, is known as fluorescence *in situ* hybridization (FISH). Fluorescence *in situ* hybridization allows the analysis of copy number of a known specific DNA sequence within intact nuclei. In reticuloendothelial malignancies and solid tumour-derived cell lines, the use of both single-copy probes and centromere alpha-satellite repeat probes on metaphase preparations has enhanced and refined classical karyotyping. Interphase FISH has been applied to solid tumour sections to assess the copy number of a known sequence in breast, prostate, bladder, brain, lung and head and neck tumours.

Fluorescence-labelled hybridization has also been combined with cytogenetics to produce the powerful technique of comparative genomic hybridization (CGH).⁶ Comparative genomic hybridization permits the rapid medium resolution screening of the entire genome by comparatively hybridizing matched tumour and normal DNA from a patient, which are labelled with different fluorochromes, on to normal metaphase chromosome preparations. Under red-green dual filter fluorescence microscopy and computer-aided image analysis, areas of

genetic 'neutrality' appear yellow, under-representation appears green, and over-representation appears red. Areas of genetic under-representation suggest the possibility of a tumour suppressor gene lying within that region while areas of over-representation may indicate the location of a putative oncogene. This technique has been applied to the rapid genetic analysis of many tumour types including squamous cell carcinomas of the head and neck. The advent of molecular cytogenetics has obviated the need for primary short-term cultures and refined the location of chromosomal aberrations in solid tumours.

POLYMERASE CHAIN REACTION

Perhaps the single molecular technique which has had the most dramatic impact on molecular biology has been the polymerase chain reaction (PCR). The original problem lay in obtaining sufficient quantities of a particular DNA sequence such that DNA profiling (e.g. sequencing) and DNA manipulation (e.g. cloning) could be achieved. The only 'requirement' is that the sequences flanking the stretch of DNA of interest is known. With that proviso, PCR achieves faithful and exponential amplification of a specific sequence of DNA by repeated cycles each consisting of dsDNA denaturation, hybridization of specific oligonucleotides (primers) and extension of the polynucleotide by rapidly altering the reaction temperature between segments of each cycle. dsDNA denaturation is achieved by raising the temperature of the reaction to 94°C for 30 seconds, thus disrupting the hydrogen bonds between the strands and exposing the hydrogen bond donor and acceptor groups to allow base pairing. The oligonucleotide primers are then allowed to hybridize to the denatured DNA (annealing) at around 55–65°C for 90 seconds before the reaction temperature is raised to 72°C to permit extension of the DNA strand by DNA polymerase in the presence of deoxynucleoside triphosphates (dNTPs). With each cycle resulting in the

doubling of the copies of the DNA sequence, a 30-cycle PCR taking approximately two hours would amplify a single copy of a DNA sequence 268 million-fold (Figure 1.3). Although the PCR was originally described by Mullis and Faloona in 1987, one practical problem prevented its instant exploitation.⁷ The DNA polymerase used in the original reaction was denatured during the DNA denaturation segment and therefore had to be added after each and every cycle. The solution came in 1989 when Lawyer isolated and characterized the DNA polymerase, Taq polymerase, from the thermophilic bacterium *Thermus aquaticus* which normally resided in temperatures above 95°C.⁸ This polymerase was therefore 'heat resistant' and did not need to be replenished between cycles.

The PCR holds a central position in many molecular biological techniques as well as clinical diagnostic methods. The fundamental principle of DNA amplification has been adapted to amplify messenger RNA and to amplify areas where the initial flanking oligonucleotide sequences are not known. It is often described as a sensitive and powerful technique, but with great power comes the potential for corruption! In theory, a single copy of DNA can be amplified. Therefore, careless experimental technique may lead to contamination of the DNA sample with other DNA (e.g. from the skin of the investigator) and consequently to an artefactual result. The Taq polymerase originally described in the technique does not have proofreading properties, but newer cloned enzymes such as Pfu polymerase incorporates a proof-reading function to increase amplification fidelity for sequencing reactions.

The sensitivity of PCR also presented a problem for the analysis of genetic alterations in certain solid tumours. Squamous cell carcinomas of the head and neck are histologically often characterized by a large stromal element within the tumour. The genetic alterations in the tumour may not be present in the stromal

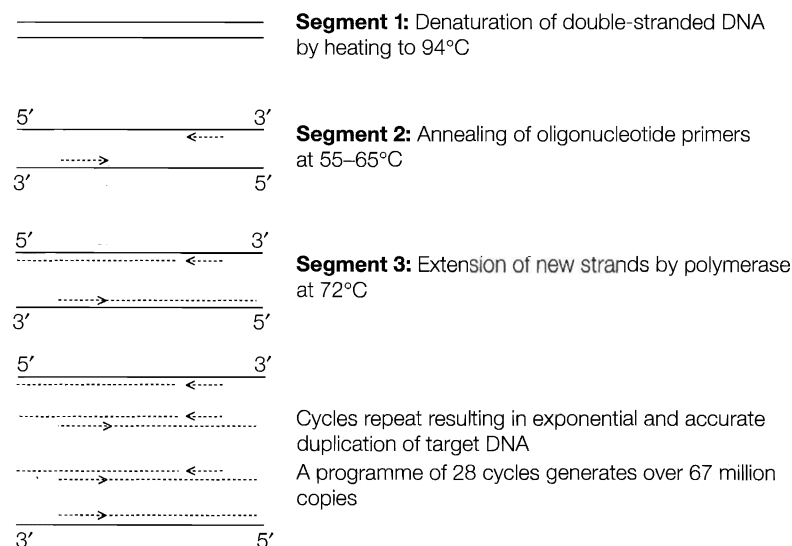


Figure 1.3 The polymerase chain reaction.

tissue and thus total DNA extracted from the tumour will contain DNA from both benign and malignant tissue. This *in situ* contamination can now be eliminated by the use of laser capture microdissection (LCM) of tumours. LCM involves the placement of a laser-activated film over a tissue specimen. When areas of 'pure' tumour cells are identified, a focal laser pulse lifts the tissue on to the film in specimens down to 30 µm in diameter.⁹

KEY POINTS

- Restriction endonucleases are enzymes that were initially identified in bacteria that can cut and join up DNA. They recognize specific target sites based on sequences of four and eight nucleotides.
- Electrophoresis is a technique for resolving the size of DNA fragments, which carry a negative charge from the phosphate groups on their backbone. Using a gel matrix with an electric current applied across it, the DNA will migrate to the positive electrode at a rate inversely proportional to its size.
- Hybridization is the specific annealing of single DNA or RNA strands (probe) to a DNA sample (target) to detect the presence of a specific sequence of DNA in the cell or hybridization membrane. Variants include the eponomously named Southern, Northern and Western blotting techniques.
- Cytogenetics is the study of chromosomal abnormalities and rearrangements important in the diagnosis of congenital syndromes characterized by numerical chromosomal abnormalities, e.g. Downs syndrome and leukaemia types.
- FISH refers to fluorescence *in situ* hybridization which involves hybridization to target DNA cells using fluorescence detection and allows the analysis of copy number of a known specific DNA sequence within intact nuclei.
- PCR achieves faithful and exponential amplification of a specific sequence of DNA by repeated cycles each consisting of:
 - DNA denaturation by heating to 94°C to denature hydrogen bonds between strands;
 - annealing (hybridization) of oligonucleotide primers to denatured DNA at 55–65°C;
 - extension of DNA strand by DNA polymerase.

MOLECULAR ABERRATIONS OF CELLULAR BIOLOGY

Loss of heterozygosity and the expression of recessive mutant alleles

Retinoblastoma is a childhood cancer, which exhibits both hereditary and sporadic occurrence, with the inherited form transmitted as a highly penetrant autosomal dominant trait. The proposition by Alfred Knudson in 1971, based upon a statistical analysis of the occurrence of retinoblastoma in children, that two genetic events were required to inactivate the gene mitigating against development of the cancer, was a major landmark in the understanding of tumour suppressor genetics.¹⁰ In hereditary retinoblastomas, a single additional somatic event in a cell that carried the inherited mutation was sufficient to give rise to the disease while two somatic events were required to produce a sporadic retinoblastoma. This became known as Knudson's 'two-hit' hypothesis. The subsequent study on matched tumour and blood DNA from patients with sporadic retinoblastoma by Webster Cavenee not only proved Knudson's hypothesis but also established the paradigm for all subsequent investigations of tumour suppressor genes.¹¹ For the first time, the now widely accepted mechanisms of tumourigenesis were reconciled, viz. that neoplasms can arise in a multistep manner, that chromosomal events can lead to tumour formation and that chromosome loss with or without reduplication can lead to expression of recessive mutations. Perhaps even more strikingly, the authors presciently suggested that development of homozygosity for recessive mutant alleles at the *Rb-1* locus may give rise to the development of other tumours and that other additional dominant mutations may be involved in the development of retinoblastoma. Cavenee proposed the various chromosomal mechanisms that could reveal recessive mutations and these are summarized for a putative tumour suppressor gene in **Figure 1.4**, adapted from the figure in his original paper. To these can now be added hypermethylation of the 5' CpG island resulting in transcriptional inactivation of the gene, discussed below.¹² The simplest way of revealing a recessive mutant allele is by deletion of the wild-type allele, resulting in hemizyosity at the particular locus on the remaining chromosome. It is inferred from this that areas of frequent allelic loss in tumours may represent the location of putative tumour suppressor genes and this hypothesis underpins the commonly employed method of molecular detection of allelic losses, loss of heterozygosity (LOH).

The practical exploitation of the concepts outlined above hinges on the presence of the previously described microsatellites, highly polymorphic noncoding DNA sequences, also referred to as simple sequence length polymorphisms (SSLP) or short tandem repeat

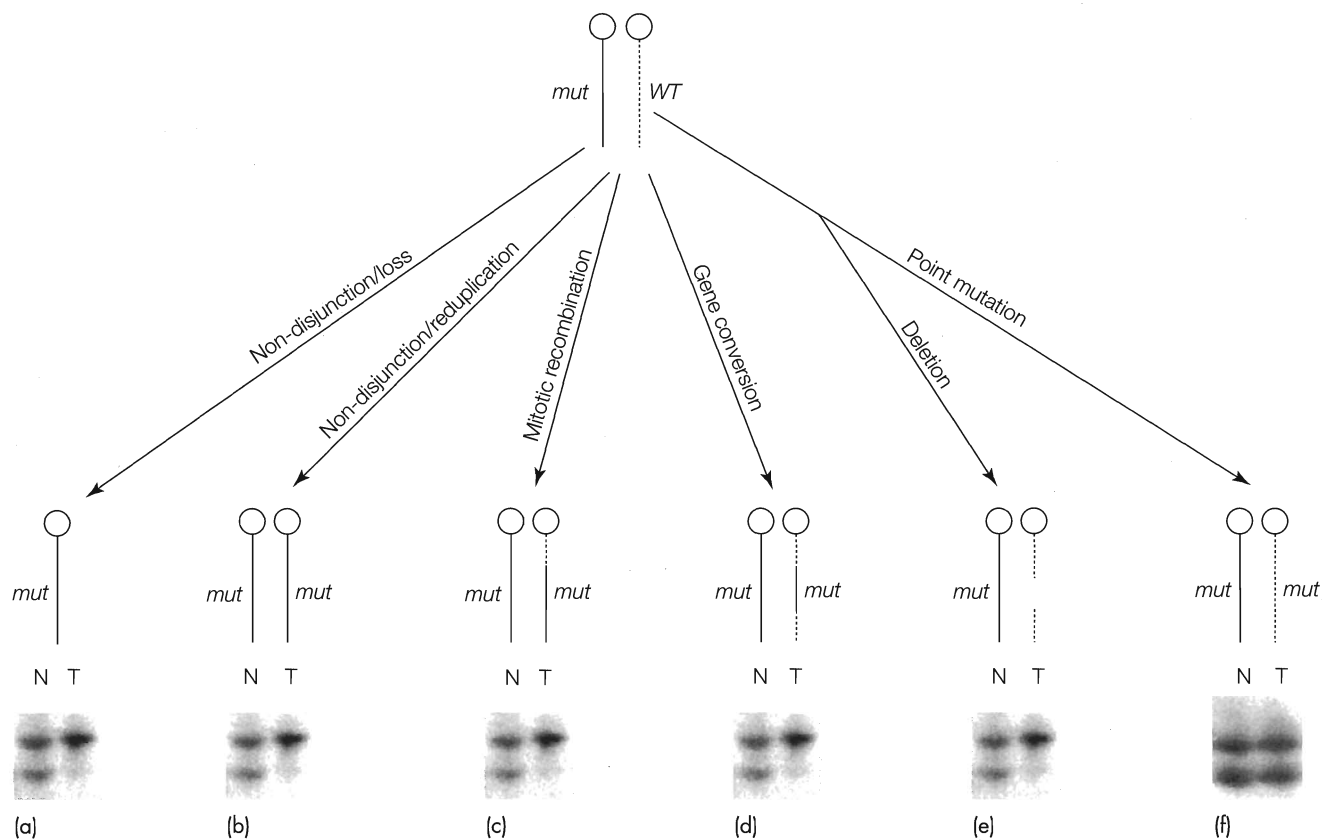


Figure 1.4 Chromosomal mechanisms that could reveal recessive mutations. In this example, before cell division, the tissue concerned carries a mutation in one copy of the hypothetical tumour suppressor gene. In each of the scenarios (a–f), the recessive mutation is revealed. If the individual is heterozygous for a microsatellite marker within or very close to the mutated gene, the hypothetical PCR results are given below each ideotype. The only mechanism which escapes observed loss of heterozygosity is F. *mut*, mutated; N, normal; T, tumour; *WT*, wildtype. After Ref. 11, with permission.

polymorphisms (STRP), which are distributed approximately every 100,000 bp throughout the human genome. These microsatellites contain small dinucleotide or trinucleotide repeat units, the number of which may differ between the two alleles in a particular person. Microsatellite markers are now available which map thousands of these sequences to chromosomal loci. When DNA sequences containing these microsatellite markers are amplified by PCR in a person heterozygous for that particular microsatellite, the PCR will yield two products of different lengths, which can be resolved on an electrophoretic gel. Where amplification of tumour DNA from such a subject yields only one product, the tumour is said to show LOH, implying allelic loss. Persons who are homozygous for a particular marker are said to be noninformative for that marker. The concept of examining the variation and extent of allelic deletion in tumours was introduced by Vogelstein in an analysis of colorectal carcinomas and termed allelotyping.¹³ Allelotypes generated in this fashion have identified several areas of frequent allelic deletion from which some of the responsible tumour suppressor genes have been cloned or identified. The most common areas of loss in HNSCC are

at chromosome 9p21, 17p21, 13q14, 4p, 5q21 and several discrete regions on 3p and 8p.^{14,15}

Inactivation of genes and oncogenic transformation

Allelic deletion is only one mechanism by which a copy of a gene can be inactivated. As there are two copies of each gene, inactivation of the gene requires inactivation of both copies of the gene, ‘the second hit’. This may occur as a result of a genetic mutation or transcriptional silencing. Conversely, a protooncogene may be converted into an oncogene by a simple increase in the copy number of the gene (gene amplification) resulting in an overproduction of protein or by point mutations that affect the control of protein activity.

Not all mutations result in alteration in function of a gene. DNA mutation may occur as a result of base substitutions, as well as nucleotide insertions and deletions. Insertions and deletions of nucleotides are very rare in coding DNA, but if they occur they may produce a shift in the ‘reading frame’ which dramatically alters the