

Mario Mandalà  
Emanuela Romano *Editors*

# Mechanisms of Drug Resistance in Cancer Therapy

---

# Handbook of Experimental Pharmacology

Volume 249

**Editor-in-Chief**

J.E. Barrett, Philadelphia

**Editorial Board**

V. Flockerzi, Homburg

M.A. Frohman, Stony Brook, NY

P. Geppetti, Florence

F.B. Hofmann, München

M.C. Michel, Mainz

C.P. Page, London

W. Rosenthal, Jena

K. Wang, Qingdao

More information about this series at <http://www.springer.com/series/164>

---

Mario Mandalà • Emanuela Romano  
Editors

# Mechanisms of Drug Resistance in Cancer Therapy

 Springer

*Editors*

Mario Mandalà  
Department of Oncology and Hematology  
Papa Giovanni XXIII Hospital  
Bergamo, Italy

Emanuela Romano  
Department of Oncology, INSERM Res  
Unit 932  
Institut Curie  
Paris Cedex 05, France

ISSN 0171-2004

ISSN 1865-0325 (electronic)

Handbook of Experimental Pharmacology

ISBN 978-3-030-10506-8

ISBN 978-3-030-10507-5 (eBook)

<https://doi.org/10.1007/978-3-030-10507-5>

Library of Congress Control Number: 2018966536

© Springer Nature Switzerland AG 2018

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, express or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Switzerland AG  
The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland

---

## Preface

The remarkable advances in precision medicine and the development of targeted cancer treatments have generated significant optimism based on the assumption that a better knowledge of genetic or molecular features sustaining cancer growth could determine long-term control of many types of cancer and potentially cure the majority of patients. After several years of basic and translational research, the oncology community has tempered this optimism by the recognition that in only a minority of patients cure can be achieved by innovative strategies such as targeted therapies, immunotherapy and cytotoxic agents. In the vast majority of cancer patients, cancer wins the battle, and the reason for this failure is drug resistance. Ultimately, advanced cancer patients die because some or all of their tumour cells exhibit or develop resistance to available therapeutic strategies. The challenge of drug resistance therefore represents an important barrier that hinders the ultimate goal of cure or at least long-term control of cancer.

As a corollary of this, response rates in the setting of tumour progression/relapse are dismal. Despite the considerable importance of drug resistance to cancer therapies, our understanding of their biological mechanisms – and plausible therapeutic avenues to intercept them – remains highly incomplete.

Knowledge of specific resistance mechanisms can inform novel therapeutic approaches to counter this phenomenon, and improvement of our understanding of key driver genes could guide new therapeutic approaches capable of eliciting meaningful tumour responses in patients with advanced malignancies.

As previously reported with chemotherapy, it is reasonable to suppose that a combination of multiple targeted therapies and immunotherapy strategies as well as planned sequence of treatments will be necessary to effectively prevent and/or treat drug-resistant cancers. The potential number of therapeutic combinations is immense; thus, new preclinical paradigms are needed to prioritize high-yield combinations and define the genetic or molecular contexts in which they would most likely be efficacious. To improve outcome of patients a strict collaboration between academia and industry will be needed so that the appropriate resources and innovation may be brought to bear on this challenge. The goal to achieve durable control of many cancer subtypes will likely require dedicated, multidisciplinary teams of preclinical and clinical experts that work together guided by rigorous translational and analytical science.

With the increasing armamentarium of anticancer agents and the advent of powerful high-throughput screening techniques, there are now unprecedented opportunities to understand and overcome drug resistance through the clinical assessment of rational therapeutic drug combinations and the use of predictive and prognostic biomarkers to enable patient stratification and tailor treatments.

The main aim of this book is to offer to the readers an updated overview on the possible reasons of failure of new and promising therapeutic opportunities.

The first part of the book covers the basic mechanisms of such hot topics. The other chapters cover specific pathways of resistance that from the knowledge of basic mechanisms translate this information in clinical routine or in translational clinical research.

Readers will discover diverse perspectives of the contributing authors (such as basic scientists, clinical pharmacologists and clinicians) and extensive discussions of issues including pharmacodynamic and pharmacokinetic mechanisms of resistance, alterations in the drug target, activation of prosurvival pathways, ineffective induction of cell death and plasticity of microenvironment.

This book will be of interest to scholars and researchers, biologists, pharmacologists, medical oncologists, haematologists and immunologists.

Bergamo, Italy  
Paris Cedex 05, France

Mario Mandalà  
Emanuela Romano

---

# Contents

<b>Molecular and Pharmacological Mechanisms of Drug Resistance: An Evolving Paradigm</b> . . . . .	1
Benedetta Colmegna, Lavinia Morosi, and Maurizio D’Incalci	
<b>Major Physiological Signaling Pathways in the Regulation of Cell Proliferation and Survival</b> . . . . .	13
Huifang Tang and Gongda Xue	
<b>Immune-Mediated and Hypoxia-Regulated Programs: Accomplices in Resistance to Anti-angiogenic Therapies</b> . . . . .	31
Diego O. Croci, Santiago P. Mendez-Huergo, Juan P. Cerliani, and Gabriel A. Rabinovich	
<b>Mechanisms of Resistance to Target Therapies in Non-small Cell Lung Cancer</b> . . . . .	63
Francesco Facchinetti, Claudia Proto, Roberta Minari, Marina Garassino, and Marcello Tiseo	
<b>Mechanisms of Drug Resistance in Melanoma</b> . . . . .	91
Matthew Winder and Amaya Virós	
<b>Mechanisms of Resistance to Immune Checkpoint Antibodies</b> . . . . .	109
Rodrigo N. Ramos, Eliane Piaggio, and Emanuela Romano	
<b>Immunotolerance as a Mechanism of Resistance to Targeted Therapies in Melanoma</b> . . . . .	129
Mario Mandalà and Daniela Massi	
<b>Plasticity of Resistance and Sensitivity to Anti-Epidermal Growth Factor Receptor Inhibitors in Metastatic Colorectal Cancer</b> . . . . .	145
Andrea Sartore-Bianchi and Salvatore Siena	
<b>Targeting DNA Repair</b> . . . . .	161
Giuseppe Curigliano	
<b>Resistance to Hormonal Therapy in Prostate Cancer</b> . . . . .	181
Alfredo Berruti and Alberto Dalla Volta	



---

<b>Mechanism of Resistance in Gastrointestinal Stromal Tumors . . . . .</b>	<b>195</b>
E. Tamborini	
<b>Mechanisms of Resistance to Targeted Therapies in Chronic Lymphocytic Leukemia . . . . .</b>	<b>203</b>
Francesca Arruga and Silvia Deaglio	
<b>Mechanisms of Resistance to Targeted Therapies in Chronic Myeloid Leukemia . . . . .</b>	<b>231</b>
Federico Lussana, Tamara Intermesoli, Paola Stefanoni, and Alessandro Rambaldi	
<b>Mechanisms of Resistance in Multiple Myeloma . . . . .</b>	<b>251</b>
Athanasios Papadas and Fotis Asimakopoulos	
<b>Cancer Evolution as the New Frontier of Precision Medicine . . . . .</b>	<b>289</b>
Giuseppe Curigliano	



# Molecular and Pharmacological Mechanisms of Drug Resistance: An Evolving Paradigm

Benedetta Colmegna, Lavinia Morosi, and Maurizio D’Incalci

## Contents

1	Introduction .....	2
2	Cellular Mechanism of Multidrug Resistance .....	4
2.1	Drug Activation and Inactivation .....	4
2.2	Expression of Drug Efflux Pumps .....	5
2.3	DNA Damage Repair .....	7
2.4	Deregulation of Apoptosis .....	8
2.5	Alteration of Drug Target .....	9
3	Conclusion .....	10
	References .....	11

## Abstract

The high heterogeneity and genomic instability of malignant tumors explains why even responsive tumors contain cell clones that are resistant for many possible mechanisms involving intracellular drug inactivation, low uptake or high efflux of anticancer drugs from cancer cells, qualitative or quantitative changes in the drug target. Many tumors, however, are resistant because of insufficient exposure to anticancer drugs, due to pharmacokinetic reasons and inefficient and heterogeneous tumor drug distribution, related to a deficient vascularization and high interstitial pressure. Finally, resistance can be related to the activation of anti-apoptotic and cell survival pathways by cancer cells and often enhanced by tumor microenvironment.

B. Colmegna • L. Morosi • M. D’Incalci (✉)

Department of Oncology, IRCCS ‘Mario Negri’, Institute for Pharmacological Research, 20145 Milan, Italy

e-mail: [maurizio.dincalci@marionegri.it](mailto:maurizio.dincalci@marionegri.it)

© Springer International Publishing AG 2017

M. Mandalà, E. Romano (eds.), *Mechanisms of Drug Resistance in Cancer Therapy*, Handbook of Experimental Pharmacology 249, DOI 10.1007/164\_2017\_20

---

**Keywords**

Apoptosis • Drug distribution • Mass spectrometry imaging • Multidrug resistance • Pharmacokinetics

---

## 1 Introduction

Although significant improvement of the therapy of several human malignancies have been achieved in the last two decades with the use of small molecules or antibodies designed to hit cancer-specific molecular targets, for the majority of solid human tumors at advanced stage pharmacological treatments have only a transient antitumor effect with small or no impact on patients' survival. The major reason for this disappointing result is the phenomenon of drug resistance.

Although most recent research has been focused on molecular and cellular mechanism of resistance, it seems likely that in many cases the lack of antitumor activity is related to pharmacokinetic reasons, essentially associated with an insufficient drug concentration in the cancer cells.

The pharmacokinetic factors that can be involved in the failure of treatment can be dependent on the route of administration. For example, in the case of orally administered drugs the variable absorption as well as the first pass effect can be responsible for an insufficient bioavailability. For many anticancer drugs the variable metabolism can be a clinically relevant problem, also in consideration of potential interactions with other drugs interfering with the activity of cytochromes involved in drug biotransformation. These pharmacokinetic factors can be investigated by monitoring plasma drug levels, thus allowing, at least in theory, a dose adjustment or a change in the route of administration.

However, the drug plasma levels that for other classes of drugs predict the drug concentration at the pharmacological target, for anticancer drug are often meaningless. The circulating drug concentrations in fact do not necessarily mirror those present in the neoplastic tissue and its metastases because of tumor architecture and microenvironment (Fuso Nerini et al. 2014).

It is known that the vascularization of solid tumors is often inefficient, as the angiogenic process occurring in growing tumors leads to unorganized capillary network with fenestrated vessels and significant leakage of proteins responsible for an increase in the interstitial pressure. Probably further factors contributing to the high interstitial pressure are the inefficient lymphatic drainage as well as a significant infiltration of inflammatory and mesenchymal cells that represent common features of many solid tumors. This peculiar abnormal vascularization and tissue architecture is responsible for a low and heterogeneous tumor drug distribution (Minchinton and Tannock 2006). The evidence of this heterogeneous distribution of anticancer drugs was already available several years ago by examining different parts of tumors. It was demonstrated that in necrotic or hypovascularized and hypoxic parts of the tumor the drug concentrations were lower than those that were normally perfused by blood.

Recently new powerful technologies have been developed to investigate drug distribution in different parts of the tumor by simultaneous visualization and quantification of the drug levels. Among these new approaches, mass spectrometry imaging (MSI) is an interesting technique to obtain specific information on the spatial distribution of a drug linking qualitative molecular information of compounds to their spatial coordinates and distribution within the investigated tissue (Morosi et al. 2013).

MSI can be applied to visualize potentially every tissue component localizing the molecule of interest on the base of its molecular mass and fragmentation pattern, without the need to label the analyte, a great advantage over other imaging techniques such as fluorescence microscopy, positron emission tomography, magnetic resonance spectroscopy, and autoradiography (Sugiura and Setou 2010).

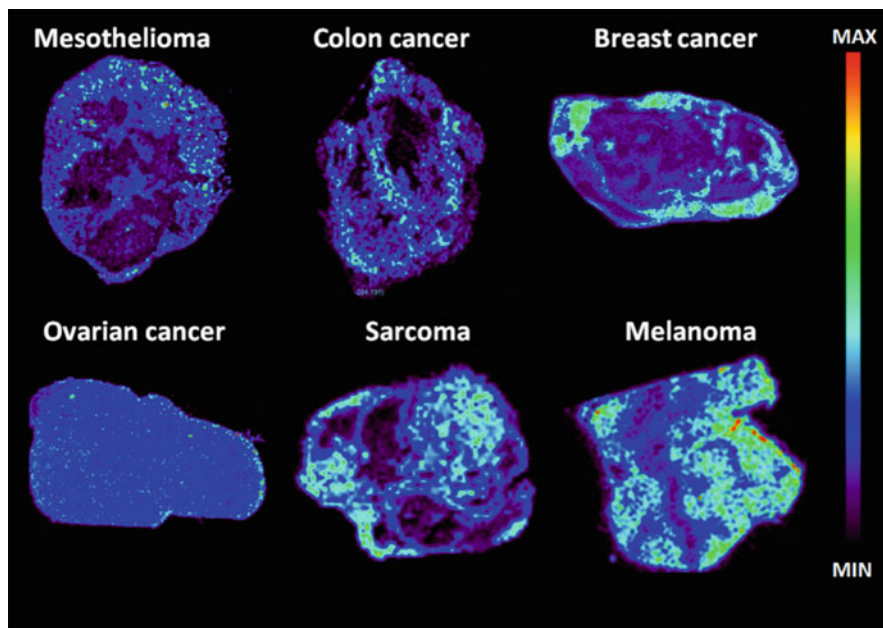
MSI was applied to localize drug distribution and penetration inside solid tumors, thanks to its superior spatial resolution (20–100  $\mu\text{m}$ ) and specificity allowing the detection of parent compound and metabolites simultaneously (Buck et al. 2015; Connell et al. 2015).

MSI can also provide the quantitative amounts of target compounds in well-defined region of interest of the examined tissue, ideally in a single pixel. This goal is extremely challenging depending on the technical possibilities and limitations of the MSI instrument hardware, but equally on the chosen calibration/standardization strategy (Giordano et al. 2016a; Rzagalinski and Volmer 2016).

MSI has also some limitation: it is only applicable to molecules that are ionizable, the selection of optimal matrix is arduous and driven by empirical results, moreover the interpretation of quantitative data is often complex especially because of ion suppression effect and the chemical noise from the matrix ions covering the drug ion signal and finally, the sensitivity is quite limited (Prideaux and Stoeckli 2012).

By applying this methodology it has become clear that drug distribution in tumors is very heterogeneous comparing different tumor models having different histopathological characteristics (Giordano et al. 2016b). Figure 1 shows the intratumor distribution of the anticancer drug paclitaxel by MSI. The drug distribution is homogeneous only in the ovarian cancer model while it is highly irregular in the other examples of tumor types. As you can see in other models such as sarcoma or breast cancer, areas of the tumor where the drug is highly concentrated and areas where it is almost absent can be observed at the same time. Broad parts of the tumor where the drug is not present at all are present especially in mesothelioma xenograft. This aspect surely contributes to explain tumor resistance. Torok et al. recently correlate the antitumor activity of several antiangiogenic drugs with their intratumoral distribution data obtained by MSI (Torok et al. 2017). Moreover, Cesca et al. observed an enhancement in paclitaxel activity combined with bevacizumab associated with the improvement of its intratumor distribution (Cesca et al. 2016).

The possibility to superimpose MSI molecular images with histological and immune-histological evaluation could help understanding the factors that impair drug penetration. Finally, this technology could be used in future to evaluate the



**Fig. 1** Paclitaxel distribution in different cancer models analyzed by MALDI mass spectrometry imaging. *Light blue* and *green* mark the drug presence on the basis of the color scale on the right

effect of therapeutic strategies aimed at modifying tumor microenvironment or altering drug properties to facilitate drug uptake and intratumor distribution.

## 2 Cellular Mechanism of Multidrug Resistance

The hallmarks of cellular anticancer drug resistance fall into distinct categories:

### 2.1 Drug Activation and Inactivation

Altered local drug metabolism and detoxification are key resistance mechanisms highly specific for each class of drugs. Lack of prodrug activation or epigenetic silencing – by promoter methylation – of genes involved in drug processing are examples of potential cause of resistance and are frequently tumor-specific. Classical examples could be the inactivation of platinum drugs by the thiol glutathione (Meijer et al. 1992) or metallothioneins (Amable 2016), or by epigenetic mechanisms, e.g., silencing – by methylation – of gene encoding for thymidine phosphorylase, an enzyme involved in the conversion of capecitabine (prodrug) to 5-fluorouracil (active drug) (Longley et al. 2003). For many DNA interacting agents, e.g., alkylating agents, anthracyclines, and platinum drugs, an important

mechanism of inactivation is related to the overexpression of Glutathione-S-Transferases (GSTs) that metabolizes the drugs into inactive molecules (Kauvar et al. 1998).

## 2.2 Expression of Drug Efflux Pumps

Plasma membrane transport proteins are key players in the uptake of nutrients, such as sugars, amino acids, nucleosides, and inorganic ions and the efflux of xenobiotic toxins, including many anticancer drugs. Transport proteins can be classified into two major families, the solute carrier (SLC) and the ATP-binding cassette (ABC) transporters and can affect drug absorption, distribution, and excretion conferring sensitivity or resistance to anticancer drugs.

The importance of ABC transporters in cancer therapy has been well documented while the impact of the solute carriers (SLC) on cancer therapy has not been extensively characterized (Table 1).

So far, the most characterized transporters belong to ABC family and include ABCB1 (also known as MDR1 or P-glycoprotein), ABCC1 (MRP1), and ABCG2 (BCRP or MXR).

**Table 1** Reports the transporters whose activity has been associated with drug resistance

Transporters	Chemotherapeutic drugs effluxed
P-gp/MDR1	Anthracyclines, taxanes, vinca alkaloids, imatinib, etoposide
MRP1	Anthracyclines, methotrexate, camptothecins, etoposide, glutathione conjugates
MRP2	Doxorubicin, methotrexate, cisplatin, vincristine
MRP3	Methotrexate, etoposide
MRP4	Methotrexate, camptothecins, thiopurines
MRP5	Methotrexate, thiopurines
MRP6	Etoposide
MRP8	5-Fluorouracil
BCRP/MXR	Doxorubicin, mitoxantrone, topotecan, flavopiridol, SN-38
LRP	Doxorubicin, cisplatin, vincristine, etoposide, paclitaxel, melphalan
Human nucleoside transporters (hENT, hCNT)	Cytarabine, gemcitabine, fludarabine, clofarabine
Copper transporters (Ctr1, Ctr2, ATP7A, ATP7B)	Cisplatin
Multidrug extrusion transporter1 (MATE1)	Cisplatin
Organic cation transporter (OCT1, -2, -3)	Platinum drugs, SN-38, mitoxantrone, melphalan
Organic anion transporter (OAT1, -2, -3)	Methotrexate, 5-fluorouracil, paclitaxel

The ATP-binding cassette (ABC) transporters superfamily is a large group of membrane proteins that are responsible for the translocation of various cytotoxic molecules out of the cell reducing their intracellular concentration and thus their cytotoxicity. ABC transporters are codified by 48 genes (divided into 7 subfamilies) and characterized by a conserved quaternary structure containing 2 transmembrane domains (TMDs) and 2 nucleotide (ATP)-binding domains (NBDs). Their ATP-dependent activity is involved in the movement of a wide variety of xenobiotics – including drugs – lipids, peptides, and metabolic products across the plasma and intracellular membranes. The ABC transporters tissue distribution reflects the complex physiological network of these proteins and reveals their important role in absorption, excretion, and distribution of drugs.

P-glycoprotein (P-gp) is a 170 kDa glycoprotein coded by ABCB1 gene mapped to chromosome 7q21.1. Its presence in normal tissue (as liver, kidney, brain, intestine, etc.) reveals the ability of modulating bioavailability and transport of wide range of substrates, especially organic molecules containing aromatic groups, and suggests its role in determining multidrug resistance (MDR) phenotype. P-gp is overexpressed in many tumors (thus causing intrinsic drug resistance) and the expression of P-gp can also be induced by chemotherapy (thus resulting in the acquired development of MDR) (Thomas and Coley 2003). Expression of P-gp fluctuates with elevated expression level in untreated cancer into higher level upon relapse after chemotherapy and undetectable or low level in the expression in drug sensitive tumors. P-gp expression has been reported in 40% of breast cancer (Kao et al. 2001), 20% of ovarian cancer (Baekelandt et al. 2000), and in more than 50% of patients with acute myelogenous leukemia experiencing relapse (Leith et al. 1999) and is now associated with treatment failure in kidney, liver, and colon cancer (Ambudkar et al. 2003). It confers the strongest resistance to the widest variety of compounds including vinca alkaloids, anthracyclines, and taxanes contributing to the failure of chemotherapy (Szakács et al. 2006). Recent reports have suggested that molecularly targeted therapies, such as some kinases inhibitors, are also substrates for drug efflux proteins (Shervington and Lu 2008).

MRP1 is a transporter involved in glutathione-linked organic compound transport and is normally expressed in the basolateral membrane of cells from testis, kidney, oropharyngeal mucosa, etc. Recently the overexpression of MRP1 has been correlated with chemoresistance in prostate, lung, and breast cancer (Holohan et al. 2013) and confers resistance to several hydrophobic compounds that are also P-gp substrates.

Screens carried out with the NCI60 cell panel indicate that there is a strong correlation between the expression of several transporter (including ABC superfamily) and decrease in chemosensitivity.

The goal of reversing MDR in the clinic through the pharmacological inhibition of ABC transporter has been pursued for years and different generation of inhibitors have been developed. Only P-gp inhibitors have been evaluated in clinical studies. However, the results of clinical trials using modulators of P-gp have been disappointing mainly because of the required marked reduction of the anticancer drug doses. In addition it is now very clear that resistance is not only due to the

overexpression of P-gp, but also to many other mechanisms, thus making unrealistic to reverse tumor resistance by pump inhibition.

The resistance of tumors originating from tissues expressing high levels of P-gp (such as colon, kidney, or the adrenocortex) often extends to drugs that are not subject to P-gp-mediated transport, suggesting that “intrinsically resistant” cancer is also protected by non-Pgp-mediated mechanisms (Szakács et al. 2006).

Evidence linking P-gp expression with poor clinical outcome is more conclusive for breast cancer, sarcoma, and certain types of leukemia. P-gp expression in patients with AML has consistently been associated with reduced chemotherapy response rates and poor survival (Pallis and Russell 2004), in contrast with MRP1 whose expression is not a significant factor in drug resistance in AML (Leith et al. 1999).

The expression of these transporters as a mechanism of acquired resistance after a specific treatment is also responsible for the phenomenon of cross-resistance that must be taken into account especially for a clinical perspective.

### 2.3 DNA Damage Repair

The cellular response to DNA damage involves several mechanisms aimed at repairing the damage, thus restoring DNA integrity or alternatively activating cell death pathways.

Many chemotherapeutic drugs induce DNA damage either directly (for example, platinum-based drugs) or indirectly (for example, topoisomerase inhibitors). Without repair, this damage can result in genetic instability, acquisition of further mutations, and cytotoxic effects (Holohan et al. 2013). The different sensitivity to DNA damaging agents can be due to a different ability to repair drug-induced damage. For example, cancer cells deficient in Homologous Recombination (HR), e.g., because of mutations of BRCA1 or BRCA2 genes, are very sensitive to DNA damage caused by platinum drugs. HR deficient cancers are very sensitive to PARP inhibitors that cause an accumulation of DNA single strand breaks. In HR proficient cells, DNA single strand breaks generate DNA double strand breaks during DNA replication that are repaired before mitosis. Instead in HR deficient cells the inhibition of base excision repair by PARP inhibitors leads to unreparable DNA double strand breaks with consequent cell death. This is the concept of synthetic lethality whose application has shown promising results in patients with breast and ovarian tumors (Farmer et al. 2005). Even in this case resistance can occur because of the acquisition of secondary mutations restoring HR normal function (Ashworth 2008).

Other pathways involved in DNA repair that have been reported to be relevant for drug resistance are nucleotide-excision repair (NER) and mismatch repair (MMR). The first one is required for the repair of DNA damage caused by many DNA-damaging drugs, such as platinum-based drugs. High expression of a key component, excision repair cross-complementing 1 (ERCC1), has been linked with poor responses to chemotherapy in non-small-cell lung carcinoma (NSCLC),



gastric, and ovarian cancer (Kirschner and Melton 2010). Notably, testicular cancers, which are very sensitive to cisplatin treatment, have very low levels of ERCC1 (Usanova et al. 2010).

The mismatch repair (MMR) system, whose key players are MLH1 and MSH2 genes, is crucial for maintaining genomic integrity, and its deficiency has been linked to high mutation rate of a large number of genes including those involved in immunogenicity and drug sensitivity. This explains why hypermethylation of MLH1 causes resistance to cisplatin and carboplatin (Fink et al. 1998). More recently, a synthetic lethal interaction between MSH2-targeted short interfering RNA (siRNA) and methotrexate was identified in MMR-deficient cancer cells; methotrexate caused the accumulation of oxidative lesions such as 8-oxoguanine (8-oxoG) in MSH2-deficient cells, which resulted in a loss of viability through apoptosis. This has led to an on-going Phase II clinical trial with methotrexate in patients with MSH2-deficient metastatic colorectal cancer using measurement of 8-oxoG lesions as a biomarker (Holohan et al. 2013).

Another DNA protein involved in resistance is *O*-6-methylguanine-DNA-methyl transferase (MGMT), whose overexpression is responsible for the resistance to methylating agents such as temozolomide. The hypomethylation of the promoter of the MGMT gene has been associated with resistance of human glioblastoma to temozolomide (Thomas et al. 2017).

## 2.4 Deregulation of Apoptosis

The resistance to anticancer drugs can be due to the fact that in cancer cells, apoptotic pathways are frequently dysfunctional. Numerous intrinsic adaptive responses can be triggered and promote survival of cancer cells exposed to DNA-damaging agents. Anti-apoptotic BCL-2 family members, inhibitor of apoptosis proteins (IAPs), and the caspases are key proteins of this mechanism. Mutations, amplifications, chromosomal translocations, and overexpression of the genes encoding for these proteins have been associated with various malignancies and linked to resistance to chemotherapy and targeted therapies.

BCL-2 family proteins have a pivotal role in dictating cell fate following chemotherapy treatment. The balance between the anti-apoptotic BCL-2 family members (such as BCL-XL and MCL1) and pro-apoptotic family members (such as BAX, BAD, and BAK, as well as various BH3-only proteins) is critical in determining the activation or not of a common pathway generally stimulated from all cytotoxic drugs that causes the mitochondrial outer membrane permeabilization (MOMP) and finally cell death. Overexpression of one or more anti-apoptotic proteins or underexpression of one or more pro-apoptotic proteins or a combination of both dysregulate apoptosis. For example, the overexpression of Bcl-2 protected prostate cancer cells from apoptosis while led to inhibition of TRAIL-induced apoptosis in neuroblastoma, glioblastoma, and breast carcinoma cells. In colorectal cancers bax(G)8 frameshift mutations could contribute to resistance of cells to anticancer treatments. In the case of chronic lymphocytic leukemia (CLL), the

malignant cells have an anti-apoptotic phenotype with high levels of anti-apoptotic Bcl-2 and low levels of pro-apoptotic proteins such as Bax (Wong 2011). The IAPs are a group of structurally and functionally similar proteins that regulate apoptosis by inhibiting caspase activity and promoting degradation of active caspases. Dysregulated IAP expression has been reported in many cancers. For example, drug resistance correlated with the expression of cIAP-2 in pancreatic cells while Apollon was found to be upregulated in gliomas and was responsible for cisplatin and camptothecin resistance (Chen et al. 1999).

The caspases are a large group of proteins that play a central role in apoptosis. As they work at two levels, *initiator caspases* (e.g., caspase-2, -8, -9, and -10) responsible for the initiation of the apoptotic pathway and *effector caspases* (caspase-3, -6, and -7) responsible in the actual cleavage of cellular components during apoptosis. Therefore low levels of these proteins or impairment function can decrease apoptosis thus contributing to resistance.

The p53 protein, also called tumor protein 53 (or TP 53), is one of the best known tumor suppressor proteins whose mutation acquired oncogenic property. Defects in the p53 tumor suppressor gene have been linked to more than 50% of human cancers (Bai and Zhu 2006) because it is not only involved in the induction of apoptosis but it is also a key player in cell cycle regulation, development, differentiation, gene amplification, DNA recombination, chromosomal segregation, and cellular senescence. A recent study reported that some target genes of p53 involved in apoptosis and cell cycle regulation are aberrantly expressed in melanoma cells and possibly related to resistance (Avery-Kiejda et al. 2011).

Several abnormalities in the death signaling pathways that can lead to evasion of the extrinsic pathway of apoptosis have been identified. Death receptors such as Fas, DR3, Trail-1, Trail-2, and ligands of the death receptors are key players in the regulation of extrinsic pathway of apoptosis. Downregulation or impairment of receptor function, abnormal expression of decoy receptors, as well as a reduced level in the death signals can contribute to impaired signaling and hence a reduction of apoptosis and acquisition of drug resistance.

## 2.5 Alteration of Drug Target

One of the most common drug resistance mechanisms involves genetic alterations of drug target such as mutations or changes in expression levels. These modifications could exist at low levels before drug treatment and undergo positive selection during exposure to chemotherapy. In oncology, genomic characterization of cancer has highlighted the importance of driver somatic mutations that give rise to an unusual reliance of cancer cells on a particular molecular pathways and their specific oncogenic kinases. These proteins are targets for many drugs and can be altered by different mechanisms.

Main categories of this type of resistance could be:

- Downregulation of gene expression (e.g., effect of doxorubicin on topoisomerase II $\alpha$ ) (Di Nicolantonio et al. 2005)
- Gene amplification (e.g., BCR-ABL amplification detected both in vitro and in imatinib-resistant CML specimens) (Gorre et al. 2001)
- Mutations in *gatekeeper residue* (e.g., EGFR-T790M in NSCLC after treatment with tyrosine kinase inhibitor (TKI) gefitinib and erlotinib (Bell et al. 2005); BCR-ABL T315 in CML patient treated with imatinib) (Gorre et al. 2001)
- Mutations that alter the conformation (e.g., mutations in ALK – F1174L, C1156Y, and L1152R – confer an increase in ATP affinity and clinical resistance to the ALK inhibitor crizotinib) (Choi et al. 2010)
- Alternative spliced form (e.g., p61BRAF<sup>V600E</sup> detected in both vemurafenib in vitro-resistant cells and from resistant patient tumor biopsies produces enhanced dimerization with other RAF family members and resistance to vemurafenib but not MEK inhibitors) (Poulikakos et al. 2011).

A possible consequence of the inhibition of specific target is also the activation of a so-called bypass signaling mechanism. This results in the activation of a critical downstream signaling effector – normally activated by the kinase and extinguished by a kinase inhibitor – through a parallel mechanism that is indifferent to the kinase-directed therapy. An illustrative example of bypass-mediated resistance has been described in EGFR-mutant NSCLC. The bypass resistance mechanisms may also involve the modulation of positive or negative feedback loops and an example could be the augmentation of AKT signaling by MEK inhibitors (Garraway and Jänne 2012).

---

### 3 Conclusion

This review article shortly summarizes the main mechanisms of resistance to anticancer drugs. In order to define possible strategies to counteract resistance mechanisms, it can be useful to divide them in *pre-target*, *target related*, and *post-target* mechanisms.

As far as *pre-target* mechanisms – that can also be classified as pharmacokinetics mechanisms – we envisage the possibility to circumvent them by increasing drug doses and improving drug delivery testing combinations with modulators of drug transport and of tumor microenvironment.

On the other side, a deeper and deeper knowledge of *target related* mechanisms, e.g., target mutations, downregulation, amplification, is instrumental to identify alternative drugs, equally effective against cancer cells, to circumvent these types of resistance.

Finally as regards *post-target* mechanisms, the modulation of apoptosis appears to be a potentially feasible approach as recently demonstrated by Croce and Reed (2016).

## References

- Amable L (2016) Cisplatin resistance and opportunities for precision medicine. *Pharmacol Res* 106:27–36. doi:[10.1016/j.phrs.2016.01.001](https://doi.org/10.1016/j.phrs.2016.01.001)
- Ambudkar SV, Kimchi-Sarfaty C, Sauna ZE, Gottesman MM (2003) P-glycoprotein: from genomics to mechanism. *Oncogene* 22:7468–7485. doi:[10.1038/sj.onc.1206948](https://doi.org/10.1038/sj.onc.1206948)
- Ashworth A (2008) Drug resistance caused by reversion mutation. *Cancer Res* 68:10021–10023. doi:[10.1158/0008-5472.CAN-08-2287](https://doi.org/10.1158/0008-5472.CAN-08-2287)
- Avery-Kiejda KA, Bowden NA, Croft AJ et al (2011) P53 in human melanoma fails to regulate target genes associated with apoptosis and the cell cycle and may contribute to proliferation. *BMC Cancer* 11:203. doi:[10.1186/1471-2407-11-203](https://doi.org/10.1186/1471-2407-11-203)
- Baekelandt MM, Holm R, Nesland JM et al (2000) P-glycoprotein expression is a marker for chemotherapy resistance and prognosis in advanced ovarian cancer. *Anticancer Res* 20:1061–1067
- Bai L, Zhu WG (2006) p53: structure, function and therapeutic applications. *J Cancer Mol* 2:141–153
- Bell DW, Gore I, Okimoto RA et al (2005) Inherited susceptibility to lung cancer may be associated with the T790M drug resistance mutation in EGFR. *Nat Genet* 37:1315–1316. doi:[10.1038/ng1671](https://doi.org/10.1038/ng1671)
- Buck A, Halbritter S, Späth C et al (2015) Distribution and quantification of irinotecan and its active metabolite SN-38 in colon cancer murine model systems using MALDI MSI. *Anal Bioanal Chem* 407:2107–2116. doi:[10.1007/s00216-014-8237-2](https://doi.org/10.1007/s00216-014-8237-2)
- Cesca M, Morosi L, Berndt A et al (2016) Bevacizumab-induced inhibition of angiogenesis promotes a more homogeneous intratumoral distribution of paclitaxel, improving the antitumor response. *Mol Cancer Ther* 15:125–135. doi:[10.1158/1535-7163.MCT-15-0063](https://doi.org/10.1158/1535-7163.MCT-15-0063)
- Chen Z, Naito M, Hori S et al (1999) A human IAP-family gene, apollon, expressed in human brain cancer cells. *Biochem Biophys Res Commun* 264:847–854. doi:[10.1006/bbrc.1999.1585](https://doi.org/10.1006/bbrc.1999.1585)
- Choi YL, Soda M, Yamashita Y et al (2010) EML4-ALK mutations in lung cancer that confer resistance to ALK inhibitors. *N Engl J Med* 363:1734–1739. doi:[10.1056/NEJMoa1007478](https://doi.org/10.1056/NEJMoa1007478)
- Connell JJ, Sugihara Y, Török S et al (2015) Localization of sunitinib in in vivo animal and in vitro experimental models by MALDI mass spectrometry imaging. *Anal Bioanal Chem* 407:2245–2253. doi:[10.1007/s00216-014-8350-2](https://doi.org/10.1007/s00216-014-8350-2)
- Croce CM, Reed JC (2016) Finally, an apoptosis-targeting therapeutic for cancer. *Cancer Res* 76:5914–5920. doi:[10.1158/0008-5472.CAN-16-1248](https://doi.org/10.1158/0008-5472.CAN-16-1248)
- Di Nicolantonio F, Mercer SJ, Knight LA et al (2005) Cancer cell adaptation to chemotherapy. *BMC Cancer* 5:78. doi:[10.1186/1471-2407-5-78](https://doi.org/10.1186/1471-2407-5-78)
- Farmer H, McCabe N, Lord CJ et al (2005) Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature* 434:917–921. doi:[10.1038/nature03445](https://doi.org/10.1038/nature03445)
- Fink D, Aebi S, Howell SB (1998) The role of DNA mismatch repair in drug resistance. *Clin Cancer Res* 4:1–6
- Fuso Nerini I, Morosi L, Zucchetti M et al (2014) Intratumor heterogeneity and its impact on drug distribution and sensitivity. *Clin Pharmacol Ther* 96:224–238. doi:[10.1038/clpt.2014.105](https://doi.org/10.1038/clpt.2014.105)
- Garraway LA, Jänne PA (2012) Circumventing cancer drug resistance in the era of personalized medicine. *Cancer Discov* 2:214–226. doi:[10.1158/2159-8290.CD-12-0012](https://doi.org/10.1158/2159-8290.CD-12-0012)
- Giordano S, Morosi L, Veglianesi P et al (2016a) 3D mass spectrometry imaging reveals a very heterogeneous drug distribution in tumors. *Sci Rep* 6:37027. doi:[10.1038/srep37027](https://doi.org/10.1038/srep37027)
- Giordano S, Zucchetti M, Decio A et al (2016b) Heterogeneity of paclitaxel distribution in different tumor models assessed by MALDI mass spectrometry imaging. *Sci Rep* 6:39284. doi:[10.1038/srep39284](https://doi.org/10.1038/srep39284)
- Gorre ME, Mohammed M, Ellwood K et al (2001) Clinical resistance to STI-571 cancer therapy caused by BCR-ABL gene mutation or amplification. *Science* 293:876–880. doi:[10.1126/science.1062538](https://doi.org/10.1126/science.1062538)

- Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG (2013) Cancer drug resistance: an evolving paradigm. *Nat Rev Cancer* 13:714–726. doi:[10.1038/nrc3599](https://doi.org/10.1038/nrc3599)
- Kao CH, Tsai SC, Liu TJ et al (2001) P-glycoprotein and multidrug resistance-related protein expressions in relation to technetium-99m methoxyisobutylisonitrile scintimammography findings. *Cancer Res* 61:1412–1414
- Kauvar LM, Morgan AS, Sanderson PE, Henner WD (1998) Glutathione based approaches to improving cancer treatment. *Chem Biol Interact* 111–112:225–238
- Kirschner K, Melton DW (2010) Multiple roles of the ERCC1-XPF endonuclease in DNA repair and resistance to anticancer drugs. *Anticancer Res* 30:3223–3232
- Leith CP, Kopecky KJ, Chen IM et al (1999) Frequency and clinical significance of the expression of the multidrug resistance proteins MDR1/P-glycoprotein, MRP1, and LRP in acute myeloid leukemia: a southwest oncology group study. *Blood* 94:1086–1099
- Longley DB, Harkin DP, Johnston PG (2003) 5-fluorouracil: mechanisms of action and clinical strategies. *Nat Rev Cancer* 3:330–338. doi:[10.1038/nrc1074](https://doi.org/10.1038/nrc1074)
- Meijer C, Mulder NH, Timmer-Bosscha H et al (1992) Relationship of cellular glutathione to the cytotoxicity and resistance of seven platinum compounds. *Cancer Res* 52:6885–6889
- Minchinton AI, Tannock IF (2006) Drug penetration in solid tumours. *Nat Rev Cancer* 6:583–592. doi:[10.1038/nrc1893](https://doi.org/10.1038/nrc1893)
- Morosi L, Zuchetti M, D’Incalci M, Davoli E (2013) Imaging mass spectrometry: challenges in visualization of drug distribution in solid tumors. *Curr Opin Pharmacol* 13:807–812. doi:[10.1016/j.coph.2013.06.003](https://doi.org/10.1016/j.coph.2013.06.003)
- Pallis M, Russell N (2004) Strategies for overcoming p-glycoprotein-mediated drug resistance in acute myeloblastic leukaemia. *Leukemia* 18:1927–1930. doi:[10.1038/sj.leu.2403511](https://doi.org/10.1038/sj.leu.2403511)
- Poulikakos PI, Persaud Y, Janakiraman M et al (2011) RAF inhibitor resistance is mediated by dimerization of aberrantly spliced BRAF(V600E). *Nature* 480:387–390. doi:[10.1038/nature10662](https://doi.org/10.1038/nature10662)
- Prideaux B, Stoeckli M (2012) Mass spectrometry imaging for drug distribution studies. *J Proteome* 75:4999–5013. doi:[10.1016/j.jprot.2012.07.028](https://doi.org/10.1016/j.jprot.2012.07.028)
- Rzagalinski I, Volmer DA (2016) Quantification of low molecular weight compounds by MALDI imaging mass spectrometry – a tutorial review. *Biochim Biophys Acta*. doi:[10.1016/j.bbapap.2016.12.011](https://doi.org/10.1016/j.bbapap.2016.12.011). pii:S1570-9639(16)30276-X [Epub ahead of print]
- Shervington A, Lu C (2008) Expression of multidrug resistance genes in normal and cancer stem cells. *Cancer Investig* 26:535–542. doi:[10.1080/07357900801904140](https://doi.org/10.1080/07357900801904140)
- Sugiura Y, Setou M (2010) Imaging mass spectrometry for visualization of drug and endogenous metabolite distribution: toward in situ pharmacometabolomes. *J Neuroimmune Pharmacol* 5:31–43. doi:[10.1007/s11481-009-9162-6](https://doi.org/10.1007/s11481-009-9162-6)
- Szakács G, Paterson JK, Ludwig JA et al (2006) Targeting multidrug resistance in cancer. *Nat Rev Drug Discov* 5:219–234. doi:[10.1038/nrd1984](https://doi.org/10.1038/nrd1984)
- Thomas H, Coley HM (2003) Overcoming multidrug resistance in cancer: an update on the clinical strategy of inhibiting p-glycoprotein. *Cancer Control J Moffitt Cancer Center* 10:159–165
- Thomas A, Tanaka M, Trepel J et al (2017) Temozolomide in the era of precision medicine. *Cancer Res* 77:823–826. doi:[10.1158/0008-5472.CAN-16-2983](https://doi.org/10.1158/0008-5472.CAN-16-2983)
- Torok S, Rezeli M, Kelemen O et al (2017) Limited tumor tissue drug penetration contributes to primary resistance against angiogenesis. *Inhibitors. Theranostics* 7(2):400–412. doi:[10.7150/thno.16767](https://doi.org/10.7150/thno.16767)
- Usanova S, Piée-Staffa A, Sied U et al (2010) Cisplatin sensitivity of testis tumour cells is due to deficiency in interstrand-crosslink repair and low ERCC1-XPF expression. *Mol Cancer* 9:248. doi:[10.1186/1476-4598-9-248](https://doi.org/10.1186/1476-4598-9-248)
- Wong RSY (2011) Apoptosis in cancer: from pathogenesis to treatment. *J Exp Clin Cancer Res CR* 30:87. doi:[10.1186/1756-9966-30-87](https://doi.org/10.1186/1756-9966-30-87)



# Major Physiological Signaling Pathways in the Regulation of Cell Proliferation and Survival

Huifang Tang and Gongda Xue

## Contents

1	Introduction .....	14
2	Signaling Pathways Involved in the Regulation of Cell Proliferation .....	14
2.1	Biological Regulation of Proliferation by mTOR/PI3K/Akt Signaling .....	15
2.2	Ras/MAPK Signaling-Regulated Cell Proliferation, Differentiation, and Survival ...	17
2.3	JAK/STAT Signaling in the Regulation of Cell Proliferation and Survival .....	20
3	Signaling Interaction Network Between PI3K/Akt, Ras/MAPK, and JAK/STAT Axes ...	20
4	Output Signals of Integrated Signaling Pathways .....	22
4.1	Redundant Targeting of Shared Substrates in Cell Growth and Metabolism .....	22
4.2	Differential Phospho-Regulation of Shared Substrates in Cell Proliferation and Survival .....	23
5	Impact of Signaling Interaction Network in Clinical Cancer Therapies .....	23
6	Final Remarks .....	24
	References .....	25

## Abstract

Multiple signaling pathways regulate cell proliferation and survival and are therefore important for maintaining homeostasis of development. The balance between cell growth and death is achieved through orchestrated signal transduction pathways mediated by complex functional interactions between signaling axes, among which, PI3K/Akt and Ras/MAPK as well as JAK/STAT play a dominant role in promoting cell proliferation, differentiation, and survival. In clinical cancer therapies, drug resistance is the major challenge that occurs in almost all targeted therapeutic strategies. Recent advances in research have suggested that the intrinsic

H. Tang (✉)

Department of Pharmacology, Zhejiang University, School of Basic Medical Sciences, Hangzhou 310058, China

e-mail: [tanghuifang@zju.edu.cn](mailto:tanghuifang@zju.edu.cn)

G. Xue

Department of Biomedicine, University Hospital Basel, Basel 4031, Switzerland

© Springer International Publishing AG 2017

M. Mandalà, E. Romano (eds.), *Mechanisms of Drug Resistance in Cancer Therapy*, Handbook of Experimental Pharmacology 249, DOI 10.1007/164\_2017\_4

13

pro-survival signaling crosstalk is the driving force in acquired resistance to a targeted therapy, which may be abolished by interfering with the cross-reacting network.

---

**Keywords**

Apoptosis • Cell cycle • Drug resistance • JAK/STAT • Lung cancer • Melanoma • mTOR/PI3K/Akt • Proliferation • Ras/MAPK • Signaling crosstalk

---

## 1 Introduction

The development of a multicellular eukaryote starts from the division of a zygote that eventually produces many types of functionally specialized cells. Proper tissue formation and growth requires a homeostatic regulation of cell cycle events that control cell proliferation and the cellular metabolism that often associates with apoptosis, a process of programmed cell death that surveils a balanced cell proliferation rate and cell number. Such regulation is usually governed by defined activation and inactivation of individual genes at different developmental stages. The selective pattern of gene expression functionally directs cell proliferation, cell differentiation, cell migration, and cell–cell communication to maintain a developmental integrity of the complex formation of tissues and organs. This phenotypic complexity is precisely mirrored by the genetic control system, an orchestrated signaling network coordinated among a few key evolutionarily conserved pathways. In this chapter, we intend to discuss recent advances of two important pathways, mTOR/PI3K/Akt and mitogen-activated protein kinase (MAPK), whose activation and interaction are critical for development and are the key regulatory elements in pathological circumstances such as malignant transformation of neoplasia and drug resistance in clinical cancer therapy.

---

## 2 Signaling Pathways Involved in the Regulation of Cell Proliferation

Cell proliferation is integrated with the overall needs of the organismal development in coordination with cell growth. It is principally mediated by cell cycle, a fundamental process that occurs when a parental cell produces two daughter cells via a series of genetic events (Hartwell and Kastan 1994; Nakayama and Nakayama 2006). Each of the daughter cells theoretically obtains the same copy of the genetic material inherited from the parental cell and are therefore genetically identical and morphologically undistinguishable. Similar to their parent, the mature daughter cells may initiate and complete the cell cycle when triggered by distinct signals in favor of the needs for growth and metabolism. Under physiological conditions, the proliferative cells are generally restricted to cells that supplement the tissue. Most tissues contain a group of multipotent cells, usually called stem cells, which can not only renew themselves but also divide asymmetrically to generate a new stem cell and a progenitor cell. In a context

dependent manner, progenitor cells may undergo further divisions, or enter terminal differentiation with endowed specialized functions. Thus, in order to maintain proper tissue development, cell proliferation is programmed to be tightly controlled and its deregulation is a hallmark of malignant transformation found in the clinic (Hanahan and Weinberg 2000, 2011).

Maintenance of an appropriate cell number is also contributed by programmed cell death, apoptosis, which actively eliminates unwanted cells including aged and damaged cells (Elmore 2007; Taylor et al. 2008). Importantly, these two cellular functions are under a variety of stringent controls to ensure tissue homeostasis. Aberrant alterations of each functional node, either hyperactivation of proliferation or deactivation of apoptosis, will ultimately result in uncontrolled cell proliferation and growth that leads to tumor formation.

Mechanistic studies have revealed a number of biological signaling axes important for controlling cell proliferation and apoptosis. These signaling pathways commonly form an integral signaling network through diverse functional interactions, which are mediated by specific feedback signaling loops. Recent advances from clinical studies have shown that therapeutic disruption of the indispensable signaling loops during tumorigenic development is one of the leading causes of drug resistance and is attributed to over-activation of parallel proliferative signaling pathways (Holohan et al. 2013; Wicki et al. 2016).

## 2.1 Biological Regulation of Proliferation by mTOR/PI3K/Akt Signaling

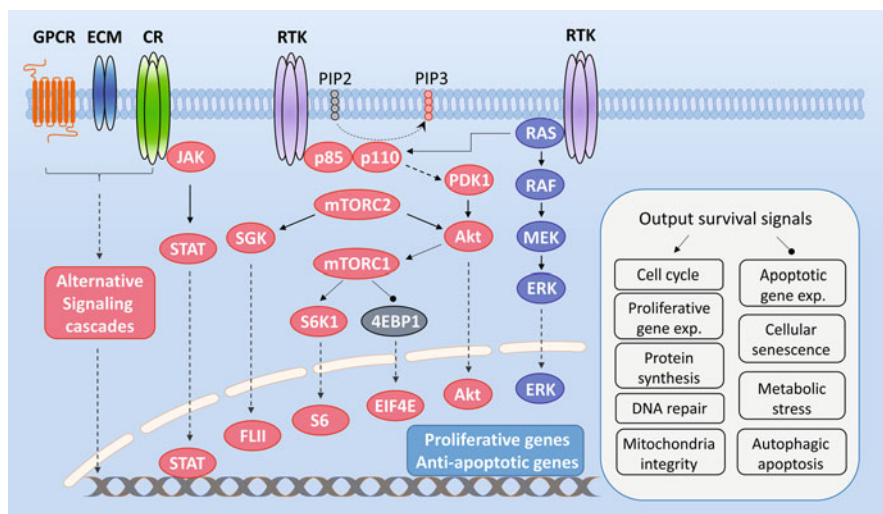
In response to the stimulation of a variety of extracellular signaling cues, a number of membrane receptors with kinase activities can be activated via oligomerization. Activated membranous kinases subsequently create binding motifs in their intracellular segments that recruit and further activate the essential signal-mediator kinases. One of these intracellular messengers is lipid kinase phosphoinositide 3 kinase (PI3K), an evolutionarily conserved intracellular kinase that can convert Phosphatidylinositol (4,5)-bisphosphate (PIP2) into the secondary messenger molecule Phosphatidylinositol (3,4,5)-trisphosphate (PIP3) on the plasmic membrane (Engelman et al. 2006). This process can be reversed by the phosphatase and tensin homolog (PTEN), which pairs with PI3K to constitute a functional switch of PIP3-dependent signaling transduction. Membrane-targeted PIP3 recruits certain proto-kinases to the inner surface of cytoplasmic membrane through binding to the pleckstrin homology (PH) domain with elevated affinity. Several important kinases are dependent on PIP3-mediated membrane targeting for activation. Akt and phosphoinositide-dependent kinase 1 (PDK1), two PH-domain-containing kinases, recognize and bind to newly generated and enriched PIP3, resulting in transient membrane anchoring and consequent activation.

Akt, also called protein kinase B (PKB), is a serine/threonine kinase that belongs to the AGC protein kinase family (Pearce et al. 2010). It comprises three isoforms in mammals with a high degree of sequence identity. Although the tissue distribution pattern is slightly different, Akt isoforms are virtually expressed in all cell types and



act as one of the most important signaling nodes to direct proper responses to exogenous or endogenous stimuli (Fayard et al. 2010).

Activation of Akt is a multistep process with characteristic phosphorylation of two key amino acids, threonine 308 (T308) in the “activation loop” and serine 473 (S473) in the “carboxy-terminal hydrophobic motif,” both of which are conserved in all three isoforms. T308 phosphorylation is contributed by PDK1 when both proteins are recruited to the plasma membrane by PIP3. Although not exclusively required, it is generally accepted that the full kinase activity of Akt is stimulated upon phosphorylation of S473 by the mammalian target of rapamycin (mTOR) complex 2 (mTORC2) (Cybulski and Hall 2009). Recent studies have also uncovered another kinase, DNA-dependent protein kinase (DNA-PK), that is capable of phosphorylating Akt on the same amino acid in response to DNA double-strand break (Bozulic et al. 2008; Surucu et al. 2008). Therefore, under physiological or pathological conditions, Akt acts as a direct functional target downstream of PI3K under the stringent control of PDK1 and mTORC2 or DNA-PK, which consolidates the theoretical basis for understanding the current model (Fig. 1). Genetic knockout of each Akt isoform or pharmacological inhibition of its activity results in a universal defect in growth in mouse models, indicating a central role of the Akt family in growth control (Fayard et al. 2010). In consistence with its roles in development, Akt is widely observed to be hyperactivated in a broad panel of cancers (Fruman and Rommel 2014; Vivanco and Sawyers 2002). In this regard, the signaling axis mTOR/PI3K/Akt has become one of the most attractive targeting pathways to restrain cancer cell proliferation. In spite of its three major upstream regulators, other kinases such as serine–threonine kinases IKK (Xie



**Fig. 1** Introduction to major proliferative signaling pathways. Three major signaling axes, mTOR/PI3K/Akt, Ras/MAPK, and JAK/STAT, are dissected to show their downstream targets that promote cell cycle progression. *RTK* receptor tyrosine kinase, *CR* chemokine receptor, *ECM* extracellular matrix, *GPCR* G-protein-coupled receptor