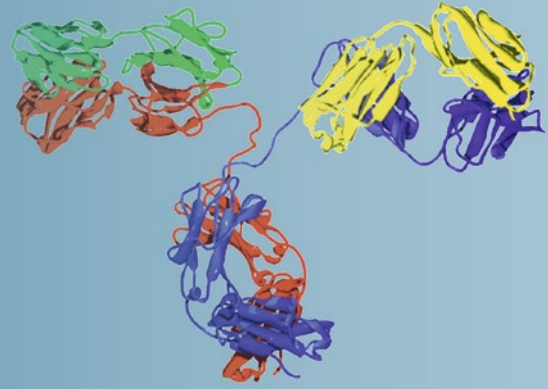


Methods in
Molecular Biology 1348

Springer Protocols



Gunnar Houen *Editor*

Peptide Antibodies

Methods and Protocols

 Humana Press

METHODS IN MOLECULAR BIOLOGY

Series Editor
John M. Walker
School of Life and Medical Sciences
University of Hertfordshire
Hatfield, Hertfordshire, AL10 9AB, UK

For further volumes:
<http://www.springer.com/series/7651>

Peptide Antibodies

Methods and Protocols

Edited by

Gunnar Houen

Statens Serum Institut, Copenhagen, Denmark

 **Humana Press**

Editor

Gunnar Houen
Statens Serum Institut
Copenhagen, Denmark

ISSN 1064-3745 ISSN 1940-6029 (electronic)
Methods in Molecular Biology
ISBN 978-1-4939-2998-6 ISBN 978-1-4939-2999-3 (eBook)
DOI 10.1007/978-1-4939-2999-3

Library of Congress Control Number: 2015948252

Springer New York Heidelberg Dordrecht London
© Springer Science+Business Media New York 2015

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, express or implied, with respect to the material contained herein or for any errors or omissions that may have been made.

Cover Illustration: Image provided by Tim Vickers.

Printed on acid-free paper

Humana Press is a brand of Springer
Springer Science+Business Media LLC New York is part of Springer Science+Business Media (www.springer.com)

Preface

The ability to design and produce peptide antibodies has had a large impact on molecular cell biology and immunology, including all the different techniques involved (immunoassays, immunoprecipitation, immunoblotting, immunohistochemistry, etc).

This large impact is a result of several technical and scientific advances: solid phase peptide synthesis, peptide carrier conjugation and immunization, genomics, transcriptomics, proteomics and elucidation of the molecular basis of antigen presentation and recognition by dendritic cells, macrophages, B cells, and T cells.

Moreover, although peptide antibodies have been available for many years, they continue to be a field of active research and method development. For example, peptide antibodies which are dependent on specific posttranslational modifications are of great interest (phosphorylation, citrullination, etc.) and different forms of recombinant peptide antibodies are gaining interest (nanobodies, single chain antibodies, TCR-like antibodies, etc.).

This volume covers basic and advanced aspects of peptide antibody production, characterization, and uses.

I thank all contributors and editorial staff for their work, especially the series editor, John Walker. Also, I want to thank all my collaborators and students throughout the years.

Copenhagen, Denmark

Gunnar Houen

Contents

<i>Preface</i>	<i>v</i>
<i>Contributors</i>	<i>xi</i>
1 Peptide Antibodies: Past, Present, and Future <i>Gunnar Houen</i>	1
2 The Structure of Natural and Recombinant Antibodies <i>Hui Ma and Richard O’Kennedy</i>	7
3 Prediction of Antigenic B and T Cell Epitopes via Energy Decomposition Analysis: Description of the Web-Based Prediction Tool BEPPE <i>Claudio Peri, Oscar C. Solé, Dario Corrada, Alessandro Gori, Xavier Daura, and Giorgio Colombo</i>	13
4 Prediction of Antibody Epitopes <i>Morten Nielsen and Paolo Marcatili</i>	23
5 Fmoc Solid-Phase Peptide Synthesis <i>Paul R. Hansen and Alberto Oddo</i>	33
6 Peptide-Carrier Conjugation <i>Paul R. Hansen</i>	51
7 Solid-Phase Peptide-Carrier Conjugation <i>Gunnar Houen and Dorte T. Olsen</i>	59
8 Analysis of Peptides and Conjugates by Amino Acid Analysis <i>Peter Højrup</i>	65
9 Characterization of Synthetic Peptides by Mass Spectrometry <i>Bala K. Prabhala, Osman Mirza, Peter Højrup, and Paul R. Hansen</i>	77
10 Interpretation of Tandem Mass Spectrometry (MSMS) Spectra for Peptide Analysis <i>Karin Hjerno and Peter Højrup</i>	83
11 Polyclonal Peptide Antisera <i>Tina H. Pihl, Kristin E. Illigen, and Gunnar Houen</i>	103
12 Production and Screening of Monoclonal Peptide Antibodies <i>Nicole Hartwig Trier, Anne Mortensen, Annette Schiolborg, and Tina Friis</i>	109
13 Production of Epitope-Specific Antibodies by Immunization with Synthetic Epitope Peptide Formulated with CpG-DNA-Liposome Complex Without Carriers <i>Dongbum Kim, Younghee Lee, and Hyung-Joo Kwon</i>	127

14	Thioredoxin-Displayed Multipetide Immunogens	137
	<i>Angelo Bolchi, Elena Canali, Andrea Santoni, Gloria Spagnoli, Daniele Viarisio, Rosita Accardi, Massimo Tommasino, Martin Müller, and Simone Ottonello</i>	
15	The Purification of Natural and Recombinant Peptide Antibodies by Affinity Chromatographic Strategies	153
	<i>Hui Ma and Richard O’Kennedy</i>	
16	Isolation of Camelid Single-Domain Antibodies Against Native Proteins Using Recombinant Multivalent Peptide Ligands	167
	<i>Norah A. Alturki, Kevin A. Henry, C. Roger MacKenzie, and Mehdi Arbabi-Ghahroudi</i>	
17	Generation of TCR-Like Antibodies Using Phage Display	191
	<i>Brian H. Santich, Hong Liu, Cheng Liu, and Nai-Kong V. Cheung</i>	
18	Structural Characterization of Peptide Antibodies	205
	<i>Anna Chailyan and Paolo Marcatili</i>	
19	Automated High-Throughput Mapping of Linear B-Cell Epitopes Using a Statistical Analysis of High-Density Peptide Microarray Data	215
	<i>Thomas Østerbye and Søren Buus</i>	
20	Characterization of Peptide Antibodies by Epitope Mapping Using Resin-Bound and Soluble Peptides	229
	<i>Nicole Hartwig Trier</i>	
21	Screening and Characterization of Linear B-Cell Epitopes by Biotinylated Peptide Libraries	241
	<i>Ida Rosenkrands and Anja Olsen</i>	
22	Bead-Based Peptide Arrays for Profiling the Specificity of Modification State-Specific Antibodies	251
	<i>Angela Filomena, Yvonne Beiter, Markus F. Templin, Thomas O. Joos, Nicole Schneiderhan-Marra, and Oliver Poetz</i>	
23	Surface Plasmon Resonance Method to Evaluate Anti-citrullinated Protein/Peptide Antibody Affinity to Citrullinated Peptides	267
	<i>Feliciano Real-Fernández, Giada Rossi, Filomena Panza, Federico Pratesi, Paola Migliorini, and Paolo Rovero</i>	
24	Specificity Analysis of Histone Modification-Specific Antibodies or Reading Domains on Histone Peptide Arrays	275
	<i>Goran Kungulovski, Ina Kycia, Rebekka Mauser, and Albert Jeltsch</i>	
25	Prion-Specific Antibodies Produced in Wild-Type Mice	285
	<i>Peter M.H. Heegaard, Ann-Louise Bergström, Heidi Gertz Andersen, and Henriette Cordes</i>	
26	Immunoblotting with Peptide Antibodies: Differential Immunoreactivities Caused by Certain Amino Acid Substitutions in a Short Peptide and Possible Effects of Differential Refolding of the Peptide on a Nitrocellulose or PVDF Membrane	303
	<i>Takenori Yamamoto, Taisuke Matsuo, Atsushi Yamamoto, Ryohei Yamagoshi, Kazuto Ohkura, Masatoshi Kataoka, and Yasuo Shinohara</i>	

27	Immunocytochemical and Immunohistochemical Staining with Peptide Antibodies	311
	<i>Tina Friis, Klaus Boberg Pedersen, David Hougaard, and Gunnar Houen</i>	
28	Designing B-Cell Epitopes for Immunotherapy and Subunit Vaccines	327
	<i>Harinder Singh, Sudbeer Gupta, Ankur Gautam, and Gajendra P.S. Raghava</i>	
29	Enterovirus-Specific Anti-peptide Antibodies.	341
	<i>Chit Laa Poh, Katherine Kirk, Hui Na Chua, and Lara Grollo</i>	
30	Therapeutic HIV Peptide Vaccine	351
	<i>Anders Fomsgaard</i>	
	<i>Index</i>	359

Contributors

- ROSITA ACCARDI • *Infections and Cancer Biology Group, International Agency for Research on Cancer-World Health Organization, Lyon, France*
- NORAH A. ALTURKI • *Human Health Therapeutics Portfolio, National Research Council Canada, Ottawa, ON, Canada; College of Applied Medical Sciences, King Saud University, Riyadh, Saudi Arabia*
- HEIDI GERTZ ANDERSEN • *Innate Immunology Group, Section for Immunology and Vaccinology, National Veterinary Institute, Technical University of Denmark, Frederiksberg, Denmark*
- MEHDI ARBABI-GHAHROUDI • *Human Health Therapeutics Portfolio, National Research Council Canada, Ottawa, ON, Canada; School of Environmental Sciences, University of Guelph, Guelph, ON, Canada; Department of Biology, Carleton University, Ottawa, ON, Canada*
- ANN-LOUISE BERGSTRÖM • *Innate Immunology Group, Section for Immunology and Vaccinology, National Veterinary Institute, Technical University of Denmark, Frederiksberg, Denmark; Department Neurodegeneration, H. Lundbeck A/S, Valby, Denmark*
- YVONNE BEITER • *Natural and Medical Sciences Institute at the University of Tuebingen, Reutlingen, Germany*
- ANGELO BOLCHI • *Biochemistry and Molecular Biology Unit, Department of Life Sciences, University of Parma, Parma, Italy*
- SØREN BUUS • *Laboratory of Experimental Immunology, Faculty of Health Sciences, Department of Immunology and Microbiology, University of Copenhagen, Copenhagen, Denmark*
- ELENA CANALI • *Biochemistry and Molecular Biology Unit, Department of Life Sciences, University of Parma, Parma, Italy*
- ANNA CHAILYAN • *Department of Biochemistry and Molecular Biology, University of Southern Denmark, Odense, Denmark*
- NAI-KONG V. CHEUNG • *Gerstner Sloan Kettering Graduate School of Biomedical Sciences and Department of Pediatrics, Memorial Sloan-Kettering Cancer Center, New York, NY, USA*
- HUI NA CHUA • *Sunway University, Kuala Lumpur, Malaysia*
- GIORGIO COLOMBO • *Department of Computational Biology, Institute for Molecular Recognition Chemistry (ICRM), Italian National Research Council, Milan, Italy*
- HENRIETTE CORDES • *Innate Immunology Group, Section for Immunology and Vaccinology, National Veterinary Institute, Technical University of Denmark, Frederiksberg, Denmark; Novo Nordisk A/S, Bagsvaerd, Denmark*
- DARIO CORRADA • *Institute for Molecular Recognition Chemistry (ICRM), Italian National Research Council, Milan, Italy; Department of Earth and Environmental Sciences, University of Milano-Bicocca, Milan, Italy*
- XAVIER DAURA • *Institut de Biociències i de Biomedicina (IBB), Universitat Autònoma de Barcelona (UAB), Barcelona, Spain; Catalan Institution for Research and Advanced Studies (ICREA), Barcelona, Spain*

- ANGELA FILOMENA • *Natural and Medical Sciences Institute at the University of Tuebingen, Reutlingen, Germany*
- ANDERS FOMSGAARD • *Virus R&D Laboratory, Department of Microbiology Diagnostics and Virology, Statens Serum Institut, Copenhagen, Denmark; Infectious Disease Research Unit, Clinical Institute, University of Southern Denmark, Odense, Denmark*
- TINA FRIIS • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- ANKUR GAUTAM • *Bioinformatics Centre, CSIR-Institute of Microbial Technology, Chandigarh, India*
- ALESSANDRO GORI • *Department of Computational Biology, Institute for Molecular Recognition Chemistry (ICRM), Italian National Research Council, Milan, Italy*
- LARA GROLO • *Swinburne University, Melbourne, Australia*
- SUDHEER GUPTA • *Bioinformatics Centre, CSIR-Institute of Microbial technology, Chandigarh, India*
- PAUL R. HANSEN • *Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark*
- PETER M.H. HEEGAARD • *Innate Immunology Group, Section for Immunology and Vaccinology, National Veterinary Institute, Technical University of Denmark, Frederiksberg, Denmark*
- KEVIN A. HENRY • *Human Health Therapeutics Portfolio, National Research Council Canada, Ottawa, ON, Canada*
- KARIN HJERNØ • *Department of Biochemistry and Molecular Biology, University of Southern Denmark, Odense, Denmark*
- PETER HØJRUP • *Department of Biochemistry and Molecular Biology, University of Southern Denmark, Odense, Denmark*
- GUNNAR HOUEN • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- DAVID HOUGAARD • *Department of Congenital Diseases, Statens Serum Institut, Copenhagen, Denmark*
- KRISTIN E. ILLIGEN • *Department of Quality Control, Statens Serum Institut, Copenhagen, Denmark*
- ALBERT JELTSCH • *Institute of Biochemistry, Faculty of Chemistry, University Stuttgart, Stuttgart, Germany*
- THOMAS O. JOOS • *Natural and Medical Sciences Institute at the University of Tuebingen, Reutlingen, Germany*
- MASATOSHI KATAOKA • *Biomarker Analysis Research Group, Health Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Takamatsu, Japan*
- DONGBUM KIM • *Department of Microbiology, College of Medicine, Hallym University, Gangwon-do, Republic of Korea*
- KATHERINE KIRK • *Swinburne University, Melbourne, Australia*
- GORAN KUNGULOVSKI • *Institute of Biochemistry, Faculty of Chemistry, University Stuttgart, Stuttgart, Germany*
- HYUNG-JOO KWON • *Department of Microbiology and Center for Medical Science Research, College of Medicine, Hallym University, Gangwon-do, Republic of Korea*
- INA KYCIA • *The Jackson Laboratory for Genomic Medicine, Farmington, CT, USA*
- YOUNGHEE LEE • *Department of Biochemistry, College of Natural Sciences, Chungbuk National University, Chungbuk, Republic of Korea*

- HONG LIU • *Eureka Therapeutics, Emeryville, CA, USA*
- CHENG LIU • *Eureka Therapeutics, Emeryville, CA, USA*
- HUI MA • *School of Biotechnology and Biomedical Diagnostics Institute, Dublin City University, Dublin, Ireland*
- C. ROGER MACKENZIE • *Human Health Therapeutics Portfolio, National Research Council Canada, Ottawa, ON, Canada; School of Environmental Sciences, University of Guelph, Guelph, ON, Canada*
- PAOLO MARCATILI • *Department of Systems Biology, Center for Biological Sequence Analysis, Technical University of Denmark, Lyngby, Denmark*
- TAISUKE MATSUO • *Institute for Genome Research and Faculty of Pharmaceutical Sciences, University of Tokushima, Tokushima, Japan*
- REBEKKA MAUSER • *Institute of Biochemistry, Faculty of Chemistry, University Stuttgart, Stuttgart, Germany*
- PAOLA MIGLIORINI • *Clinical Immunology and Allergy Unit, Department of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy*
- OSMAN MIRZA • *Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark*
- ANNE MORTENSEN • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- MARTIN MÜLLER • *German Cancer Research Center, Heidelberg, Germany*
- MORTEN NIELSEN • *Department of Systems Biology, Center for Biological Sequence Analysis, Technical University of Denmark, Lyngby, Denmark; Instituto de Investigaciones Biotecnológicas, Universidad Nacional de San Martín, Buenos Aires, Argentina*
- RICHARD O'KENNEDY • *School of Biotechnology and Biomedical Diagnostics Institute, Dublin City University, Dublin, Ireland*
- ALBERTO ODDO • *Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark*
- KAZUTO OHKURA • *Faculty of Pharmaceutical Science, Suzuka University of Medical Science, Suzuka, Japan*
- DORTHE T. OLSEN • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- ANJA OLSEN • *Department of Infectious Disease Immunology, Statens Serum Institut, Copenhagen, Denmark*
- THOMAS ØSTERBYE • *Laboratory of Experimental Immunology, Faculty of Health Sciences, Department of Immunology and Microbiology, University of Copenhagen, Copenhagen, Denmark*
- SIMONE OTTONELLO • *Biochemistry and Molecular Biology Unit, Department of Life Sciences, University of Parma, Parma, Italy; Dipartimento di Bioscienze, Università di Parma, Parma, Italy*
- FILomena PANZA • *Clinical Immunology and Allergy Unit, Department of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy*
- KLAUS BOBERG PEDERSEN • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- CLAUDIO PERI • *Department of Computational Biology, Institute for Molecular Recognition Chemistry (ICRM), Italian National Research Council, Milan, Italy*
- TINA H. PIHL • *Department of Large Animal Sciences, Medicine, and Surgery, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark*

- OLIVER POETZ • *Natural and Medical Sciences Institute at the University of Tuebingen, Reutlingen, Germany*
- CHIT LAA POH • *Sunway University, Kuala Lumpur, Malaysia*
- BALA K. PRABHALA • *Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark*
- FEDERICO PRATESI • *Clinical Immunology and Allergy Unit, Department of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy*
- GAJENDRA P.S. RAGHAVA • *Bioinformatics Centre, CSIR-Institute of Microbial Technology, Chandigarh, India*
- FELICIANA REAL-FERNÁNDEZ • *Laboratory of Peptide and Protein Chemistry and Biology, Division of Pharmaceutical Sciences and Nutraceutic, Department of NeuroFarBa, University of Florence, Sesto Fiorentino, Italy*
- IDA ROSENKRANDS • *Department of Infectious Disease Immunology, Statens Serum Institut, Copenhagen, Denmark*
- GIADA ROSSI • *Laboratory of Peptide and Protein Chemistry and Biology, Division of Pharmaceutical Sciences and Nutraceutic, Department of NeuroFarBa, University of Florence, Sesto Fiorentino, Italy*
- PAOLO ROVERO • *Laboratory of Peptide and Protein Chemistry and Biology, Division of Pharmaceutical Sciences and Nutraceutic, Department of NeuroFarBa, University of Florence, Sesto Fiorentino, Italy*
- BRIAN H. SANTICH • *Gerstner Sloan Kettering Graduate School of Biomedical Sciences, Memorial Sloan-Kettering Cancer Center, New York, NY, USA*
- ANDREA SANTONI • *Biochemistry and Molecular Biology Unit, Department of Life Sciences, University of Parma, Parma, Italy*
- ANNETTE SCHIOLBORG • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- NICOLE SCHNEIDERHAN-MARRA • *Natural and Medical Sciences Institute at the University of Tuebingen, Reutlingen, Germany*
- YASUO SHINOHARA • *Institute for Genome Research and Faculty of Pharmaceutical Sciences, University of Tokushima, Tokushima, Japan*
- HARINDER SINGH • *Bioinformatics Centre, CSIR-Institute of Microbial Technology, Chandigarh, India*
- OSCAR C. SOLÉ • *Institut de Biotecnologia i de Biomedicina (IBB), Universitat Autònoma de Barcelona (UAB), Barcelona, Spain*
- GLORIA SPAGNOLI • *Biochemistry and Molecular Biology Unit, Department of Life Sciences, University of Parma, Parma, Italy*
- MARKUS F. TEMPLIN • *Natural and Medical Sciences Institute at the University of Tuebingen, Reutlingen, Germany*
- MASSIMO TOMMASINO • *Infections and Cancer Biology Group, International Agency for Research on Cancer-World Health Organization, Lyon, France*
- NICOLE HARTWIG TRIER • *Department of Autoimmunology and Biomarkers, Statens Serum Institut, Copenhagen, Denmark*
- DANIELE VIARISIO • *German Cancer Research Center, Heidelberg, Germany*
- RYOHEI YAMAGOSHI • *Institute for Genome Research and Faculty of Pharmaceutical Sciences, University of Tokushima, Tokushima, Japan*
- ATSUSHI YAMAMOTO • *Faculty of Pharmaceutical Science, Suzuka University of Medical Science, Suzuka, Japan*
- TAKENORI YAMAMOTO • *Institute for Genome Research and Faculty of Pharmaceutical Sciences, University of Tokushima, Tokushima, Japan*

Chapter 1

Peptide Antibodies: Past, Present, and Future

Gunnar Houen

Abstract

Peptide antibodies recognize epitopes with amino acid residues adjacent in sequence (“linear” epitopes). Such antibodies can be made to virtually any sequence and have been immensely important in all areas of molecular biology and diagnostics due to their versatility and to the rapid growth in protein sequence information. Today, peptide antibodies can be routinely and rapidly made to large numbers of peptides, including peptides with posttranslationally modified residues, and are used for immunoblotting, immunocytochemistry, immunohistochemistry, and immunoassays. In the future, peptide antibodies will continue to be immensely important for molecular biology, TCR- and MHC-like peptide antibodies may be produced routinely, peptide antibodies with predetermined conformational specificities may be designed, and peptide-based vaccines may become part of vaccination programs.

Key words Peptides, Antibodies, Epitopes, Three-dimensional, Linear, Continuous, Contact residues, Recombinant, Single chain, TCR, MHC

1 The Development of Peptide Antibodies

Early work on protein structure and epitopes for antibodies (Abs) revealed that most epitopes were three dimensional (*see* **Notes 1 and 2**) and that a small percentage of Abs reacted with linear (continuous epitopes) [1–7]. Peptide Abs (Fig. 1) were described in 1980 and the use of synthetic peptides (coupled to a carrier protein) to induce specific Abs was developed in the following decades together with methods for epitope mapping and a general understanding of immunogenicity and antigenicity (Tables 1 and 2) (*see* **Notes 3–5**). This development was facilitated by the introduction of solid-phase peptide synthesis [41–43], the understanding of immunological T cell help for efficient stimulation of B cells to produce Abs [44–48], and the rapid growth in DNA and protein sequence information (Table 2).



Fig. 1 Examples of peptide epitopes in ovalbumin [52]. Three different linear/continuous epitopes for monoclonal epitopes are marked in *blue*, *red*, and *green* respectively

2 Current Status of Peptide Antibodies

Currently, peptide synthesis, conjugation, and immunization protocols have been optimized and the applications of peptide Abs have expanded to include a variety of immunoassays (e.g., sandwich assays), immunoprecipitation, immunoblotting, immunocytochemistry, and immunohistochemistry (Table 1). Moreover, posttranslational modification-specific Abs (e.g., phosphorylation and citrullination), cleavage site-specific Abs (e.g., amyloid beta 1–40/1–42), tag-specific Abs (e.g., hexa-histidine, FLAG, myc), and conformation-dependent Abs (Tables 1 and 2, [49–51]) are available for the different applications. Methods for epitope prediction have been refined but must always be verified by experimental results and compared with available structural data (*see Note 6*).

Table 1
History, status, and future developments of peptide antibodies

<i>A. History of peptide antibodies (selected publications)</i>		
Target ^a	Immunogen (residues (n))	References
MMLV putative protein	C-terminal pentadecapeptide	[8]
SV40 large T	N-terminal heptapeptide, C-terminal undecapeptide	[9]
HBV sAg	Several peptides (5–34)	[10]
FMDV VP1	Several peptides (15–40)	[11]
FMDV VP1	Hexadecapeptide	[12]
TCR	Branched lysine constructs	[13, 14]
RB	C-terminal decapeptide synthesised on carrier protein	[15]
CSFV E2	Dendrimeric peptide construct	[16]
<i>B. Current applications of peptide antibodies (i.e., methods used for detection of proteins and studies of protein modification and processing)</i>		
Application	References	
Immunoblotting	[17, 18]	
Immunoassays (direct, sandwich, etc.)	[19, 20]	
Immunocytochemistry and histochemistry	[21, 22]	
Flow cytometry	[23, 24]	
Immunoprecipitation	[25, 26]	
<i>C. Future developments of peptide antibodies</i>		
Application	References	
TCR-like Abs	[27, 28]	
Therapeutic peptide Abs/vaccines	[29, 30]	
Predesigned, conformation-specific peptide Abs	[31, 32]	
MHC-like Abs	?	

^aCSFV Classical swine fever virus, FMDV Foot and mouth disease virus, HBV Hepatitis B virus, MMLV Moloney murine leukemia virus, RB Retinoblastoma protein, SV Simian virus, TCR T cell receptor

Table 2
Peptide antibody reviews and resources

<i>A. Review and handbooks</i>	
Subject	References
Peptide Abs	[33]
Peptide vaccines	[34]
FMDV vaccines and peptide Abs	[35]
Peptide Ab immunoassays	[36]
Peptide Ab laboratory techniques	[37]
Peptide antigenicity and immunogenicity	[38]
Peptide-based autoimmune serology	[39]
Posttranslational modification-specific peptide Abs	[40]
<i>B. Websites</i>	
Epitope database	www.iedb.org
Epitope prediction	www.cbs.dtu.dk
Human protein atlas	www.proteinatlas.org
NCBI	www.ncbi.nlm.nih.gov
Uniprot/Swissprot	www.expasy.org
Protein database	www.pdb.org

3 Future Developments of Peptide Antibodies

Despite the achievements described above, the potential for peptide Abs has not been exhausted and many new uses have been recently established, are under development, or have been suggested (Table 1) including recombinant peptide Abs, single-chain peptide Abs, TCR-like Abs, predesigned conformation-dependent peptide Abs, and therapeutic peptide Abs. One of the original goals of peptide Abs, the development of clinical useful peptide vaccines, is getting closer to realization but still has to make it into clinical everyday use. MHC-like Abs, i.e., Abs, where the antibody-peptide complex mimics an MHC molecule, would be a desirable, although challenging goal.

4 Notes

1. All epitopes are three dimensional, but this term is here restricted to epitopes containing parts of a polypeptide chain not directly continuous in sequence. Thus, three-dimensional epitopes depend on a folded, native structure of the antigen (Ag). Three-dimensional epitopes may also be denoted “composite” epitopes.
2. Peptide epitopes are usually denoted “linear” or “continuous” epitopes but may also be denoted “simple” epitopes and are smaller than 20 residues continuous in sequence. The difference between peptides and polypeptides is not well defined but lies somewhere between 20 and 30 residues. All proteins are polypeptides, but this term is usually confined to polypeptides larger than 100 residues.
3. Epitope mapping: Mapping of amino acid residues with direct influence on Ab binding (e.g., by peptide scanning, X-ray crystallography, or NMR spectroscopy). Epitope residues may be contact residues or structural (conformational residues) or may contribute through backbone amide bonds.
4. Peptides may be antigenic (i.e., react with Ab (defined by Kd)) but not immunogenic (i.e., incapable of inducing an immune response (i.e., specific Abs and/or T cells)) [38]. The Ab response is quantified by the titers of a serum (defined by midpoint or endpoint titration) and by the average antigenicity of the Abs (Kd).
5. Contact residues: Amino acid residues directly interacting with the epitope or paratope (site on Ab interacting with epitope) as determined by X-ray crystallography and/or NMR spectroscopy of Ag-Ab complexes. The contact may take place between side chains or through backbone amide bonds.
6. See relevant chapters in this volume or see [53–55] for recent reviews.

References

1. Sela M, Schechter B, Schechter I, Borek F (1967) Antibodies to sequential and conformational determinants. Cold Spring Harbor Symp Quant Biol 32:537–545
2. Amit AG, Mariuzza RA, Phillips SE, Poljak RJ (1986) Three-dimensional structure of an antigen-antibody complex at 2.8 Å resolution. Science 233:747–753
3. Colman PM, Laver WG, Varghese JN, Baker AT, Tulloch PA, Air GM, Webster RG (1987) Three-dimensional structure of a complex of antibody with influenza virus neuraminidase. Nature 326:358–363
4. Sheriff S, Silverton EW, Padlan EA, Cohen GH, Smith-Gill SJ, Finzel BC, Davies DR (1987) Three-dimensional structure of an antibody-antigen complex. Proc Natl Acad Sci U S A 84:8075–8079
5. Mariuzza RA, Phillips SE, Poljak RJ (1987) The structural basis of antigen-antibody recognition. Annu Rev Biophys Biophys Chem 16:139–159
6. Colman PM, Tulip WR, Varghese JN, Tulloch PA, Baker AT, Laver WG, Air GM, Webster RG (1989) Three-dimensional structures of influenza virus neuraminidase-antibody complexes. Philos Trans R Soc Lond B Biol Sci 323:511–518
7. Scherf T, Hiller R, Naider F, Levitt M, Anglister J (1992) Induced peptide conformations in different antibody complexes: molecular modeling of the three-dimensional structure of peptide-antibody complexes using NMR-derived distance restraints. Biochemistry 31:6884–6897
8. Sutcliffe JG, Shinnick TM, Green N, Liu FT, Niman HL, Lerner RA (1980) Chemical synthesis of a polypeptide predicted from nucleotide sequence allows detection of a new retroviral gene product. Nature 287:801–805
9. Walter G, Scheidtmann KH, Carbone A, Laudano AP, Doolittle RF (1980) Antibodies specific for the carboxy- and amino-terminal regions of simian virus 40 large tumor antigen. Proc Natl Acad Sci U S A 77:5197–5200
10. Lerner RA, Green N, Alexander H, Liu FT, Sutcliffe JG, Shinnick TM (1981) Chemically synthesized peptides predicted from the nucleotide sequence of the hepatitis B virus genome elicit antibodies reactive with the native envelope protein of Dane particles. Proc Natl Acad Sci U S A 78:3403–3407
11. Bittle JL, Houghten RA, Alexander H, Shinnick TM, Sutcliffe JG, Lerner RA, Rowlands DJ, Brown F (1982) Protection against foot-and-mouth disease by immunization with a chemically synthesized peptide predicted from the viral nucleotide sequence. Nature 298:30–33
12. Pfaff E, Mussgay M, Böhm HO, Schulz GE, Schaller H (1982) Antibodies against a preselected peptide recognize and neutralize foot and mouth disease virus. EMBO J 1:869–874
13. Posnett DN, McGrath H, Tam JP (1988) A novel method for producing anti-peptide antibodies. Production of site-specific antibodies to the T cell antigen receptor beta-chain. J Biol Chem 263:1719–1725
14. Tam JP (1988) Synthetic peptide vaccine design: synthesis and properties of a high-density multiple antigenic peptide system. Proc Natl Acad Sci U S A 85:5409–5413
15. Hansen PR, Holm A, Houen G (1993) Solid-phase peptide synthesis on proteins. Int J Pept Protein Res 41:237–245
16. Li GX, Zhou YJ, Yu H, Li L, Wang YX, Tong W, Hou JW, Xu YZ, Zhu JP, Xu AT, Tong GZ (2012) A novel dendrimeric peptide induces high level neutralizing antibodies against classical swine fever virus in rabbits. Vet Microbiol 156:200–204
17. Petrasovits LA (2014) Protein blotting protocol for beginners. Methods Mol Biol 1099:189–199
18. Kurien BT, Dorri Y, Dillon S, Dsouza A, Scofield RH (2011) An overview of Western blotting for determining antibody specificities for immunohistochemistry. Methods Mol Biol 717:55–67
19. Wheeler MJ (2013) Immunoassay techniques. Methods Mol Biol 1065:7–25
20. Wild D (ed) (2013) The immunoassay handbook. Elsevier, Oxford
21. Brooks SA (2012) Basic immunocytochemistry for light microscopy. Methods Mol Biol 878:1–30
22. Ramos-Vara JA (2011) Principles and methods of immunohistochemistry. Methods Mol Biol 691:83–96
23. Davies D (2012) Cell separations by flow cytometry. Methods Mol Biol 878:185–199
24. Givan AL (2011) Flow cytometry: an introduction. Methods Mol Biol 699:1–29
25. Isono E, Schwechheimer C (2010) Co-immunoprecipitation and protein blots. Methods Mol Biol 655:377–387
26. Uljon SN, Mazzarelli L, Chait BT, Wang R (2000) Analysis of proteins and peptides directly from biological fluids by immunoprecipitation/mass spectrometry. Methods Mol Biol 146:439–452

27. Dahan R, Reiter Y (2012) T-cell-receptor-like antibodies—generation, function and applications. *Expert Rev Mol Med*. doi:[10.1017/erm.2012.2](https://doi.org/10.1017/erm.2012.2)
28. Neumann F, Sturm C, Hülsmeier M, Dauth N, Guillaume P, Luescher IF, Pfreundschuh M, Held G (2009) Fab antibodies capable of blocking T cells by competitive binding have the identical specificity but a higher affinity to the MHC-peptide-complex than the T cell receptor. *Immunol Lett* 125:86–92
29. Naz RK, Dabir P (2007) Peptide vaccines against cancer, infectious diseases, and conception. *Front Biosci* 12:1833–1844
30. Yamada A, Sasada T, Noguchi M, Itoh K (2013) Next-generation peptide vaccines for advanced cancer. *Cancer Sci* 104:15–21
31. Paduch M, Koide A, Uysal S, Rizk SS, Koide S, Kossiakoff AA (2013) Generating conformation-specific synthetic antibodies to trap proteins in selected functional states. *Methods* 60:3–14
32. Lu SM, Hodges RS (2002) A de novo designed template for generating conformation-specific antibodies that recognize alpha-helices in proteins. *J Biol Chem* 277:23515–23524
33. Sutcliffe JG, Shinnick TM, Green N, Lerner RA (1983) Antibodies that react with predetermined sites on proteins. *Science* 219:660–666
34. Shinnick TM, Sutcliffe JG, Green N, Lerner RA (1983) Synthetic peptide immunogens as vaccines. *Annu Rev Microbiol* 37:425–446
35. Brown F (1988) Use of peptides for immunization against foot-and-mouth disease. *Vaccine* 6:180–182
36. Van Regenmortel MH (1993) Synthetic peptides versus natural antigens in immunoassays. *Ann Biol Clin (Paris)* 51:39–41
37. Van Regenmortel MH, Briand JP, Muller S, Plau S (Eds) (1988) Synthetic polypeptides as antigens. *Laboratory techniques in biochemistry and molecular biology* vol 19. Elsevier: Amsterdam
38. Van Regenmortel MH (2001) Antigenicity and immunogenicity of synthetic peptides. *Biologicals* 29:209–213
39. Fournel S, Muller S (2003) Synthetic peptides in the diagnosis of systemic autoimmune diseases. *Curr Protein Pept Sci* 4:261–274
40. Papini AM (2009) The use of post-translationally modified peptides for detection of biomarkers of immune-mediated diseases. *J Pept Sci* 15:621–628
41. Merrifield RB (1963) Solid phase peptide synthesis. I The synthesis of a tetrapeptide. *J Am Chem Soc* 85:2149–2154
42. Merrifield RB (1969) Solid-phase peptide synthesis. *Adv Enzymol Relat Areas Mol Biol* 32:221–296
43. Atherton E, Sheppard RC (1989) *Solid Phase peptide synthesis: a practical approach*. IRL Press, Oxford, England. ISBN 0-19-963067-4
44. Braciale TJ, Morrison LA, Sweetser MT, Sambrook J, Gething MJ, Braciale VL (1987) Antigen presentation pathways to class I and class II MHC-restricted T lymphocytes. *Immunol Rev* 98:95–114
45. Parker DC (1993) T cell-dependent B cell activation. *Annu Rev Immunol* 11:331–360
46. Fairchild PJ (1998) Presentation of antigenic peptides by products of the major histocompatibility complex. *J Pept Sci* 4:182–194
47. Appella E, Padlan EA, Hunt DF (1995) Analysis of the structure of naturally processed peptides bound by class I and class II major histocompatibility complex molecules. *EXS* 73:105–119
48. Maffei A, Harris PE (1998) Peptides bound to major histocompatibility complex molecules. *Peptides* 19:179–198
49. Blaydes JP, Vojtesek B, Bloomberg GB, Hupp TR (2000) The development and use of phospho-specific antibodies to study protein phosphorylation. *Methods Mol Biol* 99:177–189
50. Miller DL, Potempska A, Wegiel J, Mehta PD (2011) High-affinity rabbit monoclonal antibodies specific for amyloid peptides amyloid- β 40 and amyloid- β 42. *J Alzheimers Dis* 23:293–305
51. Terpe K (2003) Overview of tag protein fusions: from molecular and biochemical fundamentals to commercial systems. *Appl Microbiol Biotechnol* 60:523–533
52. Holm BE, Bergmann AC, Hansen PR, Koch C, Houen G, Trier NH (2014) Antibodies with specificity for native and denatured forms of ovalbumin differ in reactivity between enzyme-linked immunosorbent assays. *APMIS*. doi:[10.1111/apm.12329](https://doi.org/10.1111/apm.12329)
53. Soria-Guerra RE, Nieto-Gomez R, Govea-Alonso DO, Rosales-Mendoza S (2014) An overview of bioinformatics tools for epitope prediction: implications on vaccine development. *J Biomed Inform* S1532-0464(14):00233. doi:[10.1016/j.jbi.2014.11.003](https://doi.org/10.1016/j.jbi.2014.11.003)
54. Ansari HR, Raghava GP (2013) In silico models for B-cell epitope recognition and signaling. *Methods Mol Biol* 993:129–138
55. Ponomarenko JV, van Regenmortel MHV (2009) B-cell epitope prediction. In: Bourne PE, Gu J (eds) *Structural bioinformatics*. Wiley, New York, NY, pp 849–879

Chapter 2

The Structure of Natural and Recombinant Antibodies

Hui Ma and Richard O’Kennedy

Abstract

Immunoglobulins (Ig) isotypes A, D, E, G, and M are glycoproteins which are mainly composed of a “Y”-shaped Ig monomer (~150 kDa), consisting of two light and two heavy chains. Both light and heavy chains contain variable (N-terminal) and constant regions (C-terminal). Each light chain consists of one variable domain and one constant domain, whereas each heavy chain has one variable domain and three constant domains. However, heavy-chain antibodies consisting of only heavy chains and lacking the light chains are found in camelids and cartilaginous fishes. Unlike other immunoglobulins, the heavy chain of avian antibody IgY (~180 kDa) consists of four constant domains. The single-chain variable fragment (scFv; ~25 kDa) of an antibody contains variable regions of antibody heavy and light chains. The fragment antigen-binding (Fab; ~50 kDa) region has the full antibody light chain but the heavy chain is composed of a variable region and one constant domain.

Key words IgG, IgY, ScFv, Fab

1 Structure of Immunoglobulins

Antibodies, also known as immunoglobulins (Igs), enable antigen recognition in the serum. They are produced by B-cell-derived plasma cells. Antibodies are mainly located in blood, spleen, bone marrow, egg yolk for birds, as well as interstitial fluids and exocrine secretions. Antibodies can be effectively used by the immune system to identify, kill, or neutralize invading bacteria, parasites, toxins, and viruses and to destroy other foreign compounds [1].

Mammalian immunoglobulins are classified into five isotypes, namely IgM, IgD, IgG, IgE, and IgA. The synonymous “Y” shape associated with a basic immunoglobulin unit (Ig) monomer (or subunit) (~150 kDa) consists of two light and two heavy chains, which are connected by disulfide bonds (*see* Fig. 1a) [2]. Each light chain has two regions composed of one variable region (V_L) and one constant region (C_L), whereas each heavy chain contains one variable domain (V_H) and three constant domains (C_{H1-3}). All of the antibodies perform specific binding to defined antigens through

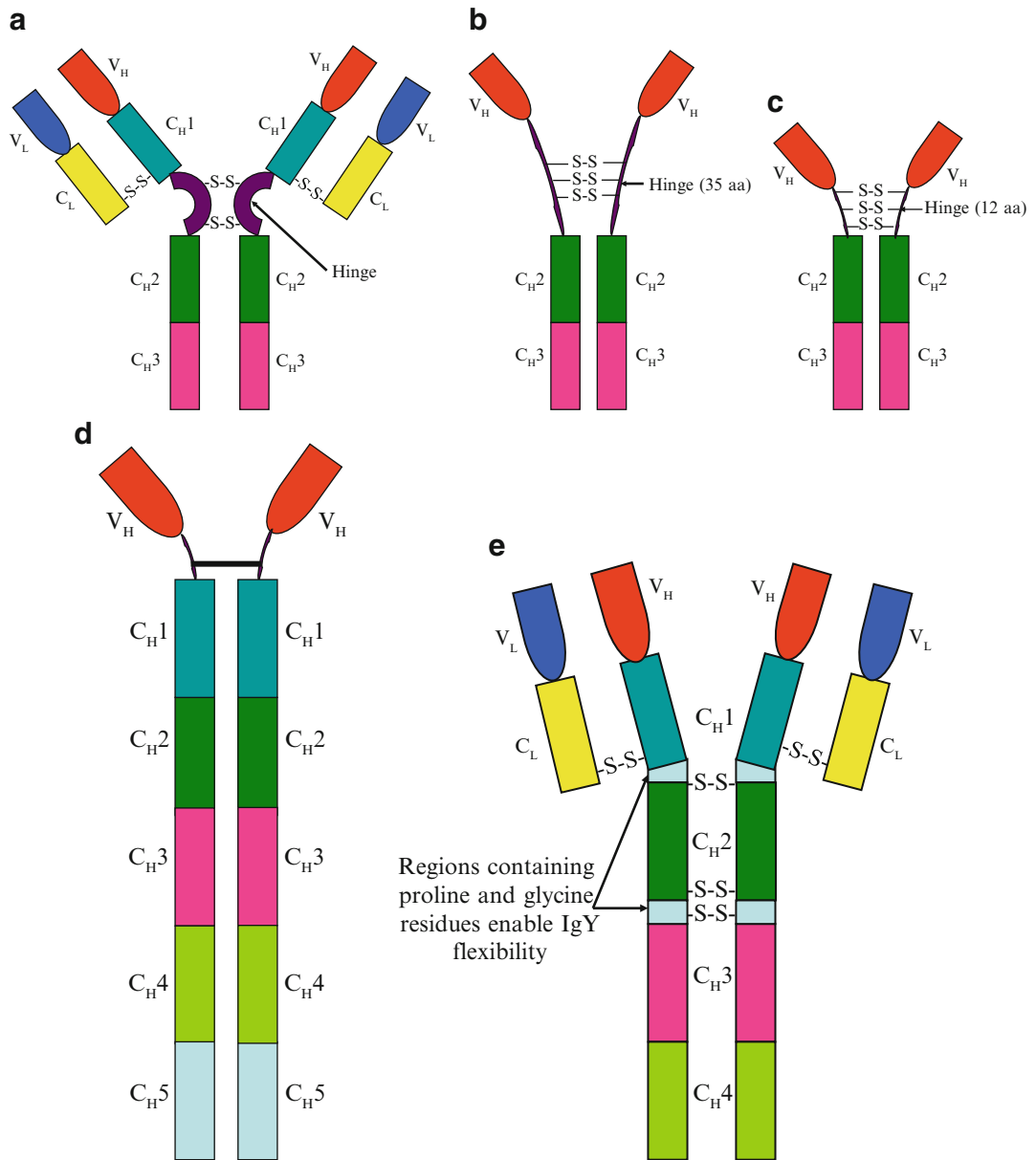


Fig. 1 Structure of basic immunoglobulins. **(a)** Shows structure of a basic immunoglobulin monomer. **(b–d)** Show structure of Camelidae IgG2 **(b)**, Camelidae IgG3 **(c)**, and avian IgY antibody **(d)**. NH₂ = amino group; COOH = carboxylic acid group; V_H = variable region of antibody heavy chain; V_L = variable region of antibody light chain; C_L = constant region of antibody light chain; C_H1,2,3,4,5 = constant domain one, two, three, four, and five of antibody heavy chain; S–S = disulfide bond; and aa = amino acid

the variable regions. Almost all five isotypes, IgA (dimer), IgD (monomers), IgE (monomer), IgG (monomer), and IgM (pentamer), are composed of the same basic immunoglobulin unit with some modifications [3]. However, all the members of *Camelidae*

family have a heavy-chain antibody, which consists of two heavy chains (one variable region and two constant regions per chain), and lacks the two light chains. There are three subclasses of IgG in camels and llamas, i.e., the conventional IgG1 (~160 kDa, with full-length light and heavy chains), IgG2 (~92 kDa; with a long hinge; *see* Fig. 1b) and IgG3 (~86 kDa; with a short hinge; *see* Fig. 1c), which lack both light chains and C_H1 [4]. Heavy-chain antibodies are also found in cartilaginous fish. They are called immunoglobulin new antigen receptors (IgNARs; ~175 kDa; *see* Fig. 1d). An IgNAR contains only two heavy chains and each chain has one variable region and five constant regions (C_H1–5) [5].

Avian immunoglobulins are of three principal classes, IgA, IgM, and IgY (the 180 kDa homologue of mammal IgG). Unlike the heavy chain of mammalian immunoglobulins, the heavy chain of IgY consists of four constant Ig domains (*see* Fig. 1e). Female chickens (hens) are favored for producing large amounts of IgY, as this can be harvested from egg yolk. The process is more convenient than isolation of antibodies from blood and other organs (e.g., spleen and bone marrow). IgG contains regions between C_H1 and C_H2, while in IgY two regions (one between C_H1 and C_H2 and the other between C_H2 and C_H3), containing proline and glycine residues, enable limited flexibility [6].

2 Structure of Recombinant Antibodies

A recombinant antibody does not exist naturally but is assembled from DNA by combining antibody heavy-chain and light-chain gene sequences. The single-chain variable fragment (scFv) and fragment antigen-binding (Fab) region are the most popular recombinant antibody formats used due to their short generation time and high antigen affinity and structural stability [7].

A scFv consists of variable (binding) regions of the antibody heavy (V_H) and light (V_L) chains, with a flexible linker [e.g., (GGGS)₃ linker] joining the terminal ends of either the V_H to V_L (or V_L to V_H) (*see* Fig. 2a). It is popular and effective to use a disease-specific scFv for targeted therapy through fusing to therapeutic proteins or genes.

The Fab fragment, which is double the size of the scFv, is formed by one variable and one constant domain of both light and heavy chains, and is linked by a disulfide bridge (*see* Fig. 2b) [8]. There are many medicines, which have been derived from Fabs that are now approved by the Food and Drug Administration (FDA).

Moreover, various kinds of scFv and/or Fab-derived antibodies have been generated for clinical applications. Bivalent or trivalent scFvs consist of two or three scFvs linked with a short amino acid

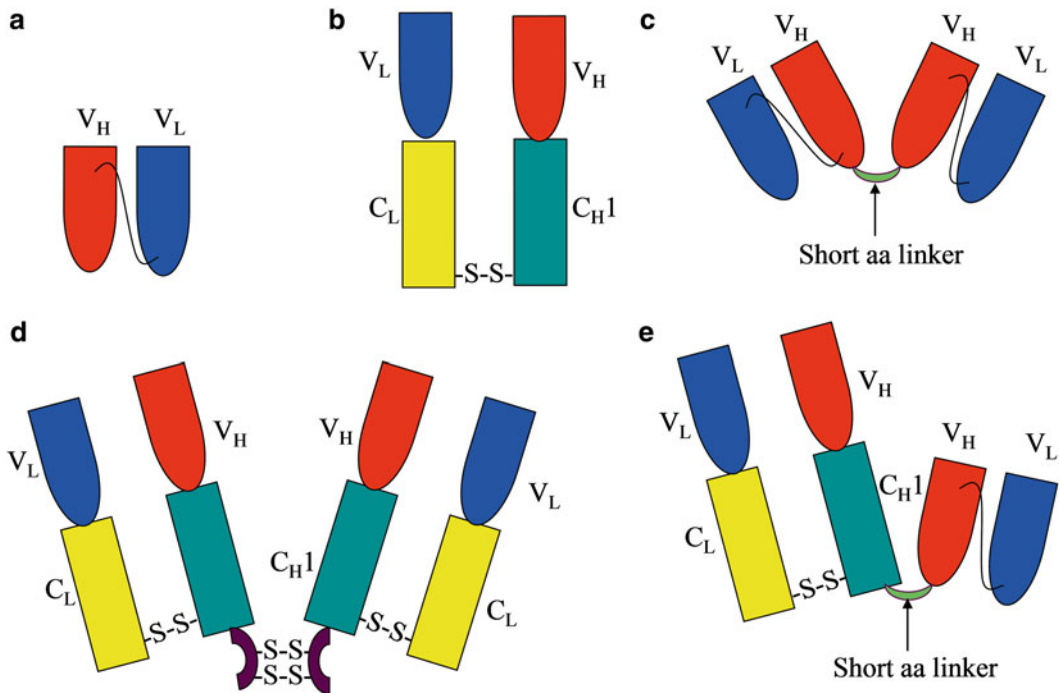


Fig. 2 Structures of a scFv (a), a Fab fragment (b), a (scFv)₂ (c), a F(ab')₂ (d), and a Fab-scFv (e). V_H = variable region of antibody heavy chain; V_L = variable region of antibody light chain; C_L = constant region of antibody light chain; C_{H1} = constant domain one of antibody heavy chain; S-S = disulfide bond; and aa = amino acid

linker [9] (see Fig. 2c). A F(ab')₂ fragment contains two Fab fragments linked by disulfide bonds. It can be obtained by cleaving whole immunoglobulins using the enzyme pepsin below the hinge region (see Fig. 2d). The Fab-scFv fusion antibody is formed by a Fab and an scFv via a short amino acid linker (see Fig. 2e).

Acknowledgement

This work is supported by Science Foundation Ireland under CSET Grant No. 05/CE3/B754 and 10/CE/B1821.

References

1. Meffre E, Casellas R, Nussenzweig MC (2000) Antibody regulation of B cell development. *Nat Immunol* 1:379–85
2. Chailyan A, Tramontano A, Marcatili P (2012) A database of immunoglobulins with integrated tools: DIGIT. *Nucleic Acids Res* 40:1230–4
3. Schroeder HW Jr, Cavacini L (2010) Structure and function of immunoglobulins. *J Allergy Clin Immunol* 125:S41–52
4. Shaker GH (2010) Evaluation of anti-diphtheria toxin nanobodies. *Nanotechnol Sci Appl* 3: 29–35